Some new observations on the cytopathology of fin erosion disease in winter flounder

Pseudopleuronectes americanus

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ABSTRACT: A light and electron microscopic study was conducted on dorsal fin tissues adjacent to acute fin erosion lesions in winter flounder from 2 polluted sites (New York Bight region and New Haven Harbor) on the northeast Atlantic Coast. The objective of this work was to evaluate these minimally affected, lesion-associated tissues which may precede the acute or severe stages of the disease. The following 4 types of pathological conditions were found in the epidermis of diseased fish from the 2 polluted sites: (1) epithelial cell hyperplasia; (2) mucous cell hyperplasia and hypertrophy; (3) spongiosis; and (4) focal necrosis. The latter 2 types of lesions have not been previously reported for fin erosion in this species. Changes in the dermis associated with these lesions included fibrosis, abnormal distribution of melanocytes, hyperemia and sclerosis of blood vessels, and hemorrhage. The possibility that hypoxia may play a role in the observed pathology is considered.

KEY WORDS: Winter flounder · Pollution · Fin erosion disease · Pathology

INTRODUCTION

Fin erosion disease in bottom-dwelling species has served as a biomarker for environmental contamination of estuarine and near-shore waters along all coasts of the United States (McCain et al. 1988, O’Connor & Huggett 1988, Overstreet 1988) and in some locations in Europe (Vethaak 1987, Dethlefsen 1988). A good correlation has been observed between the high prevalence of this disease and contaminated sediments, high levels of contaminants in fish tissues, and other histopathological abnormalities. Winter flounder Pseudopleuronectes americanus demonstrating fin erosion disease have been collected in the waters of the New York Bight (Murchelano & Ziskowski 1982) and other locations along the northeast coast of the United States (Ziskowski et al. 1987) where the sediments have been shown to be heavily contaminated by a wide variety of pollutants (Zdanowicz et al. 1986, Grønlund et al. 1991, Johnson et al. 1993). In addition, Khan et al. (1992) have observed fin erosion disease in feral winter flounder exposed to bleached kraft mill effluent (BKME).

Murchelano’s (1975) observations on the histopathology of severe fin-erosion lesions in winter flounder from the New York Bight revealed that they were most commonly present in the mid-portions of the dorsal and anal fins. He observed a thinning of and loss of epithelium in a distal-proximal direction accompanied by fin-ray degeneration and resorption. In addition, the lesions demonstrated hyperplasia of the epithelium, eosinophilic granular cells (EGC’s) and mucous cells. Melanocytes typically observed to be evenly distributed along the epidermal-dermal junction were found to be concentrated in regions where the epithelium was denuded and often located deeply within the dermis in association with blood vessels and nerves. Although Murchelano (1975) observed occasional foci of lymphocytic infiltration, no microorganisms could be detected. Changes in the dermis underlying the eroded fins included fibrosis, hyperemia, and hemorrhage. Fibrotic scarring was present in areas where fin degeneration had been severe. The objective of this study was to provide for the first time a light and
electro microscopic (EM) description of the normal cytology of fin tissues in winter flounder. In addition, through the use of higher resolution methods than those employed by Murchelano (1975), such as the examination of plastic-embedded, 1 to 2 μm thick sections by light microscopy and thin sections by EM, it was hoped that a description of the cytopathological changes in tissues adjacent to acute fin-erosion lesions would provide further information on the pathogenesis of this disease, for which there is no presently known etiology in this species.

MATERIALS AND METHODS

All the fish examined were collected by otter trawl and varied in length from 25 to 40 cm. Thirteen fin-eroded fish were collected from the New York Bight region which included Raritan or Sandy Hook Bays, NJ in 1974–1975. Seven diseased fish were captured in New Haven Harbor (Morris Cove) during 1987 to 1988, and 4 fish from Great Bay, NJ (a relatively clean site) were sampled as controls during 1975. Four additional control fish were collected in 1992 from the Niantic River (Niantic, CT), a site which had been shown to be minimally contaminated (Gronlund et al. 1991). All flounder having acute fin erosion and non-diseased controls were measured at the time of capture and killed by a blow to the head, and tissue from the tip of the dorsal fin adjacent to well-established, severe lesions found in diseased fish, as well as comparably located tissue (i.e., tip of the dorsal fin) from control fish was excised and prepared for microscopy as described below. A piece of distal fin tissue approximately 5 mm in thickness and 10 mm in length was removed from both the fin-eroded and control fish and placed on a piece of dental wax. It was then carefully divided into small pieces (4 to 6) containing at least 2 fin rays per segment, which were placed in fixative.

All tissues taken from fish captured in the New York Bight region and Great Bay were fixed in ice cold (0 to 4°C), 4% glutaraldehyde in 0.1 M phosphate buffer at pH 7.4 for at least 24 h and kept refrigerated until further processing, which was completed no later than 1 wk after collection. Microscopic examination of these tissues indicated that this fixative might be slightly hypertonic (e.g., red blood cells [RBCs] slightly crenated). Therefore, all tissues from New Haven Harbor fish were fixed in 2% glutaraldehyde using the identical buffer and procedure as stated for the New York Bight region and Great Bay specimens. Control tissues from Niantic River fish were fixed in both 2 and 4% glutaraldehyde in the same buffer as mentioned above. Subsequent to primary fixation all fin tissues were washed 4 times in 0.1 M phosphate buffer (pH 7.4) and acclimated to room temperature before post-fixation for 1 h in 1% osmium tetroxide in 0.1 M phosphate buffer at pH 7.4. The tissues were then dehydrated in a graded series of ethanols, including 2 washes in 100% ethanol and 4 washes in propylene oxide. Next, the tissues were infiltrated and embedded in Spurr’s (1969) resin. Thick sections were cut from no less than 4 blocks for each control or fin-eroded fish at 1 to 2 μm with glass knives and stained with toluidine blue dye. Because no decalcifying agents were employed, thick sections were not taken through the fin rays, but as close as possible to them. Light microscopic examination of thick sections permitted both photomicrography and selection of fields to be thin-sectioned (60 to 90 nm) with diamond knives. Thin sections were stained with alcoholic (50%) uranyl acetate and lead citrate before examination, and electron micrographs were taken with a Zeiss EM9 S2 electron microscope.

All of the 4 to 6 pieces from each fish were examined for pathological abnormalities. If any of the fin pieces from a given fish demonstrated a particular abnormality, the fish was scored positive for that abnormality with regard to the data expressed in Table 1.

RESULTS

Observations on control fish

The stratified squamous epithelium on the fins of winter flounder was principally composed of a superficial layer of variably shaped epithelial cells which frequently stained metachromatically with toluidine blue dye (Figs. 1 & 2). A middle layer of epithelial cells typically oval to elongate in appearance was several cells thick and a basal layer of cuboidal or columnar cells

Table 1. Lesions associated with fin erosion in winter flounder (stated as percent of total). No significant difference was found for any lesion type upon comparing the results from the control versus the contaminated sites using the Fisher Exact Test (2-tailed). NYB: New York Bight region (a contaminated site). NHH: New Haven Harbor, CT (a contaminated site). GTB: Great Bay, NJ (a control site). NTR: Niantic River, CT (a control site).

<table>
<thead>
<tr>
<th>Type</th>
<th>NYB (n = 13)</th>
<th>NHH (n = 7)</th>
<th>GTB (n = 4)</th>
<th>NTR (n = 4)</th>
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<tr>
<td>Focal necrosis</td>
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<td>43</td>
<td>25</td>
<td>25</td>
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<tr>
<td>Epithelial hyperplasia</td>
<td>23</td>
<td>71</td>
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<tr>
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<td>54</td>
<td>57</td>
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<td>25</td>
</tr>
<tr>
<td>Mucous cell hyperplasia</td>
<td>39</td>
<td>85</td>
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<td>92</td>
<td>57</td>
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<td>0</td>
</tr>
<tr>
<td>Hemorrhage</td>
<td>46</td>
<td>43</td>
<td>0</td>
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cytoplasm contained apical, dense layers of fine filaments, an oval nucleus with
margined heterochromatin, a well-devel-
oped Golgi complex and an endoplasmic
reticulum system (ERS), plus numerous
vesicles which in some instances (Fig. 4)
appeared to be released at their apical
plasmalemma. At their apical borders the
surface epithelial cells shared junctional
complexes. Numerous desmosomes were
observed between these cells along their
lateral and basal cell membranes. The
epithelial cells in the middle layer were
typically oval to elongate in shape and had
centrally placed, rounded nuclei with
prominent nucleoli (Fig. 5). The cytoplasm
of these cells was electron dense owing
mainly to the large number of fine fila-
ments which communicated with the num-
erous desmosomes found around the cells'
periphery. The round to oval-shaped mito-
chondria and elements of the ERS were
frequently observed in close proximity to
the cell’s nucleus. In addition to numerous
desmosomes, these cells also had marginal
plicae which interdigitated with adjacent cells. The
basal cells (Fig. 6), which rested firmly on the basal
lamina, were columnar or cuboidal in shape and had
elongate nuclei, and numerous desmosomes and pli-
cae were observed on their surfaces. The cytoplasm of

resting on a well-developed basement membrane was
also present. Densely staining, oval, eosinophilic gran-
ular cells as originally described by Roberts et al. (1971)
for the plaice Pleuronectes platessa were located in
the basal or middle cell layer (Figs. 1 & 2), and mucous
cells were interspersed at the epidermal
surface (Fig. 2). The epithelium of the skin
in close proximity to the fin rays was typi-
cally thicker (40 to 60 μm) than that found
between the rays (20 to 35 μm), but no dif-
ference in cell types was present. Further,
no difference in cell type or composition
was noted for the epidermis upon compar-
ing the pigmented and non-pigmented
sides of the fin tissues. Immediately below
the basal cells a prominent and often un-
dulating basement membrane was found,
which signaled the beginning of the der-
is, and subjacent to it thick bands of
collagen fibers (C; Figs. 1 & 2) were ob-
served.

When observed by electron microscopy,
the surface epithelium was a layer of
effete, electron lucent cells, with scattered
cells sloughing from the surface (EC;
Fig. 3). Immediately below the surface
layer of effete cells were pleomorphic,
electron dense cells (Figs. 3 & 4) that bore
the typical microridges reported for many
species of teleosts (Whitear 1986a). Their

Fig. 1 Photomicrograph of the fin epithelium (pigmented side) from a
control fish (Great Bay) near the fin rays. Note the melanophores (M) within
the thick collagen band (C) below the well-defined basement membrane.
(EGC = eosinophilic granular cell) x473

Fig. 2 Photomicrograph of the fin epithelium from a control fish (Niantic
River) midway between a pair of fin rays. The basement membrane and
associated collagen band (C) have a distinctly scalloped appearance. A
mature mucous cell (MC) is observed at the epithelial surface. (EGC =
eosinophilic granular cell; M = melanophore) x467
the basal cells contained organelles similar to the cells found in the middle layer, but these were more uniformly dispersed throughout the cytoplasm. Oval-shaped eosinophilic granular cells having eccentrically located nuclei and numerous dark granules and small lymphocytes, sometimes containing electron-dense granules, were frequently found among the basal cells (Fig. 7). Mucous cells were not observed in the basal cell layer.

On the flounder's pigmented or eyed side (Fig. 8), melanophores were located in the dermis either within or slightly below the dense band of connective tissue beneath the basement membrane (Fig. 8). In areas close to the fin rays, the stratum spongiosum beneath the thick band of collagen was composed of a loose, connective-tissue matrix that mainly contained capillaries and small groups of nerves. The stratum compactum in this area contained larger blood vessels and nerves embedded in dense connective tissue. The hypodermis, in this instance, could not be clearly identified. The dermis in the inner fin tissue between the rays, as shown in Fig. 2, was a more uniform layer of loose connective tissue containing small blood vessels, nerves, and a few melanophores.
some instances represented a 4- to 5-fold increase in height from that of normal skin. Occasionally, melanomacrophages as well as mucous cells were found at the surface of the epithelium. In all instances, the hyperplastic foci lacked the stroma or dermal papillae described for epidermal papillomas (Peters 1984, Smith et al. 1989).

Spongiosis

Epidermal and mucous cell hyperplasia was frequently observed to be accompanied by spongiosis (intercellular edema) as defined for fish integument by Roberts & Bullock (1976) and Roberts (1989). When present in the epithelium in close proximity to the fin rays, the epithelial cells in the middle epidermal layer had either an oval (Fig. 11) or elongate (squamous) shape (Fig. 12). The abnormally large extracellular spaces permitted one to visualize the spinous processes between the affected cells. The basal cells and surface cells remained more or less intact. In the thinner fin tissue between the rays (Fig. 13), spongiosic lesions in the epithelium were more limited in scope and demonstrated widened intercellular spaces which had a cleft-like appearance. Frequently, the underlying blood vessels within the dermis were hyperemic.

Observations on diseased fish

Six types of abnormalities were commonly observed in the fin tissues of diseased fish from either the New York Bight region or New Haven Harbor (Table 1). Owing to the small sample size, no significant difference was observed for the prevalence of any lesion type upon comparing the results from the control versus the contaminated sites after applying the Fisher Exact Test (2-tailed).

Epithelial and mucous cell hyperplasia

In areas between the fin rays where the normal skin often had a folded configuration, the hyperplastic epidermis frequently demonstrated an irregular, villus-like shape (Fig. 9). A greatly increased number of mucous cells, many of which appeared to be hypertrophic or empty (spent), were sometimes present. Eosinophilic granular cells were also found (EGC; Fig. 9) scattered throughout the hyperplastic tissue but did not appear to be in greater numbers in the hyperplastic epithelium of fish from the contaminated sites when compared with the epithelium of fish from control sites. The epidermis in fin tissue near the fin rays (Fig. 10) was thickened (e.g., 200 to 300 μm) and in

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Fig. 5. Electron micrograph of a mid-level epithelial cell from a control fish (Great Bay). Note the dense, filamentous cytoplasm (F) around the cell's periphery, and the desmosomal junctions (D) between adjacent cells. [N = nucleus] ×6285

Fig. 6. Electron micrograph of a columnar-shaped basal cell from a control fish (Great Bay) that was separated from a large band of collagen (C) by a prominent basal lamina (BL). Like the epithelial cells found in middle layer of the epithelium, the basal cells had numerous plicae (P) on their surfaces and desmosomal junctions (D) were readily observed between them. [N = nucleus] ×7550
regions containing either oval (Fig. 14) or squamous cells (Fig. 15) that were in the beginning stages of degeneration. Sometimes, the intercellular spaces between these cells contained an amorphous material of low electron density (Fig. 14) and the cellular processes of the epithelial cells frequently exhibited multiple and/or fused desmosomal junctions. Within the cytoplasm of these cells, some changes associated with cell death, such as enlarged mitochondria, moderate vacuolation, and accumulation of glycogen, were observed. Pathological changes within necrotic cells present in spongiotic tissues (Fig. 16) included the following: (1) vesiculate nuclei; (2) high amplitude mitochondrial sweling and loss of cristae; (3) a breakdown and/or condensation of the highly ordered cytoplasmic filaments; (4) extensive vacuolation; (5) vesiculation of the plasmalemma and loss of cytoplasmic components, particularly glycogen, to the intercellular space; and (6) a loss of integrity of desmosomal junctions between epithelial cells. Ultrastructural observations of spongiotic changes in the thinner,
Focal necrosis

Necrotic regions of limited dimension within the fin epithelium differed from the pathological conditions described above in that they always involved the death and disruption of the surface epithelial cells. In fin tissues close to the rays (Fig. 20), necrosis and sloughing were observed at the epithelial surface and were sometimes accompanied by cell death and necrosis in the middle layer of the epidermis. In fin tissues distant from the rays (Fig. 21), the epidermis of diseased fish sometimes had a villus-like configuration and the surface epithelial cells were swollen and necrotic. In some instances, the epithelial cells within the epidermis demonstrated nuclear pyknosis. Densely staining EGC cells were accompanied by marked leucocytic infiltration.

DISCUSSION

The dorsal fin integument of normal winter flounder lacks both scales and the highly specialized cell types found in other fishes (Whitear 1986a), and is struc-
mast cells and have been shown to degranulate when stimulated by bacterial extracellular products (Ellis 1985) or neurotransmitters (Powell et al. 1991). The dermis in normal fin tissue of winter flounder was not well organized into a stratum spongiosum, stratum compactum, and hypodermis as has been described for the skin of other fish species (Bullock & Roberts 1974, Whitear 1986b). The stratum spongiosum and stratum compactum and the structures they typically include were found only in close proximity to the fin rays, but were not present in the thinner tissue between the rays. Comparison between the structure of the dermis observed for the fin tissues in winter flounder and that for other fish species was difficult, as the author was unable to find a thorough histological description of the dermis underlying the fin tissues of other species. Typically, morphological studies on the fins of fish have focused on the histology of the lepidotrichia and their associated structures (Lanzen 1976, Becerra et al. 1971), except that the irregularities in thickness observed for the epidermis found on the dorsal fin of the plaice were not observed in this species. Unlike the surface epithelial cells described for the plaice, however, those in the dorsal fin tissues of winter flounder stained metachromatically with toluidine blue dye, indicating the presence of acidic glycoproteins (Clark 1981), and appeared to be secretory based on the fine structural nature of their cytoplasm. Thus, these cells may provide secretions that bathe the skin as has been described by Whitear & Mittal (1984) for the blenny Blennius pholis (L.) and for several species of Indian carp (Singh & Mittal 1990). Lymphocytes routinely found in the fin epithelium of winter flounder, but not described in the plaice, may be involved in the generation of antibacterial immunoglobulins found in the skin mucus of other fish species (Peleteiro & Richards 1985, Alexander & Ingram 1992). Eosinophilic granular cells (EGCs), structurally similar to those described in the skin of the plaice (Roberts et al. 1971), were regularly observed in the dorsal fin epithelium of winter flounder. Morphologically similar cells have been extensively studied in salmonids where they are believed to be analogous to mammalian mast cells and have been shown to degranulate when stimulated by bacterial extracellular products (Ellis 1985) or neurotransmitters (Powell et al. 1991). The dermis in normal fin tissue of winter flounder was not well organized into a stratum spongiosum, stratum compactum, and hypodermis as has been described for the skin of other fish species (Bullock & Roberts 1974, Whitear 1986b). 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heavily contaminated with a wide variety of anthropogenic substances and various types of liver lesions have been reported in winter flounder collected from them (Murchelano & Wolke 1985, Gronlund et al. 1991, Johnson et al. 1993).

Unlike fish from the polluted sites mentioned above, winter flounder from the Great Bay control site off the southern New Jersey coast have not demonstrated high fin erosion disease prevalence (Ziskowski et al. 1987), liver lesions, or high contaminant levels in the sediments (Zdanowicz et al. 1986, Johnson et al. 1993). Similarly, the Niantic River, from which control tissues were also taken for this study, has been shown to be a minimally contaminated region of Long Island Sound (Gronlund et al. 1991).

Roberts & Bullock (1976) and Roberts (1989) have indicated that epidermal hyperplasia in fish results in the addition of cells at all levels of the epithelium and may be caused by a variety of factors including chemical pollutants. The observations reported herein regarding epidermal hyperplasia, melanophore aggregation, and dermal fibrosis are in good agreement with earlier histopatho-
Fig. 15. Electron micrograph of a spongiotic lesion in the middle epithelial layer of squamous cells near a fin ray from a fish collected at a contaminated site (New York Bight). Note the eosinophilic granular cell (EGC) and lymphocyte (L) in the expanded extracellular space. ×5278

Fig. 16. Electron micrograph of an advanced spongiotic lesion in the middle layer of necrotic, oval-shaped epithelial cells near the fin ray from a fish collected from a contaminated site (New Haven Harbor). Note the vesiculate nuclei (VN), the vesiculations of their plasmalemmae (V), the high amplitude swelling of their mitochondria (M), the loss of glycogen (G) to the intercellular space, and the breakdown of their intercellular junctions. (D = desmosomes) ×5278
Bodammer: Fin erosion disease in winter flounder

Fig. 17. Electron micrograph of a spongiform lesion in the thin, inter-ray epidermis found in a fish from a contaminated site (New York Bight). The extensive intercellular space between the degenerating cells has resulted in several cleft-like spaces (CL) within the necrotic tissue. (PN = pyknotic nucleus; MM = melanomacrophage; BM = basement membrane) ×5429

logical studies of more severe fin erosion lesions in winter flounder (Murchelano 1975), starry flounder and English sole from the Duwamish River Estuary (Wellings et al. 1976), and perch or goldfish exposed to BKME (Lindesjöö & Thulin 1994). EGCs were observed at various levels within the epithelium but did not appear to be as numerous as reported by Murchelano (1975) or Wellings et al. (1976). As Roberts et al. (1971) have shown for the plaice, melanomacrophages were occasionally observed near the surface of the normal epidermis. Although epidermal papillomas have been widely observed in fish from both polluted and clean environments (Peters 1984, Smith et al. 1989), reports on nonpapillomatus epidermal hyperplasia in feral fish appear much more limited but have been observed in fish from polluted environments (Haensly et al. 1982, Bucke et al. 1983, Bruno & Ellis 1988, Dethlefsen 1988).

Mucous cells, particularly those found in the delicate tissues of the gill, respond to a wide variety of toxicants and/or irritants by undergoing hyperplasia or hypertrophy. If the noxious stimulus persists, cellular de-

Fig. 18. Photomicrograph of the epidermal-dermal border (--) from a fish collected in a contaminated site (New York Bight) illustrating the abnormal aggregation of melanocytes (M) at the junction between the epidermis (EP) and the underlying fibrotic dermis (FD). Congested blood vessels (BV) and hemorrhage (H) were also observed. ×364
Fig. 19. Photomicrograph of the dermis from the same fish illustrated in Fig. 18 but at an adjacent and deeper location. Note the melanocytes (M) within the fibrotic dermis (FD) and in association with the sclerotic blood vessels (SV). Hemorrhage (HE) is also apparent. ×364

specimens, a comparison between mucous cells observed in the control tissue and those found in in some of the diseased specimens suggested that those present in the latter were frequently both hyperplastic and hypertrophic. In addition, the existence of a number of empty mucous cell profiles in the diseased fin tissues indicated that cell depletion may have occurred. These observations are comparable to those of others studying fin erosion in flatfish. Murchelano (1975) described mucous cell hyperplasia in winter flounder, and Wellings et al. (1976) observed both mucous cell hyperplasia and depletion in 2 flatfish species, the starry flounder Platichthys stellatus and English sole Parophrys vetulus from Puget Sound. Mucous cell hyperplasia has also been reported in eroded fins of goldfish Carassius auratus exposed to BKME (Sharples & Evans 1996), and in English sole exposed to the water soluble fraction of crude oil (Hawkes 1977). Daye & Garside (1976) observed hypertrophy and excess mucous secretion on the skin of brook trout Salvelinus fontinalis exposed to elevated levels of pH. Mucous cell depletion has been reported in the integument of the brown bullhead Ictalurus nebulosus after long-term exposure to 3 ppm of copper (Benedetti et al. 1989), and in the carp Cyprinus carpio after exposure to organically fertilized pond water (Iger et al. 1988).

Spongiosis or intercellular edema most probably resulting from an inflammatory response of the skin (Roberts & Bullock 1976, Roberts 1989) was observed in the fin epithelium of winter flounder from both of the contaminated sites. Murchelano (1975) did not mention spongiosis in his histopathological description of fin erosion in winter flounder, nor has it been described by other investigators who have conducted histopathological studies of other feral fish species which had fin erosion disease (Klontz & Bendele 1973, Wellings et al. 1976, Lindesjö & Thulin 1994, Sharples & Evans 1996). Although spongiosis was not mentioned by Lindesjö & Thulin (1994) in their study of goldfish exposed to BKME, it appears to have been present in the epithelium of the fish they studied which had acute fin erosion lesions (e.g., Figs. 17 & 19).

The pathological changes observed in the dermis as reported herein are in agreement with those found in fin-eroded tissues of both winter flounder (Murchelano 1975) and goldfish (Lindesjö & Thulin 1994) and included hemorrhage and blood vessel congestion where epidermal hyperplasia and/or spongiosis was present. Murchelano (1975) was the first to consider the possibility that ischemia resulting from hemorrhage and/or hyperemia in the dermis affected the blood supply to the overlying epithelium in the diseased fins of winter flounder. There is, however, no experimental evidence

Fig. 20. Photomicrograph of a focal lesion (FL) adjacent to a region of hyperplastic epidermis (HP) in the epithelium close to the fin rays of a fish collected at a contaminated site (New Haven Harbor). The cells at both the epithelial surface and within the middle cell layer (ML) are necrotic. ×350
to prove that ischemia is the cause of the pathology observed in the diseased fins of winter flounder. Roberts (1989) has indicated that degenerative changes in epidermis of fish are often the result of a 'vascular change' within the dermis. Therefore, it is possible that hypoxia may have been the cause of the spongiosic condition observed within the epidermis of the fish studied herein. Similarly, Lindesjöö & Thulin (1994) believe an undefined 'physiological disturbance' may have resulted in the pathological changes in the dermis leading to the necrosis of fin tissues in goldfish exposed to BKME.

Whereas the necrosis and loss of small areas of epithelium were not reported by Murchelano (1975) for eroded fins in winter flounder, Wellings et al. (1976) reported small ulcers of this type in the English sole and starry flounder. Small lesions were prevalent in the fin tissues of fish found at both of our polluted sites. Focal necrosis of the fin tissues was found in areas close to the fin rays or in the skin between them, and unlike the other types of severe or extensive lesions that have been described by others, degeneration and sloughing of surface cells were typically observed. Recently, Sharples & Evans (1996) described progressive, severe necrosis commencing at the level of the surface epithelium (accompanied by a marked lymphocytic infiltration) as the principal mechanism leading to acute fin erosion disease in feral goldfish exposed to BKME. Unlike the observations by Sharples & Evans (1996), no marked infiltration of lymphocytes or other types of leucocytes were observed to be associated with the focal necrotic lesions or any other type of lesions observed in this study. Lesions in the surface epithelia of fish may result from exposure to a variety of organochlorine (e.g., DDT) and other pesticides (Meyers & Hendricks 1985), exposure to heavy metals (Gardner 1975, Bodammer 1985, Benedetti et al. 1989), and abnormal levels of environmental pH (Daye & Garside 1976).

As is indicated in Table 1, a low percentage of several lesion types was observed in the fin epithelium of fish from the control sites. There are a number of reasons why this may have occurred. Microscopic lesions may have been present in the dorsal fins of fish that did not demonstrate grossly visible lesions. The Great Bay site in southern New Jersey waters served as a control location for the extensive studies being conducted by Murchelano (1975) during the early and mid 1970s because the water and sediment quality of the area was considered to be 'relatively pristine' in comparison with the conditions found in the New York Bight region. However, as the extensive studies of Ziskowski et al. (1987) on the prevalence of fin erosion disease in winter flounder along the northeast coast of the United States have shown, fish from the southern New Jersey coastline, which includes Great Bay, had a low prevalence of fin erosion disease. Therefore, it is possible that the fin tissues of fish collected from Great Bay for this study had some of the microscopic changes associated with fin erosion disease but were undetected for the reason stated above. Although the prevalence of fin erosion disease in winter flounder at the Niantic River control site was not studied by Ziskowski et al. (1987), this site serves as a reproductive area for winter flounder which may have migrated there from various regions in Long Island Sound, where fin erosion disease was observed (Ziskowski et al. 1987). While winter flounder are not considered to be a highly migratory species (Gray 1990), the problem of their possible migration and the relationship of this behavior to fin erosion disease is borne out in the fact that Ziskowski et al. (1987) found, much to their surprise, that the highest prevalence of fin erosion disease in winter flounder was observed in fish captured in the Gulf of Maine, which is not considered to be a 'polluted' area. They hypothesized that the winter flounder from the Gulf of Maine which had fin erosion disease were migrants from 'nearby environmentally stressed estuarine populations'.

Because contaminant concentrations in fish tissues, sediments, or the water column were not investigated...
in the present study, it is not possible to designate a specific contaminant or a combination thereof as the causal factor for the lesions that were observed on the dorsal fins of winter flounder as reported herein. Because fin erosion disease in bottom-dwelling fish has been observed in urban and industrially polluted waters along all coasts of the United States and in sites of poor water quality in Europe, it would seem unlikely that a specific pollutant or identical combinations of pollutants would be found in similar concentrations at the various locations where fin erosion disease has been reported. Given all the etiological possibilities that may exist at the numerous locations where this ubiquitous disease has been observed, it seems safest to assume at this time that fin erosion disease in winter flounder, as well as in other fish species, results from the generalized responses of the fin epithelium and dermis (e.g., hyperplasia, spongiosis, necrosis, dermal fibrosis, hyperemia, etc.) to a potentially wide variety of toxic substances. Thus, the disease appears to remain, as Murchelano (1982) has previously stated, as one of 'uncertain etiology'.

This report is the first in which both light and electron microscopy have been used to examine both the normal and pathological fin tissues in winter flounder which have fin erosion disease. The results obtained from tissues located in close proximity to well-established fin erosion lesions provide general support for the earlier work on winter flounder (Murchelano 1975), other feral flatfish species, and goldfish exposed to BKME (Lindesjöo & Thulin 1994). No evidence was found in this study for a bacterial or viral etiology for fin erosion disease, and both epidermal and mucous cell hyperplasia observed in the integument of diseased fish were in keeping with the generalized reactions of fish skin to irritants and/or toxic insults. Spongiosis and focal necrotic lesions of limited scope are reported for the first time in winter flounder with fin erosion disease. Vascular and fibrotic changes within the dermis of diseased fish were observed that may be responsible for the breakdown of the normal epidermal-dermal junction, and could thereby precede or initiate further pathologic change in the epithelium owing to hypoxia. A multiplicity of lesion types were found in close proximity to each other in the affected integument, any or all of which could ultimately lead to necrosis of the fin tissue.

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