

Lysogeny and prophage induction in coastal and offshore bacterial communities

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ABSTRACT: The influence of solar radiation and hydrogen peroxide on induction of lysogens, and the resulting effect on bacteriophage production and bacterial mortality was investigated for coastal and oceanic marine bacterial communities at 6 stations in the western Gulf of Mexico. The percentage of lysogenic cells induced by mitomycin C was also determined. Solar radiation and hydrogen peroxide were not as effective as mitomycin C at inducing phage production. The burst size of cells induced by mitomycin C was estimated by transmission electron microscopy, assuming that cells completely filled with viral particles were on the verge of bursting. The smallest estimates of burst size were associated with oligotrophic oceanic stations and ranged from 15 to 28 viruses produced per lytic event, while in more productive coastal waters the estimated burst sizes ranged from 33 to 64. The mitomycin C-induced phage production and burst size were used to estimate the number of lysogenic bacterial cells. On average, the percentage of inducible lysogens was higher at offshore (1.5 to 11.4%) than at coastal (0.8 to 2.2%) stations. However, with the exception of 1 station, less than 5% of the bacteria could be induced to produce phage, suggesting that lysogens only occasionally comprised a significant component of these bacterial communities. The proportion of lysogens that could be induced by sunlight, relative to those that could be induced by mitomycin C, was lower at oceanic than coastal stations. This implies that prophages in optically transparent offshore waters were more resistant to induction by solar radiation, or that most lysogens that could be triggered by sunlight were already induced. Based on a steady-state model, induction of lysogenic bacteria by solar radiation or hydrogen peroxide could result in between 0 and 3.5% or 0.9 and 3.4% of the total bacterial mortality, respectively. Our results imply that solar radiation and hydrogen peroxide induced lysogenic phage production were not an important source of phage production or bacterial mortality in offshore or coastal waters of the western Gulf of Mexico.

KEY WORDS: Lysogeny · Prophage · Marine viruses · Bacteriophage · UV radiation

INTRODUCTION

Viruses are abundant and dynamic components of aquatic microbial food webs. They cause mortality of microorganisms, which can affect community composition and cycling of organic matter (Fuhrman & Suttle 1993, Thingstad et al. 1993). The role of viruses in microbial food webs depends on their mode of replication. There are 2 major pathways of phage replication (Luria & Darnell 1967). In the lytic cycle, the phage genome replicates immediately after infection and

newly formed virus particles are released during lysis of the host cell. In the lysogenic cycle, the genome of a temperate phage integrates into the host genome where it is carried in a dormant form (prophage) until the lytic cycle is induced by factors such as exposure to UV. Thus, lytic phage production depends on encounter rates between phages and host cells, whereas, lysogenic phage production depends on the number of lysogenized cells in the bacterial community and on the presence of an inducing agent.

Hypotheses have been put forward to explain the advantages of lysogeny (e.g. Steward & Levin 1984). For example, since lysogenized cells are immune

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against infection by phages that are closely related to the prophage, they may have a competitive advantage over non-lysogenized cells. Lysogeny is also a potential survival strategy to maintain phage populations when host abundance is too low for populations to be maintained by lytic infection. As well, temperate phages can add metabolic properties to bacterial strains, such as antibiotic resistance and production of virulence factors (Waldor & Mekalanos 1996). In some temperate phages, low nutrient concentrations and a high ratio of viruses to host cells seems to favour the establishment of lysogeny; however, evidence for this is sparse for marine microbial communities (Wilson & Mann 1997).

It has been suggested that lysogenic bacteria predominate in planktonic systems (Thingstad et al. 1993, Bratbak et al. 1994). Moebus (1983) found that about 10% of the phages isolated from marine systems were temperate, while Jiang & Paul (1994) were able to induce 43% of marine bacterial isolates to produce phage. A lower percentage of isolates from eutrophic than coral-reef and reef-lagoon environments could be induced. In other studies, Paul and co-workers detected lysogeny in ca 50% of marine samples (Jiang & Paul 1996, Cochran & Paul 1998, Cochran et al. 1998). In contrast, Weinbauer & Suttle (1996) found lysogeny in all samples, but estimated that <5% of bacteria in coastal waters were lysogenized. Wilcox & Fuhrman (1994) found no evidence for lysogenic bacteriophage production in natural bacterial communities from coastal seawater, while in oligotrophic Lake Superior, Tapper & Hicks (1998) estimated 0.1 to 7.4% of bacteria in natural communities were lysogenized. Current evidence suggests that lysogenic cells typically comprise a small proportion of bacteria in natural communities, but occasionally can be abundant.

Mitomycin C and UV-C are commonly used to induce lysogens (Ackermann & DuBow 1987). Based on studies in the phage lambda (Gottesman & Oppenheim 1994), mitomycin C and UV-C damage the DNA, which activates the SOS repair system, including the enzyme RecA. The RecA protein cleaves a repressor and induces the lytic cycle.

The factors that induce lysogenic phage production in nature are largely unknown. Solar radiation causes DNA damage in bacterioplankton (Aas et al. 1996, Jeffrey et al. 1996) and may be an inducing agent. Also, hydrogen peroxide can induce lysogenized cells by damaging the host DNA (Lwoff 1952, Ackermann & DuBow 1987, Imlay & Linn 1987). Because this molecule occurs in seawater (Van Baalen & Marler 1966, Zika et al. 1985), it might be an inducing agent in nature. Increases in temperature and vigorous cell growth are also known to induce the lytic cycle (Gottesman & Oppenheim 1994), and polyaromatic

hydrocarbons can induce lysogens in natural bacterial communities (Jiang & Paul 1996).

Potentially, increased UV-B in the upper layer of the ocean because of atmospheric ozone depletion (Jones & Shanklin 1995) would result in higher induction rates of lysogens. In turn, this may affect species composition and the way viruses influence cycling of nutrients and trace gases. Elevated UV-B would also potentially increase induction rates via hydrogen-peroxide production from photodegradation of organic matter (Petasne & Zika 1987, Johnson et al. 1989). Our study estimated the percentage of lysogens that could be induced by mitomycin C, solar radiation and hydrogen peroxide along a transect from oceanic to neritic waters in the western Gulf of Mexico. Sunlight effects on virus production and bacterial mortality were also investigated.

MATERIALS AND METHODS

Study sites and sampling. This study was done during cruises in the western Gulf of Mexico on June 20 to 28 (Stns A to F), and November 1, 1995 (Stn J, Fig. 1). Before sunrise, water for incubations was pumped from the surface (0.5 m depth) or collected from the deep chlorophyll maximum (DCM; 72 m at Stn E) using Niskin bottles mounted on a rosette equipped with a Sea-Bird CTD (model SBE plus) and fluorometer. The depths of the DCM and mixed layer were determined from multiple profiles of salinity, temperature and fluorescence. Duplicate samples for viral and bacterial abundances, and burst size were preserved with glutaraldehyde (2% final concentration) immediately after collection, and stored in the dark at 4°C. Samples for chlorophyll *a* (chl *a*) were collected onto 0.45 µm pore-size filters and frozen for subsequent analysis.

Incubations. Freshly collected seawater (4 l) was filtered through 1.2 µm nominal pore-size glass-fiber filters (147 mm; Gelman GC50), and the bacteria in the filtrate concentrated 10-fold using a 0.2 µm pore-size polycarbonate cartridge filter (Nuclepore). An aliquot (20 ml) of the bacterial concentrate was made up to 2000 ml with virus-free ultrafiltered (Amicon S10Y30 and S1Y30 cartridges; 30000 MW cut-off) seawater made immediately before the experiments as previously described (Suttle et al. 1991). This procedure decreased the concentration of viruses by about 100-fold and bacterial numbers by about 10-fold compared to the *in situ* abundances. Reducing the abundances of viruses and bacteria decreased contact rates between them, and thereby reduced the production of new viruses by the lytic cycle. Encounter rates between viruses and bacteria were calculated using the model of Murray & Jackson (1992). Assuming the diameter of

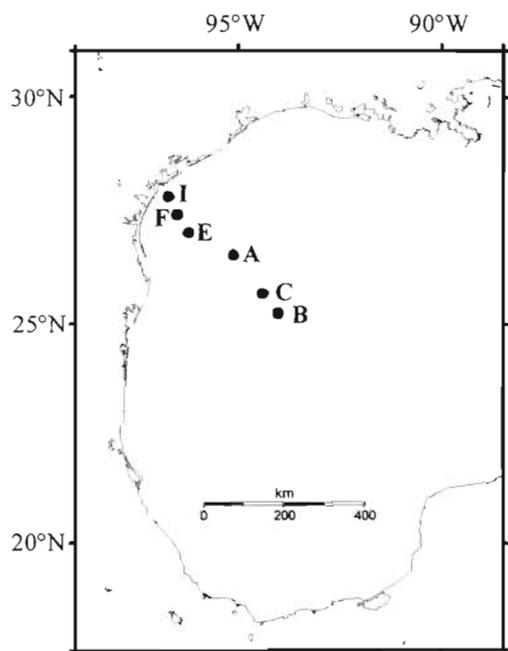


Fig. 1. Location of sampling sites in the western Gulf of Mexico

a typical bacterial cell to be $0.5 \mu\text{m}$, subsequent to dilution $<0.7\%$ of the bacterial community was contacted per hour by a virus. Therefore, the observed virus production was primarily the result of induction of lysogenized cells rather than lytic infection. Decreasing the viral abundance also allowed viral production to be estimated more accurately by reducing the background concentration of viruses. Furthermore, dilution of the samples with ultrafiltrate minimised the possibility that the experiments were compromised by nutrient limitation of the microbial community.

Aliquots (200 ml) of diluted samples were added to UV-transparent polyethylene bags (Fisher). For the chemical treatments, either mitomycin C (Sigma Chemical Co. No. M-0503; 1 mg ml^{-1} deionized-distilled water) or hydrogen peroxide (J.T. Baker Chemical No. 2186; 3% in deionized-distilled water) was added to the incubations at final concentrations of $1.0 \mu\text{g ml}^{-1}$ or 0.1 to $1.35 \mu\text{M}$, respectively. Samples were also exposed to solar radiation from ca 08:00 to 17:00 h, except for the sample from E_{DCM} , which was exposed from 12:00 to 17:00 h. Following exposure the samples were kept in the dark. Untreated dark incubations served as controls. Previous studies indicated that there was no difference in viral and bacterial numbers between incubations in the dark and incubations in which UV-B was excluded by Mylar film (Weinbauer & Suttle 1996). Treatments were performed in duplicate. The bags were placed in a flowing seawater bath to maintain *in situ* temperature. Subsamples for the enu-

meration of viruses and bacteria, and estimation of burst size were removed periodically and preserved with glutaraldehyde (2% final concentration). Untreated incubations were used as controls to check for lytic phage production.

Quantification of bacteria, viruses and chlorophyll *a*. Bacterial and viral abundances were determined within 19 d of sample collection. Aliquots of the samples were stained with 4'6-diamidino-2-phenylindole (DAPI; final concentration: $1 \mu\text{g ml}^{-1}$) and enumerated by epifluorescence microscopy (Suttle 1993) using the modifications of Weinbauer & Suttle (1997). Samples for bacteria and viruses were stained without DNase treatment for 30 min and filtered onto $0.02 \mu\text{m}$ pore-size Anodisc filters (Whatman) and enumerated using an Olympus BX40 epifluorescence microscope. Chl *a* was determined fluorometrically (Parsons et al. 1984).

Determination of burst size. Bacteria from up to 40 ml of sample were collected quantitatively onto Formvar-coated, 400-mesh electron microscope grids by centrifugation in a swinging-bucket rotor (SW 40; $70\,950 \times g$ for 20 min). The grids were stained for 30 s with 1% uranyl acetate and rinsed 3 times with deionized-distilled water. The time and speed of centrifugation avoided disruption of infected bacteria. Furthermore, phages within bacteria were easily distinguished as few viruses in solution were pelleted. Burst size was estimated by averaging the number of virus-like particles (VLPs) within a minimum of 15 cells that were completely filled with VLPs (Weinbauer & Peduzzi 1994).

RESULTS

Characterisation of sampling sites

Water temperatures ranged from 21.3 to 28.8°C and salinity from 29.3 to 36.6 ppt (Table 1). Chl *a* at the oceanic stations (A, C, D) ranged from 0.04 to $0.08 \mu\text{g l}^{-1}$, and at the neritic stations (E, F, I) from 0.21 to $0.38 \mu\text{g l}^{-1}$. At the oceanic stations bacterial abundances ranged from 0.4×10^6 to $0.6 \times 10^6 \text{ ml}^{-1}$, while viral abundances ranged from 0.2×10^6 to $0.3 \times 10^6 \text{ ml}^{-1}$. At the neritic stations bacteria and viruses were more abundant and ranged from 1.0×10^6 to 3.5×10^6 bacteria ml^{-1} and from 5.1×10^6 to 8.7×10^6 viruses ml^{-1} .

Bacterial dynamics

Bacterial abundance typically increased in the controls and sunlight-exposed incubations, but the response was more variable in incubations treated with

Table 1. Temperature, salinity, chlorophyll *a* concentrations, bacterial and viral abundances, and burst sizes at the various sampling sites at the different stations. Stn A: 26°44'N, 95°19'W; Stn B: 25°19'N, 94°07'W; Stn C: 25°41'N, 94°31'W; Stn E: 27°00'N, 96°14'W; Stn F: 27°32'N, 96°45'W; Stn I: 27°45'N, 96°51'W. DCM: deep chlorophyll maximum. Chl *a* concentrations are integrated over the mixed layer with the exception of Stn E_{DCM}. Values for bacteria, viruses and burst size are expressed as means \pm standard deviation of duplicate incubations

Stn	Temperature (°C)	Salinity (ppt)	Chl <i>a</i> ($\mu\text{g l}^{-1}$)	Bacteria (10^6 ml^{-1})	Viruses (10^6 ml^{-1})	Burst size (viruses cell $^{-1}$)
Offshore						
A	28.2	35.7	0.08	0.5 \pm 0.03	0.3 \pm 0.03	28 \pm 7.9
B	28.2	36.6	0.04	0.4 \pm 0.03	0.3 \pm 0.03	23 \pm 5.1
C	28.5	36.5	0.07	0.6 \pm 0.06	0.2 \pm 0.01	15 \pm 5.3
Coastal						
E	28.4	30.1	0.14	1.8 \pm 0.10	5.1 \pm 0.36	40 \pm 9.5
E _{DCM}	21.3	36.2	0.36	1.0 \pm 0.11	5.9 \pm 0.53	33 \pm 9.1
F	28.8	29.3	0.20	2.5 \pm 0.19	6.6 \pm 0.40	47 \pm 5.6
I	21.1	30.6	0.35	3.5 \pm 0.22	8.7 \pm 0.09	64 \pm 19.1

hydrogen peroxide. In contrast, abundance remained comparatively stable in incubations treated with mitomycin C (Figs. 2 & 3). The effect of solar radiation and hydrogen peroxide on bacterial production was estimated from the change in bacterial abundance between the beginning and end of each experiment. At the end of the experiments (about 24 h), abun-

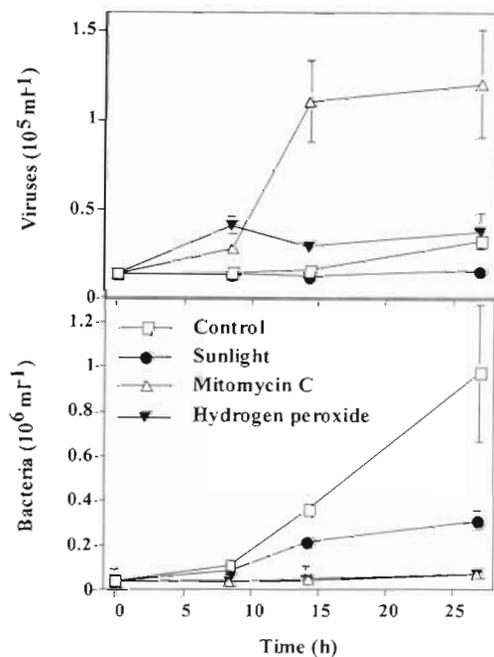


Fig. 2. Virus and bacterial abundances in experiments using seawater collected at an offshore station (Stn C) on June 28, 1995. When error bars are not shown, the range calculated from duplicate incubations was less than the width of the symbols

dances in the solar-radiation-treated incubations were 29 to 84 % (average: 55 \pm 25%, $n = 7$) of those in the dark controls. Percentages did not differ significantly between neritic and oceanic stations (2-tailed *t*-test; $p > 0.4$). The inhibition of bacterial growth was not correlated significantly to PAR ($r = 0.430$; $p > 0.3$). Hydrogen peroxide had an even stronger effect on bacterial production. Bacterial abundances in hydrogen-peroxide treatments were between 3 and 71 % (average: 25 \pm 25%, $n = 7$) of those in control incubations. The inhibition of bacterial growth due to hydrogen peroxide was not correlated significantly ($r = 0.284$; $p > 0.5$) to the hydrogen-peroxide concentration added and did not differ significantly between coastal and offshore stations (2-tailed *t*-test; $p > 0.8$).

Viral dynamics

Typically, virus abundances remained comparatively stable in the controls during the first 7 to 17 h, but increased when treated with 1.0 $\mu\text{g ml}^{-1}$ mitomycin C (Figs. 2 & 3). The difference in viral abundance between mitomycin C-treated and untreated incubations

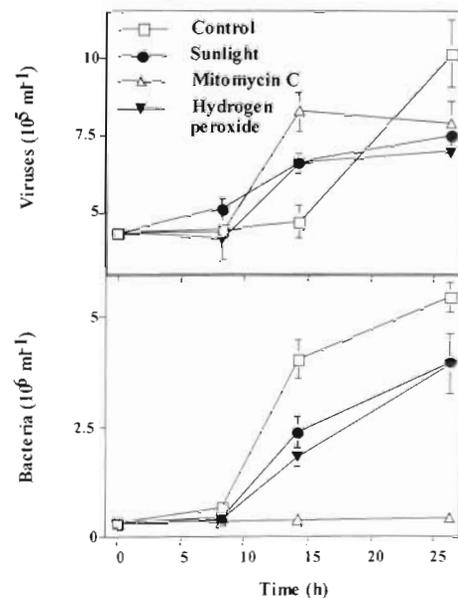


Fig. 3. Virus and bacterial abundances in experiments using seawater collected at a coastal station (Stn F) on November 1, 1995. When error bars are not shown, the range calculated from duplicate incubations was less than the width of the symbols

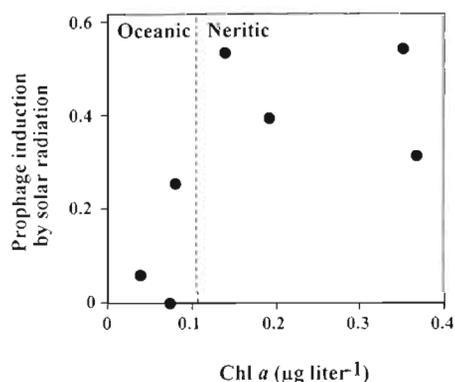


Fig. 4. Prophage induction as the result of exposure to solar radiation relative to that of mitomycin C. The effect of solar radiation on prophage induction was calculated using data from Table 2. It is expressed as the ratio of viruses produced in the sunlight-treated incubations to the viruses produced in the mitomycin C-treated incubations

during this period was assumed to have resulted from lysogenic phage production. After this initial period, viral abundances remained comparatively stable in the mitomycin C treatments, but increased in the control incubations of some experiments.

After 7 to 17 h of incubation, viral abundances were higher in sunlight-exposed samples than dark controls (except for Stn C); however, the abundances were always lower than in mitomycin C treatments (Figs. 2 & 3). This indicates that solar radiation induced a portion of the lysogenized bacteria. The ratio of sunlight-induced viral production to that induced by mitomycin

C ranged from 0 to 0.54 (average, 0.29 ± 0.21 ; $n = 7$) and was significantly (2-tailed *t*-test; $p < 0.05$) lower at oceanic (average, 0.12 ± 0.15 ; range, 0 to 0.25; $n = 3$) than neritic stations (average, 0.42 ± 0.12 ; range, 0.31 to 0.54; $n = 4$; Fig. 4).

Hydrogen-peroxide-induced phage production after 7 to 17 h was lower than that induced by mitomycin C, but higher than in dark controls (Figs. 1 & 2), indicating that hydrogen peroxide can induce lysogenic phage production. At Stns C, E and E_{DCM} the hydrogen-peroxide-induced virus production ranged from 3 to 6% of that induced by mitomycin C (calculated from Table 2). At Stn F, it was 15% of that induced by mitomycin C, and ranged from 48 to 53% at Stns A, B and I. Hydrogen-peroxide-induced phage production was not correlated significantly to the final concentration of hydrogen peroxide ($r = 0.313$; $p > 0.5$). At Stns B and C, hydrogen peroxide induced a greater number of viruses than treatment with solar radiation.

Burst size and percentage of lysogenic bacteria

Estimates of burst size in mitomycin C-treated incubations ranged from 20 to 45 (average: 33; $n = 7$; Table 2), similar to estimates for untreated natural bacterial communities (15 to 64, average 36, $n = 7$; Table 1). The difference in burst-size estimates between seawater samples and mitomycin C-treated incubations was not significant (2-tailed *t*-test; $p > 0.3$).

Seven to 17 h after the start of the experiments, viral abundances were higher in the mitomycin C-treated incubations than in the controls; the difference was attributed to phage production as a result of lysogen induction. The number of induced bacteria (N) was calculated as $N = (V_m - V_c)/B$, where V_m = viral abundance in mitomycin C treatments, V_c = viral abundance in control incubations and B = burst size. Burst sizes were determined in samples collected after 7 to 17 h from the mitomycin C incubations. The percentage of lysogenic cells in the starting bacterial communities was estimated to range from 1.5 to 11.4% at the oceanic stations and from 0.8 to 2.2% at the neritic stations (Table 2). Significantly higher percentages of lysogenized bacteria were found in oceanic than neritic waters during the June cruise, as well as when the data from the June cruise were combined with those collected during a previous study (Weinbauer & Suttle 1996) (2-tailed *t*-test; $p < 0.05$).

Table 2. Estimation of the percentage of lysogenic bacteria in the total bacterial community based on induction by mitomycin C. V_m : viral abundance (ml^{-1}) in the mitomycin C treatment; V_s : viral abundance (ml^{-1}) in incubations that were exposed to solar radiation from about 08:00 to 17:00 h; V_h : viral abundance (ml^{-1}) in the hydrogen peroxide incubations; V_c : viral abundance (ml^{-1}) in the control incubations. Values for viruses and bacteria are expressed as means \pm range calculated from duplicate incubations. Bacterial abundance is given at the onset of the experiment. Percentage of lysogenic bacteria calculated as $\{(V_m - V_c)/B/A\} \times 100$, where B = burst size and A = bacterial abundance

Stn	$V_m - V_c$	$V_s - V_c$	$V_h - V_c$	Burst size	Bacteria (10^5 ml^{-1})	% lysogenic bacteria
Offshore						
A	16700	4850	7980	22 ± 6.2	0.49 ± 0.025	1.5
B	10370	630	4960	20 ± 5.4	0.20 ± 0.163	2.6
C	94420	0	2640	25 ± 3.5	0.33 ± 0.053	11.4
Coastal						
E	61200	19130	3520	36 ± 5.9	2.20 ± 0.962	0.8
E_{DCM}	72900	38900	3610	29 ± 4.7	2.96 ± 0.830	0.8
F	65580	25980	9480	45 ± 11.2	1.83 ± 0.353	0.9
I	358080	194790	190370	51 ± 9.7	3.19 ± 0.229	2.2

DISCUSSION

This study provides empirical estimates of the proportion of lysogenic cells in marine bacterial communities, based on induction by mitomycin C. A higher proportion of cells were induced in oceanic than neritic waters. Solar radiation and hydrogen peroxide also induced lysogenic phage production, although they were not important mechanisms for phage production and bacterial mortality.

Response of bacteria to solar radiation and hydrogen peroxide treatment

Solar radiation inhibited bacterial production relative to dark incubations. Similar effects on bacterial abundance and production have been observed for other environments (e.g. Herndl et al. 1993, Jeffrey et al. 1996). The effect of solar radiation did not differ significantly between coastal and offshore stations. This is consistent with observations that DNA damage was similar in sunlight-exposed natural viral communities from neritic and oceanic stations (Weinbauer et al. 1999). When compared with solar radiation, hydrogen peroxide had an even more pronounced effect on bacterial production. Even at the lowest concentration added (0.1 μM), bacterial abundance was strongly reduced relative to dark incubations. Since hydrogen-peroxide concentrations up to 0.2 μM are found in the Gulf of Mexico (Van Baalen & Marler 1966, Zika et al. 1985), it might be important in controlling bacterial biomass and production.

Percentage of lysogenized bacteria

The data in this paper indicate that 0.8 to 11.4% (average, 2.9%; $n = 7$) of the natural bacterial community was induced to produce phage by the addition of mitomycin C (Table 2). Paul and co-workers (Jiang & Paul 1996, Cochran et al. 1998, Cochran & Paul 1998) estimated that mitomycin C-induced lysogens in ca 50 to 60% of the samples they examined. They inferred that the percentage of bacteria that were lysogenized varied from undetectable to 100%. In contrast, we detected low levels of lysogenic phage production in all samples. Our approach can detect induction by <0.1% of the bacterial community (Weinbauer & Suttle 1996); whereas, the lowest percentage of lysogeny detected by Paul and co-workers was 1.3%, a level near or above several samples in our study. Our lower detection level was because reducing virus concentrations decreased the background, and because estimates of viral abundance

made by epifluorescence microscopy are more accurate and precise than those made by transmission electron microscopy (Hennes & Suttle 1995, Weinbauer & Suttle 1997). If the percentage of lysogens was near zero when Paul and co-workers did not detect lysogenic phage production, then the average percentage of lysogens ranged from ca 5 to 10%. This is only slightly higher than the 2 to 4% measured in our studies and similar to the 0.1 to 7.4% estimated by Tapper & Hicks (1998).

Another approach for estimating the proportion of lysogenic cells is to determine the decrease in bacteria in response to the addition of mitomycin C. Jiang & Paul (1996) estimated that, on average, 35% of cells in natural bacterial communities lysed following mitomycin C exposure. However, toxic effects of mitomycin C may be the reason for the high loss rates of bacteria. It is also possible that chemicals such as polychlorinated biphenols may also be effective inducing agents (Cochran et al. 1998). Finally, if many bacteria in aquatic environments are dead (Zweifel & Hagström 1995) or metabolically inactive (Choi et al. 1996), the percentage of lysogens may be underestimated.

The percentage of inducible lysogens was generally lower in samples collected in coastal than in offshore waters in this study (Table 2) and that of Weinbauer & Suttle (1996). Similarly, Jiang & Paul (1994) found that more isolates were lysogenic from oligotrophic than eutrophic environments, and a higher percentage of bacteria could be induced in communities collected offshore than from coastal waters (Jiang & Paul 1996, Cochran et al. 1998). However, Jiang & Paul (1996) found the highest percentage of inducible lysogens in estuarine environments. They suggested that because bacterial replication and DNA repair depends on growth, fewer lysogens would be induced at oligotrophic stations where cells would be metabolically less active. In our experiments, bacteria were diluted with ultrafiltered seawater to 10% of their initial abundance. This resulted in substantial growth during the experiments, suggesting that bacteria were not nutrient limited. Jiang & Paul (1996) also used a constant burst size to calculate the percentage of lysogenized bacteria. However, estimates of burst size were higher for coastal than offshore waters (Table 2; Weinbauer et al. 1993). Since the number of lysogens is calculated as induced virus production divided by burst size, the use of a constant burst size may underestimate the percentage of lysogens in oligotrophic environments and overestimate them in more productive environments. This might partially explain the difference between the studies. Finally, lysogeny is harder to detect in oligotrophic waters, since a smaller burst size results in less viral production.

Lenski (1988) argued that lysogeny should be favored in environments with high particle loads, because infectivity of free viruses is destroyed by adsorption. Sunlight also destroys viruses (Suttle & Chen 1992, Wommack et al. 1996). UV-B penetrated deeper at offshore than at coastal stations, resulting in higher decay rates of infectivity (Wilhelm et al. 1998). Our results support the view that lysogeny will predominate in environments with high destruction rates of free viruses. High loss rates because of particle adsorption in estuaries and solar radiation offshore may explain why the percentage of lysogens is higher in these environments.

Lysogenic phage production and solar radiation

DNA damage induces SOS repair mechanisms, including activation of protease activity by the RecA protein. In turn, this can initiate the lytic pathway of virus replication in lysogens. UV-B causes DNA damage in marine bacteria (Aas et al. 1996, Jeffrey et al. 1996), and can induce RecA activity (Miller et al. 1995). Thus, the increase in phage production observed in our study as the result of exposure to sunlight may have been caused by activation of RecA and the subsequent induction of lysogens. Others have not found evidence for induction of lysogens when bacterial communities were exposed to solar radiation (Wilcox & Fuhrman 1994, Jiang & Paul 1996). This might be the result of the error associated with using transmission electron microscopy to count viruses, as well as the relatively weak effect of sunlight on inducing lysogens (Table 2).

UV-B penetrated deeper offshore than in coastal waters (Wilhelm et al. 1998). However, the formation of cyclopuridine dimers in DNA dosimeters deployed at the surface for the solar day did not differ between oceanic and neritic stations (data not shown), indicating similar UV exposure in all surface samples. The ratio of viral production in sunlight-exposed versus mitomycin C-treated samples was lower offshore than in coastal waters. This suggests that damaging wavelengths were less effective at inducing lysogens in waters receiving high doses of solar radiation. Virus particles are vulnerable to destruction by sunlight (Suttle et al. 1993, Wommack et al. 1996), particularly in optically transparent offshore waters (Wilhelm et al. 1998). Consequently, selection should favor prophages that are not induced by sunlight, since they benefit from the DNA repair mechanisms of their hosts. Alternatively, sunlight may be less effective at inducing lysogens offshore, because the higher doses of UV-B may have already induced most of the susceptible prophages.

Lysogenic phage production and hydrogen peroxide treatment

Virus production was higher in bacterial communities exposed to hydrogen peroxide than in controls, indicating induction of lysogens. In surface waters of the Gulf of Mexico, hydrogen-peroxide concentrations can be as high as 0.2 μM (Van Baalen & Marler 1966, Zika et al. 1985), and turnover in about a day (Johnson et al. 1989). Rain can increase the concentration more than 10-fold (Cooper et al. 1987). Therefore, the concentrations used in the incubations (0.1 to 1.35 μM) correspond to peak natural levels. Hydrogen peroxide can also result from photochemical degradation of organic matter (Petasne & Zika 1987, Johnson et al. 1989). Thus, solar radiation not only induces lysogenic phage production by directly damaging the DNA, but also indirectly by the production of hydrogen peroxide.

Bacterial mortality and lysogenic phage production

A steady-state model was used to estimate the mortality that could result from induction of lysogens. The model assumes that when bacteria divide, 1 daughter cell dies and the other survives to divide again. Therefore, something that kills 50% of bacteria accounts for 100% of the mortality (Proctor & Fuhrman 1990). Bacterial mortality resulting from induction by sunlight or hydrogen peroxide was calculated using burst size and induced viral production (Table 2). The simultaneous induction of lysogens by either solar radiation or hydrogen peroxide would have caused the lysis of a maximum of 1.7% of the bacterial community, representing 3.5% of the estimated mortality. Given that about 20% of bacteria in the sea are infected by viruses (Suttle 1994), induction of lysogens represents a small percentage of the phage-inflicted mortality.

Simultaneous induction of lysogens by sunlight or hydrogen peroxide would also have a modest effect on viral abundance. The viral production if all lysogens were induced was estimated from data in Tables 1 & 2. The induced viral production was calculated as the % of lysogenic bacteria \times bacterial abundance in the undiluted samples \times burst size/100. The viral production was also expressed as a % of the viral standing stock by dividing the estimated viral production by the viral abundance in the undiluted samples and multiplying by 100. The number of viruses produced as the result of exposure to sunlight ranged from 0 to 2.0×10^6 phages ml^{-1} , or 0 to 22.2% (average: 8.8%) of the standing stock, respectively (Table 2). The range in the number of viruses produced by induction due to hydrogen peroxide was 1.4×10^4 to 1.9×10^6 phages ml^{-1} , or 0.6 to 36.3% (average: 15.3%) of the standing stock,

respectively. These estimates suggest that lysogenic phage production would only occasionally be an important contributor to total phage production (e.g. when lytic phage production is low). These data are consistent with those of Wilcox & Fuhrman (1994) who showed that lytic infection rather than induction of lysogenic bacteria was primarily responsible for phage production of in coastal waters of Southern California.

This paper presents estimates of the percentage of lysogens that can be induced by mitomycin C, solar radiation and hydrogen peroxide in marine bacterial communities. Induction of lysogens by sunlight and hydrogen peroxide was not an important cause of bacterial mortality and was unlikely to have contributed significantly to phage production in oceanic and neritic waters. Consequently, the effects that phages have on mortality, nutrient flow and species diversity in these systems appears to depend primarily on the lytic pathway of infection. The data also indicate that higher percentages of bacteria are lysogenized in oligotrophic waters than in more productive coastal waters. If higher abundances of lysogenic bacteria are caused by high loss rates of infectious bacteria, then increases in UV-B because of atmospheric ozone depletion could result in an increase in the frequency of lysogens in bacterial communities. In turn, this would affect the role that viruses play in microbial food webs.

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