NOTE

**Synechocystis** MCCB 114 and 115 as putative probiotics for *Penaeus monodon* post-larvae

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ABSTRACT: *Synechocystis* MCCB 114 and 115 were segregated as putative probiotics for shrimp larvae from a collection of 54 cyanobacterial cultures enriched from seawater. On feeding *Penaeus monodon* post-larvae with the cyanobacteria, the generic diversity of the intestinal bacterial flora could be enhanced with substantial reduction or total absence of *Vibrio* spp. A significant difference (p < 0.001) in the percent survival of batches of post-larvae fed on the cyanobacterial cultures was observed and, on repeated challenge with *V. harveyi*, the relative percent survival of those batches of larvae fed on *Synechocystis* MCCB 114 and 115 was significantly higher. The *Synechocystis* MCCB 114 and 115 cultures were found to contain high levels of protein (34 to 43%), in addition to carotenoids.

KEY WORDS: Probiotic · Vibriosis · Antagonism · *Vibrio* · Aquaculture · *Synechocystis* · Cyanobacteria · Shrimp

INTRODUCTION

Probiotics are defined as live microbial adjuncts that have beneficial effects on their host by modifying the host-associated or ambient microbial community, ensuring improved use of feed by enhancing its nutritional value, host’s response to disease, or by improving the quality of the host’s ambient environment (Verschuere et al. 2000). Cyanobacteria (blue-green algae) have not yet been suggested as candidate probiotics in aquaculture. However, marine cyanobacteria in particular have been in focus for quite some time as a potential source of antagonistic compounds (Pergament & Carmeli 1994, Mundt et al. 2001). The secretion of antimicrobial substances is an important part of the competition process in the natural environment, having a beneficial effect on the host by modifying the host-associated ambient microbial community (Verschuere et al. 2000). Cyanobacteria are a good source of nutrients (Morist et al. 2001) and their cellular contents may promote the growth of gut bacteria and accelerate the process of nutrient absorption. Considering the above, the present study was undertaken to screen and segregate specific cyanobacterial cultures from the marine environment as putative probiotics to be used in shrimp hatcheries. This selection was based on (1) *in vitro* antagonism, (2) ability to regulate the intestinal bacterial flora, and (3) enhancement of the survival of *Penaeus monodon* post-larvae after challenge with *Vibrio harveyi* subsequent to being fed the cyanobacteria.

MATERIALS AND METHODS

Cyanobacterial cultures. Water samples (n = 9) were collected from surface waters of the sea surrounding Vypeen Island (09° 58'N, 76° 17'E) using a Niskin...
water sampler (General Oceanics). Samples were transferred aseptically to sterile glass bottles of 100 ml capacity, and brought to the laboratory in an ice chest within 2 h; 1 ml of each sample was then inoculated in each of 6 enrichment media: BG-11, ASN-111 and MN (Rippka et al. 1979), modified Chu-11 (Cohen 1975), Allen and Nelson’s medium (Allen & Nelson 1910) and Walne’s medium (Walne 1974), all prepared in sea water. Samples were incubated at 28 ± 0.4°C under 12 h illumination at 40 µE m−2 s−1, provided by cool fluorescent lamps (Philips), resulting in 54 cyanobacterial enrichment cultures.

**Screening.** Primary and secondary screening were accomplished by evaluating the antimicrobial activity of cell-free culture supernatants (Sample 1), cell lysate after sonication (Sample 2), cell debris separated by centrifugation of the sonicated cells (Sample 3) and intact washed cells (Sample 4) on a panel of 14 species of *Vibrio* (*V. alginolyticus, V. harveyi*, *V. splendidus, V. parahaemolyticus, V. vulniﬁcus, V. cinc- natiensis, V. marinus, V. mimicus, V. proteolyticus, V. mediterranei, V. ﬂuvialis, V. proteolyticus* and *V. cholerae*) via the standard disc diffusion method (Jayaprakash et al. 2005). The *Vibrio* spp. were previously isolated from shrimp larval rearing systems, characterized following Alsina & Blanch (1994), and maintained at the National Centre for Aquatic Animal Health (NCAAH) as part of its culture collection.

The effect of cyanobacterial cultures (segregated during the secondary screening) on the intestinal bacterial flora of *Penaeus monodon* post-larvae (PL-20; 0.1 to 0.15 g) was tested by administering them with commercial feed (Higashimaru India) on a daily basis. The feed was coated with cyanobacteria at the time of feed-coating using ‘Bindex’ (Matrix Vet Formulations) at 1:1 ratio, dried in a desiccator at room temperature (28 ± 0.1°C) for 2 h, and fed ad libitum to *P. monodon* for 20 d. The control tanks were maintained with normal feed. The PL-20 (n = 50) used for the experiment were maintained in duplicate fiberglass containers with 25 l sea water (30 ppt). NH4+–N and NO2–N of the rearing water were measured following Strickland & Parsons (1972). Chlorophyll content (APHA 1995), carotenoids (Barnes & Blachstock 1973) and lipids (Hedge 1962) and proteins (Bradford 1976) of cells were determined.

**Biochemical analysis.** The cyanobacterial cultures were harvested by centrifugation at 10,000 × g for 15 min at 4°C, washed repeatedly in saline (0.85% NaCl), and dried in a vacuum desiccator at room temperature (28 ± 0.5°C) to constant weight. Samples were centrifuged at 10,000 g for 15 min at 4°C for 15 min every 3 d, and the anti-*Vibrio* activity of the supernatant determined via standard disc diffusion using selected isolates of *Vibrio* spp. The media and the stage/phase at which each culture exhibited maximum activity were recorded.

**Growth phase vs. antagonistic activity of cyanobacterial cultures.** To determine the phase of growth of the cyanobacterial cultures at which maximum anti-*Vibrio* activity was exhibited, the cultures were inoculated in the above media and incubated until the stationary phase. During this period, aliquots of 2 to 4 ml from each culture were centrifuged at 10,000 × g at 4°C for 15 min every 3 d, and the anti-*Vibrio* activity of the supernatant determined via standard disc diffusion using selected isolates of *Vibrio* spp. The media and the stage/phase at which each culture exhibited maximum activity were recorded.
RESULTS

Out of 54 cyanobacterial cultures, 52 showed antagonism to at least 1 isolate of *Vibrio* spp. and, from this group, 7 cyanobacterial cultures (C-44, C-45, C-48, C-50, C-51, C-53 and C-54) could be segregated in such a way that antibacterial activity could be detected against all the *Vibrio* spp. isolates. Whenever antagonistic activity in the culture supernatant was detected (Sample 1), it could also be demonstrated in the cell lysate (Samples 2 and 3). However, at times the activity was observed only in the cell lysate, not in the culture supernatant.

*Vibrio* spp. could not be recovered from the intestines of post-larvae fed with C-45, C-51, C-53 and C-54. However, heterotrophic bacterial flora in the intestines of post-larvae fed on the experimental diets with cyanobacterial cultures C-45, C-51 and C-54 exhibited comparatively higher generic diversity indices (Table 1).

A significant difference (p < 0.001) in the percent survival of batches of post-larvae fed on different cyanobacterial cultures was observed. The least significant difference (LSD) at the 0.1% level was calculated in order to identify the significant treatments, and determined to be 1.1051. Administration of cyanobacteria C-45, C-48, C-50, C-51, C-53 and C-54 resulted in a significantly higher survival rate in the test group of post-larvae, which were apparently healthy and pigmented compared with the control. On repeated challenge with *Vibrio harveyi*, the RPS of the batches of larvae fed on C-51 and C-54 were significantly higher than those of all others (Table 2). Based on our observations, the cultures C-51 and C-54 were segregated as the putative probionts for shrimp larval rearing systems.

*Synechocystis* C-51 exhibited antagonistic activity in Walne’s medium during the log phase, attaining a maximum at the end of the phase. However, maximum antagonistic activity of *Synechocystis* C-54 in ASN-111 was observed during the stationary phase. For generating biomass under the experimental conditions, the media Chu-11 and BG-11 were found to be most suitable for C-51 and C-54, respectively.

<table>
<thead>
<tr>
<th>Protein (mg 100 mg⁻¹)</th>
<th>C-51</th>
<th>C-54</th>
</tr>
</thead>
<tbody>
<tr>
<td>34.74 ± 2.92</td>
<td>43.43 ± 7.43</td>
<td></td>
</tr>
<tr>
<td>Carbohydrate (mg 100 mg⁻¹)</td>
<td>22.82 ± 5.76</td>
<td>20.57 ± 2.27</td>
</tr>
<tr>
<td>Lipid (mg 100 mg⁻¹)</td>
<td>33.18 ± 12.23</td>
<td>28.25 ± 2.19</td>
</tr>
<tr>
<td>Chlorophyll a (µg mg⁻¹)</td>
<td>6.89 ± 1.83</td>
<td>3.99 ± 0.64</td>
</tr>
<tr>
<td>Carotenoid (µg mg⁻¹)</td>
<td>0.85 ± 0.13</td>
<td>0.39 ± 0.08</td>
</tr>
<tr>
<td>Allophycocyanin (mg 100 mg⁻¹)</td>
<td>4.48 ± 1.62</td>
<td>2.14 ± 0.55</td>
</tr>
<tr>
<td>C-phycocyanin (mg 100 mg⁻¹)</td>
<td>1.26 ± 0.39</td>
<td>0.5 ± 0.08</td>
</tr>
<tr>
<td>Phycoerythrin (mg 100 mg⁻¹)</td>
<td>Nil</td>
<td>Nil</td>
</tr>
</tbody>
</table>

DISCUSSION

Over the years, application of probiotics in aquaculture has gained an impressive momentum, which is focused on sustainability in the production process. Different groups of microorganisms such as *Lacto-
bacillus spp., Bacillus spp., Pseudomonas spp., Micrococcus spp., certain strains of Vibrio, yeast and microalgae have been reported as putative probiotics. (Gomez-Gil et al. 2000, Verschuere et al. 2000, Irianto & Austin 2002). Several of these microorganisms have been identified as possessing antagonistic properties and have been designated as 'biocontrol agents' (Verschuere et al. 2000). It is in this context that the present study was undertaken to screen cyanobacteria enriched from the marine environment to be used as candidate probiotics in shrimp larval production systems. More than 96 % of the cyanobacterial cultures examined in this study exhibited in vitro antagonism to at least 1 Vibrio isolate.

The anti-Vibrio property of the cyanobacteria investigated here derived from both extracellular and intracellular sources, strengthening the argument of Chetsumon et al. (1995) that bioactive molecules are initially accumulated intracellularly. If so, the cells might be used as probiotics to regulate the intestinal bacterial flora favorable to shrimp post-larvae through antagonism.

These cultures showed antagonism against known aquaculture pathogens such as Vibrio harveyi and V. parahaemolyticus (Karunasagar et al. 1994, Jayaprakash et al. 2005). It is postulated that the survival of those Penaeus monodon post-larvae fed the Synechocystis C-51 and C-54 and subjected to repeated challenge with V. harveyi may be a result of an antagonistic probiotic effect. Moreover, the higher protein content of Synechocystis C-51 and C-54 (equivalent to that of commercial feed) and the presence of carotenoids also make the cultures ideal feed additives, which would contribute to the weight gain and colouration of shrimp post-larvae. During this study, those post-larvae fed with Synechocystis were found to be visibly more pigmented than the control animals. Furthermore, as suggested by Becker (1994), phycobiliproteins such as C-phycocyanin and allophycocyanin from different species of cyanobacteria have growth-promoting activities and immunostimulatory properties as well.

In addition to coating feed particles with the cyanobacterial cells, the Synechocystis C-51 and C-54 can be bioaccumulated in rotifers (Sakamoto et al. 1996) and fed to crustacean larvae. Additionally, the cyanobacterial cultures as such or the washed cells can be added directly to the larval rearing system.

When the cyanobacterial cultures were fed to shrimp post-larvae, the generic diversity index of the intestinal heterotrophic bacterial flora increased along with either a concomitant reduction in or total absence of Vibrio spp. in those individuals fed with Synechocystis C-51 and C-54. This is in accordance with the observations of Rico-Mora et al. (1998), who reported that a bacterial strain associated with Skeletonema costatum had prevented the establishment of V. alginolyticus in crustacean larvae.

In conclusion, this study suggested that Synechocystis C-51 and C-54 could be considered as candidate probiotic organisms suitable for shrimp culture. These cyanobacterial cultures form part of the microbial culture collection of the NCAAH, Cochin University of Science and Technology, and are catalogued as Synechocystis MCCB 114 and Synechocystis 115.

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