Mesenchymal tumor in the mantle of the mussel *Modiolus difficilis* from Amursky Bay in the Sea of Japan

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ABSTRACT: Morphology, cell composition and histochemistry of a gross mantle tumor of *Modiolus difficilis* are described. The well-differentiated tumor contained numerous cells atypically rich in neutral polysaccharides and bundles of fibrous tissue and muscles. Mitotic activity of the cells of the tumor base ranged from 0 to 0.8%. Architectural disorganization, the presence of collagen and necrotic cells, cellular and nuclear changes, the absence of a protective epithelium and hemocytic infiltration of the tumor base permitted the tumor to develop as a result of chronic irritation and inflammation of the anomalous developing mantle.

KEY WORDS: Tumor Proliferation Mitotic index Mantle Mussel

INTRODUCTION

Tumors of Bivalvia have been investigated more extensively than in other invertebrates (Pauley 1969, Khudoley & Syrenko 1977, Lauckner 1983, Sparks 1985, Mix 1986, Peters 1988, 1994, Pekkarinen 1993). There are some works devoted to the description of external tumors in marine mollusks (Smith 1934, Sparks et al. 1964, 1969, Pauley & Sayce 1966, Pauley 1969, Dix 1972, Dinarnani & Wolf 1973, Harschbarger 1976, and others). External tumors in mollusks of the Far East Region have not been previously reported. We have examined some bivalve species common in the Sea of Japan from a bay near Vladivostok to estimate the frequency of tumor occurrence. The purpose of this work is to describe the microanatomy, cell composition and histochemistry of a large polyoid tumor in the mantle of the mussel *Modiolus difficilis* and to interpret its nature.

MATERIALS AND METHODS

In May 1989, 60 mussels *Modiolus difficilis* were collected from Amursky Bay, Sea of Japan, within the city precincts of Vladivostok, in Vtoraya Rechka Inlet, an area intensively polluted by industrial and domestic sewage. One external tumor was found. The tumor was excised and fixed in phosphate buffered 10% neutral formalin (pH 7.3 to 7.4), containing 3% NaCl, for 36 h at 4°C. Samples were gradually dehydrated and embedded in polyethylene glycol 3000 (Efimov 1984). Sections 4 to 5 μm thick were stained with either Mayer’s hematoxylin and eosin (H&E), methylene blue, Heidenhain’s azan, Van Gison’s stain, Giemsa stain, period acid-Schiff (PAS) reactive-alcian blue-hematoxylin, Feulgen staining, or fast green (Lillie 1965, Pearse 1968). The mitotic index (MI) was determined in 2 zones of the tumor base and in the protective epithelium after examination of 10 tumor sections. The MI was calculated per 170 000 tumor cells of the first zone, per 42 000 tumor cells of the second zone and per 10 000 cells of protective epithelium, respectively.

RESULTS

One large pink polyoid growth was found on the mantle near the adductor muscle of a mussel 12 cm in shell length (Fig. 1). The growth was of a solid consistency, was non-pedunculated, measured 15 mm in...
length and was 4 mm thick. Distinctive portions of the structure were observed (Fig. 2, a–e). The tumor surface was covered by a single-layered columnar ciliated epithelium with associated gland cells (granular eosinophilic cells with proteins, agranular basophilic cells with glucosaminoglycans and fuchsinophilic cells with neutral polysaccharides). Parts of the tumor protective epithelium were absent because of destruction and sloughing (Fig. 2, c–e). We found atypical portions of the protective epithelium: non-convoluted low columnar or cuboidal epithelium on the left side of the tumor body (Fig. 2, d), deeply convoluted epithelium at the top of the tumor and epithelium at the base of the tumor with some mitotic cells and cells with clearly marginated transparent nuclei and large conspicuous nucleoli (Fig. 3).

The outer part of the tumor base consisted of smooth muscular and collagen fibers and some connective tissue cells. Three small bundles of longitudinal muscular and collagen fibers originated from the base and ran to the top (Fig. 2, b). The inner central part of the tumor base (zone I with a slight hemocytic infiltration) was not fibrillar and consisted mainly of cells with a thin-granular cytoplasm and normal vessels. The marginal right side of the tumor base (zone II with a strong hemocytic infiltration) was located under the epithelium and was rich in hemal sinuses, granular eosinophilic and hyaline hemocytes, some minute hemocytes with basophilic nuclei and some atypical hemocytes with abnormal gross nucleus (Fig. 4).

Undifferentiated small cells with basophilic cytoplasm, with large basophilic rounded or oval nuclei and not always noticeable nucleoli, were found in the tumor base. Mitoses were found in cells of different sizes (Figs. 5 & 6), but mainly in small cells. The cytoplasm of mitotic cells usually lacked inclusions, but some large mitotic cells had a granular cytoplasm resembling that of tumor cells. Some mitotic binucleate cells and abnormal mitoses were found. Average mitotic activity in tumor cells was 0.015% in zone I and 0.026% in zone II of the entire tumor base. The overall ML range of the tumor cells of several tumor sections was 0 to 0.8%. The ML of protective epithelial cells was 0.26%, and focal concentrations of mitoses in some tumor sections examined were as much as 2%. Prophases prevailed (50%) among mitotic tumor cells, whereas meta- and anaphases were most numerous (75%) among mitotic epithelial cells (Figs. 3 & 5).

The major part of the tumor consisted of cells with a fine granular cytoplasm (Fig. 7). The tumor cells were intensively stained blue with methylene blue or with Giemsa stain and formed compact accumulations near vessel-like tubules (Fig. 8). They were stained red or lilac with Heidenhain's azan, or pale yellow with Van Gison's stain. Cells containing proteins were atypically rich in neutral polysaccharides and glycogen, whereas cells of the tumor top and base had small amounts of neutral polysaccharides (Figs. 9 to 11). The tumor cells were pleomorphic with a strongly marked nuclear polymorphism. Cells were revealed which contained heterochromatic and large nuclei and nucleoli, micro-
nuclei and chromatin clumps in cytoplasm, fragments of the chromosome bridge, and different Feulgen-stained nuclei (Fig. 12). Undifferentiated small cells (with large basophilic nuclei and small nucleoli) were rare in the major portion of the tumor, with only 2 normal and 1 abnormal mitoses (anaphase with lagging chromosomes) observed here, whereas 64 mitoses were found at the tumor base. The outer right side of

Figs. 3 to 8. *Modiolus difficilis*. Histological structure of mantle polypoid tumor. Fig. 3. Covering epithelium of the tumor base with mitotic cells (arrows) and cells with marginated transparent nuclei and conspicuous nucleoli. H&E. Scale bar = 50 µm. Fig. 4. Cells of the tumor base in zone II (strong hemocytic infiltration and atypical large nuclei) (arrow) H&E. Scale bar = 50 µm. Fig. 5. Prophase in small cells of the tumor base. Scale bar = 50 µm. Fig. 6. Gross telophase in hemal sinus. H&E. Scale bar = 50 µm. Fig. 7. Cells of the tumor body with a granular cytoplasm. Scale bar = 50 µm. H&E. Fig. 8. Accumulations of intensively stained tumor body cells near vessel-like tubules. Giemsa stain. Scale bar = 100 µm.
DISCUSSION

We classified the large polypoid growth on the mussel mantle as a mesenchymal benign tumor, and not as a teratoma or a malignant tumor. The growth originated from multiple locations in the mantle tissue, did not produce metastasized tissues and was characterized by architectural disorganization and pattern atypia rather than by similarity with a normal mantle. The cell pattern consisted of well-differentiated cells and did not resemble that of other mesenchymal tumors of Bivalvia (Smith 1934, Sparks et al. 1964, 1969, Pauley 1969, Dix 1972, Dinamani & Wolf 1973, Harschbarger 1976, Sparks 1985).

We distinguished in the tumor bulk several kinds of fine granular cells, atypical for normal mussel mantle. Some of them were myocytes (positive staining with Van Gison's stain, Heidenhain’s azan and fast green). Others appeared to be connective-tissue cells and differed morphologically from normal mantle storing cells. A third group appeared to be modified hemocytes, because they were colored as intensively with Giemsa stain (Fig. 8) as the hyaline hemocytes of mussels were (Rasmussen et al. 1985). Unlike normal mussel hemocytes (Moore & Loowe 1977) and tumor base cells, all tumor cells types had an abnormally high content of neutral polysaccharides and glycogen.

Similar to other mesenchymal molluscan tumors (Sparks 1985), undifferentiated and mitotic cells were found rarely in the tumor body. However, undifferentiated cells were observed more often in the base portion of the tumor. Mitotic activity in the cells of this portion greatly exceeded that observed in the tumor body cells, but did not reach the average level of mitotic activity which occurs in cells of molluscan mature disseminated neoplasias or of normal renewing and regenerating tissues (Hillman 1963, Usheva & Leibson 1988, Reno et al. 1994). The MI of the tumor base cells...
ranged from 0.01 to 0.03 % and was highest in the zone of strong hemocytic infiltration. Both undifferentiated cells and hemocytes may perhaps proliferate here, and the former became differentiated as cells of the fibroblast line. As they grew, cells protruded outward from the mantle, were modified and formed the tumor bulk. The M₁ of protective epithelium cells was an order higher (0.3%) than that of tumor base cells and comparable with that of normal renewing and regenerating molluscan tissues (Hillman 1963, Usheva & Leibson 1988). This high mitotic level seems to be related to an inflammation of the mussel mantle. The covering epithelium of the growth seemed to be in a state of regeneration.

Births of inflammation of the covering epithelium and some tumor parts under the epithelium were found both in the *Modiolus difficilis* tumor and in other mesenchymal molluscan tumors (Sparks et al. 1964, 1969, Pauley 1969, Dix 1972, Dinamani & Wolf 1973, Harschbarger 1976, Sparks 1985). However, we found an unusual pattern of inflammation in the *M. difficilis* tumor: hemocytic infiltration and cell reproduction occurred mainly in the tumor base whereas a clear atrophy, a marked destruction of muscular tissue and collagen formation were observed on the top and in the right portions. Muscle atrophy may have been related to nutrient destruction, and the vessel-like tubules with thickened walls may have developed as a result of chronic inflammation of normal vessels. These structures did not resemble the pseudotubes or rosette-like structures found in oyster mesenchymal tumors (Sparks et al. 1969), but were similar to pathological vascular vessels of diseased mollusks *Mya arenaria* (Barry et al. 1971). Mantle vessel-like tubules had tumor cells whereas these cells were not found in normal vessels of the tumor base. The cells of the vessel-like tubules seem to be modified hemocytes and/or endothelial cells.

These facts confirm the supposition that a mantle tumor in *Modiolus difficilis* had developed due to inflammation and chronic irritation caused by an unknown agent. Destruction, cell sloughing, detachment of necrotic flaps from the top of the tumor, partial ulceration and deep invaginations of the covering epithelium suggest protracted irritation of this tumor. We assume that the irritant had a viral or chemical nature, taking into account the fact that the mollusks examined were collected from the polluted inlet of Amursky Bay where some Bivalvia have even become extinct (Tkalin et al. 1993, Silina & Ovsyanykova 1995). The cytogenetic anomalies of the interphase and mitotic cells seem to be related more to the influence of the pollutants than to dysplasia-like changes of tissues and result in a breakdown of cell homeostasis. The same cytogenetic anomalies associated with the polluted environment were observed in the larvae of marine animals (Klumpp & Von Westernhagen 1995).
Thus, the mussel mantle tumor was characterized as an unusual pattern of fibromyoma in a state of irritation. Cell proliferation was mainly observed in the base of the tumor, whereas the detachment of flaps was found at the top of the tumor.

LITERATURE CITED


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