Microenvironment and photosynthesis of zooxanthellae in scleractinian corals studied with microsensors for $O_2$, pH and light

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ABSTRACT: During experimental light-dark cycles, $O_2$ in the tissue of the colonial scleractinian corals Favia sp. and Acropora sp. reached >250% of air saturation after a few minutes in light. Immediately after darkening, $O_2$ was depleted rapidly, and within 5 min the $O_2$ concentration at the tissue surface reached <2% of air saturation. The pH of the tissue changed within 10 min from about 8.5 in the light to 7.3 in the dark. Oxygen and pH profiles revealed a diffusive boundary layer of flow-dependent thickness, which limited coral respiration in the dark. The light field at the tissue surface (measured as scalar irradiance, $E_o$) differed strongly with respect to light intensity and spectral composition from the incident collimated light (measured as downwelling irradiance, $E_d$). Scalar irradiance reached up to 180% of $E_d$ at the coral tissue surface for wavelengths subject to less absorption by the coral tissue (600 to 650 nm and >680 nm). The scalar irradiance spectra exhibited bands of chlorophyll a (chl a) (675 nm), chl c (630 to 640 nm) and peridinin (540 nm) absorption and a broad absorption band due to chlorophylls and carotenoids between 400 and 550 nm. The shape of both action spectra and photosynthesis vs irradiance ($P$ vs $I$) curves depended on the choice of the light intensity parameter. Calculations of initial slopes and onset of light saturation, $I_s$, showed that $P$ vs $E_d$ curves exhibit a lower initial slope and a higher $I_s$ than corresponding $P$ vs $E_o$ curves. Coral respiration in light was calculated as the difference between the measured gross and net photosynthesis, and was found to be >6 times higher at a saturating irradiance of 350 µEin m$^{-2}$ s$^{-1}$ than the dark respiration measured under identical hydrodynamic conditions (flow rate of 5 to 6 cm s$^{-1}$).

KEY WORDS: Light-enhanced respiration · $O_2$ and pH dynamics · Action spectra · $P$ vs $I$ curves · Scalar irradiance

INTRODUCTION

Symbiotic photosynthetic dinoflagellates (= zooxanthellae) are common inhabitants of many metazoan animals in the tropics, e.g. foraminifera, porifera and cnidarians (Trench 1981). In the polyp tissue of reef corals, high densities of zooxanthellae belonging to the genus Symbiodinium are found (Porter et al. 1984). These are among the most important primary producers in tropical reef communities, estimated to account for 1 to 10% of total benthic production on a global scale (Muscatine 1990).

Studies of the photophysics of zooxanthellae and their interaction with the host coral have inherent methodological problems due to the close interaction of zooxanthellae and coral metabolism within the <0.5 to 1 mm thin layer of tissue growing on the calcified coral skeleton. Carbon fixed by the zooxanthellae
might be used for biosynthesis and respiration within the symbionts or translocated to the surrounding animal tissue, where it is used for respiration and biosynthesis (Muscatine et al. 1981, Falkowski et al. 1984). Part of the fixed C is excreted e.g. as mucus, or is incorporated into skeletal carbonate. Oxygen produced by zooxanthellae photosynthesis is used partly for symbiotic respiration and partly for coral respiration. This intimate association between autotrophic and heterotrophic processes in corals (and in many other algal-animal symbioses) makes the quantitative and spatial separation of respiratory and photosynthesis difficult when \(^{14}\text{C}\) isotopes or \(\text{O}_2\) exchange methods are used (Muscatine et al. 1981, Muscatine 1990). Similar methodological problems exist in measurements of the primary production in sediments inhabited by microalgae and in photosynthetic microbial mats (Revsbech et al. 1981). A method for measuring gross photosynthesis independent of respiration is available based on the use of \(\text{O}_2\) microelectrodes (Revsbech et al. 1981, Revsbech & Jørgensen 1983).

The microenvironment of the zooxanthellae is virtually unstudied, although microsensors for pH and \(\text{O}_2\) and the light-dark shift technique for photosynthesis measurements, have previously proven useful in the study of chemical gradients and transport mechanisms within algal-animal symbiosis (Jørgensen et al. 1985, Revsbech & Jørgensen 1986). The few published \(\text{O}_2\) microelectrode measurements in corals have dealt with the diffusive boundary layer (DBL), which regulates the diffusive exchange between the coral tissue and the surrounding seawater (Patterson 1992, Shashar et al. 1993). The dynamics of pH and \(\text{O}_2\) in layers of endolithic algae in the coral skeleton has also been studied with \(\text{O}_2\) microelectrodes and pH minielectrodes (Shashar & Stambler 1992).

Together with the chemical microenvironment, the light field inside the coral tissue is the most important factor regulating zooxanthellae photosynthesis. The optical properties of coral tissue are not known and only a few studies of the light field near coral reefs have been published [Roos 1967 (cf. Falkowski et al. 1990), Jaubert & Vasseur 1974, Brakel 1979, Williams & Carpenter 1990]. Recently, fiber-optic microprobes have been developed to measure directional radiant fluxes (field radiance) and spherically integrated total radiant flux (scalar radiance) (Jørgensen & Des Marais 1986, Kuhl & Jørgensen 1992, Lassen et al. 1992). Microscale light measurements with these probes in sediments and microbial mats demonstrated that the light field near the sediment surface may be very different in spectral composition and intensity from the incident radiant flux (irradiance) (Kuhl & Jørgensen 1992, 1994, Lassen et al. 1992, Kuhl et al. 1994a, b). The scalar irradiance in sediments reached up to 200% of incident irradiance at the sediment surface due to intense multiple scattering. The combination of the incident collimated light with diffuse scattered light, which was spectrally altered due to absorption by photopigments in the sediments, resulted in a strong spectral distortion relative to the incident light. Similar effects of light scattering have been postulated to occur in corals (Brakel 1979, Falkowski et al. 1990) but to our knowledge no measurements of the light field at the level of single corals are available.

In this study we present direct measurements of \(\text{O}_2\), pH, light and photosynthesis in the tissue of scleractinian corals. The goal was to characterize the chemical and physical microenvironment of the zooxanthellae and their surrounding coral host. Our results represent point measurements within single coral polyps and demonstrate basic principles of respiration and photosynthesis regulation in corals.

**MATERIALS AND METHODS**

**Sampling and experimental setup.** Small branches of *Acropora* sp. and *Favia* sp. were sampled by SCUBA diving from 5 to 10 m depth in the Gulf of Aqaba next to the H. Steinitz Marine Biological Laboratory, Eilat, Israel. The *in situ* seawater temperature was 20 to 23°C and maximal light intensities at the sampling depth reached 980 \(\mu\text{Ein} \cdot \text{m}^{-2} \cdot \text{s}^{-1}\) downwelling irradiance (Shashar et al. 1993). Most specimens had been exposed to natural light conditions and exhibited pale yellow-brownish/green tissue. One colony head of *Favia* sp. had been exposed for an extended period (>2 yr; Razi Wigo pers. comm.) to very low light conditions in the shade of a larger coral assemblage and had dark chocolate-brown tissue.

After sampling, the corals were transferred directly to an aquarium with continuously aerated seawater kept at room temperature (21 to 23°C). Water movement in the aquarium was created by blowing air from an aquarium pump onto the water surface through a Pasteur pipette. The approximate flow velocity of the water over the coral was determined by timing the movement of small suspended particles in the water under a dissection microscope. Visible light was incident vertically from above using a tungsten-halogen lamp of constant color temperature but variable light intensity (Schott KL1500 with neutral density filters) and a daylight filter inserted in the light path. The lamp was equipped with a fiber-optic cable with a collimating lens. Experiments were done within 24 to 42 h after sampling.

**Oxygen and pH measurements.** Oxygen concentration was measured with Clark-type \(\text{O}_2\) microelectrodes (Revsbech & Ward 1983) connected to a custom-built...
In the *Favia* sp. measurements the O₂ microelectrodes were also equipped with a guard cathode (Revsbech 1989). Signals were either recorded on a strip-chart recorder or collected by a computer via an A/D card and a custom-made Pascal program. The electrodes had a tip diameter of 6 to 12 μm, a 90% response time of <0.5 s, and a stirring sensitivity of 1 to 2%. Linear calibration of the O₂ microelectrodes was done from readings of the electrode current in air-saturated seawater and in nitrogen-flushed seawater. The O₂ concentration in air-saturated seawater was calculated from measurements of salinity and temperature using the formula given by Garcia & Gordon (1992).

pH was measured with glass microelectrodes (Revsbech & Jorgensen 1986) connected to a high impedance electrometer (Radiometer pH meter or Keithley type 617) with a calomel electrode (Radiometer pH meter or Keithley type 617) with a calomel electrode (Radiometer type 401) as a reference. The tip diameter of the pH microelectrodes was 30 to 50 μm. Signals were recorded on a strip-chart recorder. Electrodes were calibrated at room temperature in standard pH buffers (pH 4, 7, 8 and 9) and exhibited a near-Nernstian response of 58 pm to 60 mV per pH unit.

**Light measurements.** Incident downwelling quantum irradiance was measured with an underwater 400 to 700 nm scalar irradiance meter (LiCor) or with an underwater scalar irradiance meter (1.5 cm probe diameter; Biospherical Instruments) positioned over a light well of black PVC at approximately the same distance and position in the collimated light field as the coral sample. Microscale light measurements were done in *Favia* sp. corals with a fiber-optic scalar irradiance microprobe (Lassen et al. 1992, Kühl et al. 1994b) connected to a custom-built light meter consisting of a photomultiplier (Hamamatsu HC120-05) equipped with a hot mirror (Melles Griot) transmitting only visible light to the detector. By covering the coral tissue with a small piece of black velvet, measurements of downwelling light intensity were done with the fiber-optic scalar irradiance sensor positioned at approximately the same distance from the light source as the coral tissue surface. The measurements of scalar irradiance presented in this study were all normalised to this measure of downwelling irradiance. A direct conversion of microscale light measurements into absolute units of light intensity was not possible as the fiber-optic light meter was not spectrally calibrated.

Besides measurements of visible light integrated from 400 to 700 nm, we also measured spectral scalar irradiance. This was done by the same procedure as described above but with a continuous 400 to 720 nm interference filter (HBW 10 to 14 nm, Schott) mounted in a holder at the output of the fiber-optic lamp (more details in Jorgensen & Des Marais 1986). By sliding the interference filter in the holder it was possible to scan through the visible spectrum in 10 nm steps illuminating the coral with monochromatic light.

**Positioning of microsensors.** All microsensors were positioned by either a manually operated or a motorized micromanipulator (Mätzähäuser Germany; L.O.T. Germany) controlled by a computer and custom-made Pascal software. In the *Acropora* sp. samples, measurements were done with the microelectrodes mounted vertically and the light incident at an angle. In most of the *Favia* sp. experiments, the direction of measurement was at 45° relative to the vertically incident light. Measurements were done in vertical steps of 50 to 100 μm. Determination of the tissue surface position was done by observation of the microsensors under a dissection microscope while slowly approaching the surface. In the following, all depths are related to the tissue surface (depth = 0), where negative and positive depths indicate depth above and below the tissue surface, respectively.

**Gross photosynthesis measurements.** Gross photosynthetic rates (nmol O₂ cm⁻² s⁻¹) in the tissue were determined with O₂ microelectrodes by the light-dark shift method (Revsbech & Jorgensen 1983). By this method the gross rate of photosynthesis is estimated as the rate of O₂ production during 1 to 2 s after the onset of darkness, assuming (1) a steady-state O₂ profile before darkening, (2) an identical O₂ consumption before and during the dark period, and (3) identical diffusive fluxes during the measurements [a detailed discussion of the light-dark shift method can be found in Revsbech & Jorgensen (1983) and Glud et al. (1992)]. The gross photosynthetic rates determined by use of O₂ microelectrodes may include carbon equivalents that are subsequently lost through photorespiration (Glud et al. 1992).

**Net photosynthesis and respiration.** Diffusive fluxes of O₂, J(x), were calculated from steady-state O₂ profiles by Fick's first law of diffusion (Revsbech & Jorgensen 1986):

\[
J(x) = -\Phi D_e \frac{\partial C(x)}{\partial x}
\]

where \(\partial C/\partial x\) is the slope of the O₂ profile at depth \(x\), \(\Phi\) is the porosity, and \(D_e\) is the apparent diffusion coefficient of O₂. In seawater \(D_e = D_n\), the molecular diffusion coefficient of O₂ in water, which is a function of temperature and salinity (Broecker & Peng 1974, Li & Gregory 1974, N. B. Ramsing & J. Gundersen unpubl. data). In our case \(D_n = 2.02 \times 10^{-5} \text{ cm}^2 \text{s}^{-1}\) [calculated from a seawater temperature of 21°C and a salinity of 40‰ by the formula in Li & Gregory (1974)].

The total flux of O₂ out of the photosynthetic part of the coral tissue, i.e. the sum of the downward flux into
tissue. Only in a few measurements did we succeed in penetrating the tissue directly with the microelectrode without inducing enhanced mucus secretion, while in most cases the tissue was only compressed by the microsensors. We tried to cut the tissue with a small hypodermic needle or with the tip of a scalpel before inserting the microsensors into the tissue. This resulted in the secretion of large amounts of mucus, which coated the tip of the microsensors and significantly affected their response. In most cases our measurements were thus limited to the upper 0.0 to 0.2 mm of the tissue and the overlying water. However, this still allows reliable measurements of $P$ vs $I$ curves and of action spectra of the zooxanthellae photosynthesis.

**RESULTS**

**Oxygen and pH dynamics**

In the light, the zooxanthellae photosynthesis resulted in a build-up of $O_2$ in the tissue of up to 250% air saturation and a tissue pH of up to 8.6, i.e., 0.7 pH units above the pH value of the overlying seawater, due to intense $CO_2$ fixation (Figs. 1 & 2). In the dark, $O_2$ was depleted by the polyp and zooxanthellae respiration, and almost anoxic (<2% air saturation) conditions were found in the coral tissue, which now had a low pH of 7.3 to 7.4 (Fig. 2B). Both $O_2$ and pH profiles demonstrated the presence of a 200 to 300 μm thick DBL that separated the coral tissue and the overlying flowing seawater.

**Fig. 1.** Acropora sp. Depth profiles of $O_2$, pH and gross photosynthesis (bars) in the tissue of a branched stony coral sampled from 5 to 6 m water depth. Incident photon irradiance (400 to 700 nm) was 535 μEin m$^{-2}$ s$^{-1}$
Coral tissue

Photosynthesis (nmol O₂ cm⁻³ s⁻¹)

Fig. 2. Favia sp. Depth profiles of (A) O₂ and (B) gross photosynthesis and pH in the tissue of a stony coral sampled from 5 to 6 m water depth. (●) Profiles measured in darkness; (○) profiles measured at an incident photon irradiance of 350 µEin m⁻² s⁻¹. Photosynthetic rates are indicated by bars for every 0.1 mm interval in the coral tissue (flow velocity was 5 to 6 cm s⁻¹).

The thickness of the DBL varied with the flow velocity and increased from a thickness of 200 to 300 µm at a flow velocity of 5 to 6 cm s⁻¹ to 500 to 600 µm at a flow velocity of 1 to 2 cm s⁻¹, and to >1 mm thickness in stagnant water (Fig. 3, DBL thickness indicated by arrows). The O₂ concentration in the coral tissue decreased with increasing DBL thickness, and anaerobic conditions were reached in stagnant water.

During experimental light-dark cycles, the O₂ concentration and pH of the coral tissue exhibited large variations. The O₂ concentration in the upper 0.1 mm tissue layer of a Favia sp. coral started to decrease immediately after the onset of darkness and changed from 240% air saturation to <2% saturation in <5 min (Fig. 4A). Oxygen started to build up as soon as the light was turned on again and reached >100% air saturation in <1 min. The pH in the upper 0.1 mm exhibited a similar variation to the O₂ concentration although the observed changes were slower (Fig. 4B). Variations between a steady-state tissue pH of 7.3 to 7.4 in darkness and pH 8.4 to 8.5 in light were observed within 5 to 10 min. The surrounding seawater had a pH of 7.9. The steady-state tissue pH in Favia sp. varied with the incident light intensity (Fig. 5). Largest changes of tissue pH occurred between darkness and ~100 µEin m⁻² s⁻¹ of incident irradiance. Similar dynamic changes of pH and O₂ with light intensity were also found in the polyp tissue of an Acropora sp. branched coral (data not shown; Revsbech & Jørgensen 1986).

Light intensity and spectral composition

The scalar irradiance reached a maximum in the outer tissue layers of a high-light adapted Favia sp. coral (Fig. 6). The scalar irradiance maximum reached 125 to 180% of incident irradiance at 550 to 650 nm and >180 to 200% at wavelengths >690 nm. The highest maximum was found in the thin tissue covering coral skeleton ridges. In between ridges the scalar irradiance maximum was less pronounced (maximal values of 100 to 130% of incident irradiance). At a depth of ca 0.2 mm below the tissue surface, only light at wavelengths >690 nm reached >100% of incident irradiance.

Integrated 400 to 700 nm scalar irradiance reached maxima of up to 160 to 180% of incident irradiance in the tissue of a high-light adapted Favia sp., while scalar irradiance in a dark-colored, low-light adapted
Fig. 4. *Favia* sp. (A) Oxygen and (B) pH variations in the upper 0.1 mm of the tissue of coral during light-dark shifts. Incident photon irradiance was 350 μEin m⁻² s⁻¹. Dotted lines: O₂ content and pH of the ambient seawater.

*Favia* sp. reached maxima of up to 130 to 140 % of incident irradiance in the upper 0.0 to 0.1 mm of the tissue (data not shown).

The scalar irradiance spectra exhibited distinct minima at absorption wavelengths of the major zooxanthellae chlorophylls (chl a at 675 to 680 nm, chl c at 625 to 640 nm). A shoulder in the spectra around 590 nm was probably due to a secondary absorption maximum of chl c (Shibata & Haxo 1969). Carotenoids caused a broad absorption band at 450 to 550 nm with shoulders in the spectra at 480 to 490 nm and at 540 to 550 nm indicating the presence of the dinoflagellate carotenoid peridinin (cf. Halldal 1968, Shibata & Haxo 1969).

**Spatial heterogeneity**

The distribution of gross photosynthetic activity was measured in a transect across a single *Favia* sp. polyp (Fig. 7). Photosynthetic activity varied from <5 nmol O₂ cm⁻³ s⁻¹ in the relatively flat area surrounding the mouth opening of the contracted polyp up to 32 nmol O₂ cm⁻³ s⁻¹ in the tissue covering the inner ridges of coral skeleton. High activity was also found in the tissue of the oral disk.

**Action spectra**

The most distinct feature of the action spectrum of zooxanthellae photosynthesis in the tissue of *Acropora* sp. was the relatively broad maximum at 620 to 680 nm due to chl a (peak at 675 nm and shoulder at 620 nm) and chl c (small peak at 580 to 590 nm and shoulder at 640 to 650 nm) (Fig. 8). In the blue part of the spectrum, photosynthetic activity due to chl a absorption at 430 to 450 nm, chl c absorption at 460 to 470 nm, and peridinin absorption at 490 to 540 nm was found as shoulders or small peaks in the action spectrum (photopigment absorption wavelengths according to Halldal 1968, Shibata & Haxo 1969, Prézelin et al. 1976).
Photosynthesis versus irradiance

The gross photosynthesis of the zooxanthellae in an Acropora sp. as a function of the incident integrated 400 to 700 nm irradiance (PAR) is shown in Fig. 9. The hyperbolic tangent function of Jassby & Platt (1976) was fitted to the \( P \) vs \( I \) data by a nonlinear least-square Levenberg-Marquardt algorithm (Origin 3.0, MicroCal Software, Inc.) giving an initial slope \( \alpha = 0.04 \) and a maximum photosynthetic rate \( P_{\text{max}} = 11 \text{ nmol O}_2 \text{ cm}^{-3} \text{ s}^{-1} \). The irradiance at the onset of light saturation, \( I_k \), could thus be calculated as \( I_k = P_{\text{max}}/\alpha = 275 \text{ µEin m}^{-2} \text{ s}^{-1} \). The hyperbolic tangent function has been shown to give the best fit to \( P \) vs \( I \) data for corals (Chalker 1981).

Another \( P \) vs \( I \) curve was measured in a Favia sp. (Fig. 10). When the gross photosynthesis \( P \) was plotted against the incident irradiance \( E_{\text{in}} \), we obtained an initial slope \( \alpha = 0.09 \), a maximum photosynthetic rate \( P_{\text{max}} = 13.65 \text{ nmol O}_2 \text{ cm}^{-3} \text{ s}^{-1} \), and an onset of light saturation at \( I_k = 152 \text{ µEin m}^{-2} \text{ s}^{-1} \) from the fitted hyperbolic tangent function. If \( P \) was plotted against the
scalar irradiance (assuming a scalar irradiance of 180% of the incident irradiance) the respective parameters from the curve fit were $\alpha = 0.05$, $P_{\text{max}} = 13.65 \text{ nmol O}_2 \text{ cm}^{-2} \text{ s}^{-1}$, and $I_g = 273 \mu\text{Ein m}^{-2} \text{ s}^{-1}$.

**Net photosynthesis and respiration**

The O$_2$ budget for the coral tissue of *Favia* sp. in the dark and at a saturating irradiance of 350 µEin m$^{-2}$ s$^{-1}$ is shown in Table 1. The fluxes of O$_2$ across the DBL into the tissue (dark respiration) and out of the tissue (net photosynthesis) were calculated from the dark and light steady-state O$_2$ profiles respectively, shown in Fig. 2. Total gross photosynthesis was calculated by depth integration. Respiration was found to be 0.055 nmol O$_2$ cm$^{-2}$ s$^{-1}$ in the dark and 0.366 nmol O$_2$ cm$^{-2}$ s$^{-1}$ in the light, and thus accounted for 11 and 77%, respectively, of the O$_2$ production by gross photosynthesis.

**DISCUSSION**

**Dynamics of the chemical microenvironment**

The high metabolic activity of zooxanthellae and the surrounding polyp tissue, coupled with diffusion as the major exchange process with the surroundings, creates a dynamic microenvironment in the coral tissue (Figs. 1 to 5). At irradiances of >100 µEin m$^{-2}$ s$^{-1}$, the photosynthetic activity of the zooxanthellae resulted in supersaturation with respect to O$_2$ (>250% air saturation), and to alkaline conditions within the coral tissue (>0.5 pH units above the ambient seawater level) due to CO$_2$ fixation by the microalgae. Our results show that the tissue of *Favia* sp. and *Acropora* sp. living in shallow water is supersaturated with O$_2$ during most of the day, thus supporting earlier observations reported in the literature (e.g. Crossland & Barnes 1977, Shick 1990, Patterson et al. 1991, Shashar et al. 1993).

In the dark, coral respiration depleted O$_2$ to near-anoxic conditions (<5% air saturation), and a low pH (>0.5 pH units below seawater pH) was found throughout the tissue. The change from a steady-state light to a steady-state dark situation or vice versa occurred within minutes after a change in the light field (Figs. 4 & 5). This demonstrated the dynamic balance between respiration and photosynthesis, and consequently how the chemical microenvironment in the coral is regulated by fluctuations in light intensity (e.g. shading by clouds passing the sun). The pH changes were slower (5 to 10 min) than the O$_2$ variations (<1 min) due to the relatively large buffering capacity of the bicarbonate pool in the tissue and surrounding seawater. Although no light-dark cycle measurements in the tissue were presented, similar dynamics of O$_2$ and pH have been found in the DBL above a *Favia favus* coral (Shashar & Stambler 1992), in the skeleton of the coral *Porites compressa* (Shashar et al. 1993) and in the symbiotic foraminiferan *Globigerinoides sacculifer* (Jørgensen et al. 1985).

**Diffusive boundary layers and diffusion geometry**

The DBL and its function as a diffusion barrier for solute exchange between the coral tissue and the surrounding turbulent water is discussed by Patterson et al. (1991) and Patterson (1992). The presence of a DBL over the coral tissue was evident from the measured steady-state microprofiles of O$_2$ and pH in *Favia* sp. (Figs. 2 & 3). As there is no significant production or consumption of O$_2$ within the DBL, the 1-dimensional concentration gradients of O$_2$ should be linear within the DBL with a gradual transition at the upper DBL boundary to the O$_2$ concentration of the overlying tur-
bulent water (Jørgensen & Des Marais 1990). The DBL thickness can be found by extrapolation of the linear DBL gradient out to the bulk concentration (Jørgensen & Revsbech 1985). In the corals, DBL thickness increased with decreasing flow velocity resulting in a higher mass transfer resistance and thus a lower increased with decreasing flow velocity resulting in a

Both in stagnant water and at a flow velocity of 2 cm s⁻¹ the DBL thickness was ~200 μm. The effect on the diffusion geometry of the surrounding ridge tissue 0.5 to 1 mm away was thus negligible, and solute transport between the coral tissue and the overlying water could be described by 1-dimensional diffusion. The DBL thickness increased to ~600 μm at 1 to 2 cm s⁻¹ and a 3-dimensional, tubular geometry would be required to adequately describe the diffusion geometry. The O₂ profiles measured above the tissue at low flow velocities were thus affected by the O₂ consumption of the surroundings, and 1-dimensional flux calculations were no longer valid. This effect of diffusion geometry was even more pronounced under stagnant conditions (Fig. 3), where O₂ in the 1 to 2 mm thick layer of overlying water was depleted by the surroundings along the measurement direction, and where the O₂ profile exhibited a curvature characteristic of a 3-dimensional diffusion geometry (cf. Crank 1975, Jørgensen et al. 1985).

Shashar et al. (1993) studied the DBL over single Favia favus and Stylophora pistillata coral polyps and demonstrated how the DBL follows polyp morphology. Both in stagnant water and at a flow velocity of 5 cm s⁻¹ the thickest DBL (1 to 5 mm) was found over the central oral disk of the polyp, while 1 to 2 mm thick DBLs were found near the skeleton ridges of the investigated corals. The DBL thickness and the dimensions of the coral polyps (1 to 10 mm polyp width) reported by Shashar et al. (1993) indicate that at least the central part of the polyps exhibited a 3-dimensional diffusion geometry under the given experimental conditions. Equations applicable to tubular diffusion geometries in corals were described by Patterson (1992).

The presence of a microsensor may also impose changes in the locally measured diffusive flux by compressing the DBL, thus resulting in a steeper concentration gradient across the DBL (Glud et al. 1994). For 5 μm thin microelectrodes the compression of the DBL was found to increase the O₂ flux by ~10 to 60% in various marine sediments relative to measurements in which the microelectrode was inserted from below the sediment surface. The compression effect by the microelectrode was ascribed to perturbations of the local flow velocity field, which were, however, independent of the flow velocity over a flat surface. In a recent study of microbial mats, it was shown that the boundary layer compression only showed significant effects on the O₂ concentration in biofilms with a smooth and homogeneous surface, while no effect was found when the surface was irregular (Lorenzen et al. 1994). When present, the compression effect did not affect the gross photosynthesis measured by the light-dark shift technique. The extent of DBL compression in our coral measurements and how this effect interacts with the diffusional geometry within coral polyps awaits combined high-resolution measurements of the DBL and the flow field around corals under more defined hydrodynamic conditions. One-dimensional diffusion equations should be used with care in corals, which exhibit a complex polyp morphology that interacts with the water flow and leads to flow-dependent differences in diffusion geometry within the polyp.

The DBL for other solutes like CO₂, Ca²⁺ and nutrients will be similar to the oxygen DBL in corals (Patterson et al. 1991, Patterson 1992). Water motion and its effect on DBL thickness is thus a crucial factor in the overall regulation of coral metabolism and increased flow velocity has been shown to increase respiration, photosynthesis and calcification in corals (Dennison & Barnes 1988, Barnes & Chalker 1990, Patterson et al. 1991).

**Photosynthesis and light-enhanced respiration**

In comparison with traditional gas-exchange techniques, a major advance of the O₂ microelectrode light-dark shift technique for photosynthesis measurements is the independence of the measured gross photosynthetic rate from respiration (Revsbech et al. 1981, Revsbech & Jørgensen 1983, Glud et al. 1992). The high spatial and temporal resolution of the method thus enabled us to measure directly gross photosynthesis of the zooxanthellae while exposed to the chemical and physical gradients present within the coral tissue (Figs. 1, 2, 7 to 10). Gross photosynthesis of corals is traditionally measured indirectly as the sum of dark respiration and net photosynthesis under the assumption that dark respiration is identical to the respiration rate in light (Muscatine et al. 1981, Edmunds & Davies 1988, Muscatine 1990).

The respiratory activity of the coral tissue in the light cannot be inferred from gas-exchange or ¹⁶C measure-
ments (Muscatine 1990) but can now be obtained from O₂ microelectrode measurements. Steady-state O₂ profiles measured in the coral express the net outcome of photosynthesis, respiration and diffusive transport. Thus, net photosynthesis in the coral tissue can be calculated from steady-state O₂ concentration profiles as the diffusive flux of O₂ out of the coral tissue in the light (Table 1). An analogous calculation of the diffusive flux into the coral tissue from steady-state O₂ profiles in the dark yields the dark respiration of the coral. These measures of net photosynthesis and dark respiration, although point measurements, are comparable to the data traditionally obtained with gas-exchange studies of coral photosynthesis and respiration (e.g. Crossland & Barnes 1977, Muscatine 1980). The difference between the depth-integrated gross photosynthesis measured by the light-dark shift technique and the net photosynthesis calculated from steady-state O₂ profiles yields a measure of the total respiration of the coral tissue in the light (Jensen & Revsbech 1989). We used this approach in a Favia sp. coral and found the coral respiration under saturating light conditions to be 6 times higher than the dark respiration (Table 1). A higher coral respiration in light has been postulated from observations of a post-illumination enhancement of respiration in corals (Edmunds & Davies 1988), a higher dark respiration in light-adapted vs shade-adapted corals (Porter et al. 1984), and from measurements of higher coral dark respiration in hypoxic water (Shick 1990). To our knowledge, we present here the first data of daytime respiration measured in illuminated corals.

The high O₂ concentration and low CO₂ concentration (indicated by the high pH) found in the illuminated coral tissue create favorable conditions for photorespiration, i.e. the O₂ consumption caused by a detoxification of glycylate produced by the oxygenase activity of the enzyme ribulose biphosphate carboxylase (Ogren 1984, Beardall 1989). Although the presence of a functional photorespiratory pathway in zooxanthellae has been demonstrated, the significance of photorespiration in zooxanthellae is still largely unknown (Muscatine 1990). At present we cannot therefore rule out that part of the enhanced O₂ consumption in light is due to photorespiration. Microsensor measurements in corals under experimentally controlled O₂/CO₂ ratios similar to the experiments described by Glud et al. (1992) could probably give an estimate of the importance of photorespiration in corals.

Another mechanism for light-enhanced O₂ consumption could be a true stimulation of the cell metabolism of zooxanthellae, the host or both, e.g. by organic compounds freshly translocated from the zooxanthellae to the host tissue during photosynthesis (Edmunds & Davis 1988). Such a mechanism for light-enhanced respiration has been demonstrated in phytoplankton (Falkowski et al. 1985).

Yet another mechanism causing light-enhanced O₂ consumption in corals was demonstrated by Shick (1990), who showed that the dark respiration of corals and other anthozoans was limited by the diffusive supply of O₂. This was demonstrated by an enhanced dark respiration when corals were incubated in hypoxic water. For the branched coral Stylophora pistillata up to 20% higher dark respiration was observed in hypoxic water relative to the respiration in air-saturated water. The diffusion limitation of dark respiration is caused by the presence of a DBL (Patterson 1992, Shashar et al. 1993, Figs. 2 & 3 in this study). By using hypoxic water in the dark incubations, the effect of the DBL on O₂ supply was alleviated and respiration thus no longer O₂ limited. A similar alleviation of the diffusion limitation of O₂ was postulated to occur at daytime, due to the internal O₂ production by zooxanthellae photosynthesis. Our data confirm this hypothesis by showing the presence of hyperoxic conditions in the coral tissue, which lead to a net efflux of O₂ to the overlying water in light (Fig. 2).

The observed stimulation of coral respiration in the light, most probably by a combination of the above-mentioned mechanisms, has profound consequences for the calculation of energy and carbon budgets in corals. Our data indicate a potential daytime respiration 6 times higher than the dark respiration (Table 1). Edmunds & Davies (1988) and Shick (1990) report a 10 to 60% higher dark respiration either after previous illumination or under hypoxic conditions. Under the traditional assumption of equal respiration in the dark and the light, daytime respiration is consequently underestimated and will result in a corresponding underestimation of gross photosynthesis. The quantification of daytime respiration thus seems crucial for future studies of carbon and energy budgets in corals.

**Light and photosynthesis**

Apart from the chemical microenvironment, which is highly dependent on diffusional exchange processes, the key parameters for photosynthesis regulation are the intensity and spectral composition of the light field surrounding the zooxanthellae. The light field available for the zooxanthellae in the tissue differed strongly from the incident irradiance due to the combined action of multiple scattering and absorption, resulting in a wavelength-dependent local maximum of the spherically integrated quantum flux (i.e. scalar irradiance) in the upper tissue layers (Fig. 6). Similar results have been found in studies of the light field in
plant tissue, sediments and microbial mats (e.g. Vogelmann & Björn 1986, Jørgensen & Des Marais 1988, Kühl & Jørgensen 1992, Kühl et al. 1994a, b). The mechanism behind the observed local maximum of scalar irradiance relative to incident irradiance can be explained in terms of multiple scattering and diffuse reflection of light within the coral tissue and from the surrounding coral skeleton, which is not taken into account in the measurement of incident downwelling irradiance. Diffuse reflection from the coral skeleton is definitely an important component of the scalar irradiance maximum. The light field exhibited horizontal heterogeneity at the level of a single coral polyp and the highest values of scalar irradiance were found on ridges of coral skeleton covered with the thinnest tissue layers (Fig. 6). Furthermore, if the refractive index of the coral tissue is higher than that of water, diffuse backscattered light can be internally reflected at the tissue-water interface, where collimated light is continuously incident. Light can thus enter more easily into the tissue from the water than it can escape back into the water. The significance of such an internal reflection mechanism for the observed scalar irradiance maximum has been demonstrated in animal tissue by Anderson et al. (1989).

Multiple scattering increases the pathlength of photons per vertical distance traversed. The increased pathlength due to multiple scattering is thus equivalent to a longer residence time of photons within a given layer of the optically dense coral tissue, which results in a higher photon flux density close to the tissue surface where collimated light is continuously incident. Light can thus enter more easily into the tissue from the water than it can escape back into the water. The significance of such an internal reflection mechanism for the observed scalar irradiance maximum has been demonstrated in animal tissue by Anderson et al. (1989).

Multiple scattering increases the pathlength of photons per vertical distance traversed. The increased pathlength due to multiple scattering is thus equivalent to a longer residence time of photons within a given layer of the optically dense coral tissue, which results in a higher photon flux density close to the tissue surface where collimated light is continuously incident. The amplification effect of the increased pathlength on the scalar irradiance at the tissue surface will be most pronounced at wavelengths outside the major absorption regions of coral photopigments, e.g. infrared light, while in the spectral regions of maximal photopigment absorption the increased pathlength due to multiple scattering leads to more efficient light absorption. Scattering thus also leads to an amplification of the spectral differences between regions of low and high absorption, respectively. A more detailed discussion of the optical mechanisms behind the observed scalar irradiance maximum is beyond the scope of this study and can be found elsewhere (e.g. Vogelmann & Björn 1986, Anderson et al. 1989, Kühl & Jørgensen 1994).

The properties of the microscale light field experienced by the zooxanthellae must be taken into account in studies of their photophysiology. The zooxanthellae live in a strongly scattering environment and as such the relevant light intensity measure for the symbiont photosynthesis is scalar irradiance rather than incident irradiance (Weinberg 1976, Kirk 1983). We measured the action spectrum of zooxanthellae photosynthesis in Acropora sp. as a function of the incident collimated light, i.e. the downwelling irradiance (Fig. 8). The zooxanthellae do, however, also receive a significant amount of scattered light from their surroundings and a true action spectrum of the microalgae is thus related to the scalar irradiance (Jørgensen et al. 1987). The contribution of scattered light, which was not taken into account in the recalculation of photosynthetic rates to the same incident light intensity ('Material and methods'), will be wavelength dependent (Fig. 6) thus leading to relatively more scattered light at wavelengths outside the major absorption regions of the coral photopigments. The action spectrum calculated with the downwelling irradiance as the light intensity measure does not take this effect into account and will thus be distorted. This might be the explanation for the relative low photosynthetic activity in the blue region of chl a absorption in the action spectrum presented here (solid curve in Fig. 8). In order to estimate the degree of distortion we recalculated the action spectra using a scalar irradiance spectrum obtained in a Favia sp. (dotted curve in Fig. 8). The recalculated action spectrum shows a much higher photosynthetic activity in the blue part of the spectrum and is similar to action spectra presented for isolated zooxanthellae (Halldal 1968) and planktonic dinoflagellates (e.g. Prézelin et al. 1976).

The P vs I curve and derived parameters of zooxanthellae photosynthesis also depend on the measure of light intensity. If photosynthesis in Favia sp. was plotted against scalar irradiance $E_d$ instead of downwelling irradiance $E_o$, the $P$ vs $E_d$ curve (dotted curve in Fig. 10) exhibited a lower initial slope than the $P$ vs $E_o$ curve (solid curve in Fig. 10). The maximum photosynthetic rate is independent of the choice of light intensity parameter, and thus the $P$ vs $E_d$ curve exhibited a higher onset of light saturation ($I_k = 273 \mu$Ein m$^{-2}$ s$^{-1}$) than the $P$ vs $E_o$ curve ($I_k = 152 \mu$Ein m$^{-2}$ s$^{-1}$). This effect of scalar irradiance on the parameters of photosynthetic performance is probably an overestimate due to the anticipation of a scalar irradiance of 180% of incident irradiance throughout the visible spectrum. Nevertheless, our data indicate that $P$ vs $E_o$ curves measured in photosynthetic communities where light scattering is significant exhibit higher initial slopes and thus result in lower $I_k$-values for photosynthesis than equivalent calculations based on $P$ vs $E_d$ curves. Preferentially, studies of the photosynthetic performance of zooxanthellae and e.g. photosynthetic microorganisms in sediments and biofilms should thus take scattered light into account by measurements of scalar irradiance.

Although the coral tissue was supersaturated with $O_2$ in light, the $P$ vs $I$ curves in both Acropora sp. and Favia sp. exhibited no indications of photoinhibition at
high light intensities (Figs. 9 & 10). High levels of the protective enzymes superoxide dismutase, catalase, and ascorbate peroxidase are present in coral tissue (Lesser & Shick 1989, Shashar & Stambler 1992). An efficient protection against toxic effects of high O₂ concentrations in the tissue (i.e. photooxidation) by the enzymes mentioned is probably a major mechanism behind the absence of photoinhibition often observed in shallow water corals and associated algal communities (Carpenter 1985, Falkowski et al. 1990). Another important mechanism is the presence of high levels of protective pigments, especially UV-absorbing compounds in corals, which protect against both direct UV damage and synergistic effects of UV and high O₂ in the coral (e.g. Shibata 1969, Falkowski et al. 1990, Drollet et al. 1993, Kinzie 1993).

The previously mentioned variation of the DBL (Fig. 3) (Shashar et al. 1993) and the observed differences in the light field within single polyps (Fig. 6) may lead to a spatial heterogeneity of photosynthesis and respiration. The spatial variation of e.g. photosynthetic activity within a single polyp (Fig. 7) might, however, also result from differences in zooxanthellae densities in various parts of the polyp. Thus a direct comparison of microsensor measurements with whole-colony respiration and photosynthesis obtained by the O₂ exchange techniques would therefore require a more detailed mapping of the DBL and the light field together with measurements of photosynthesis, respiration and zooxanthellae densities under defined illumination and hydrodynamic conditions.

In conclusion, we found the regulation of coral physiology to be strongly dependent on the gradient microenvironment of the zooxanthellae and the polyp tissue. The chemical microenvironment results from combined host and symbiont metabolism and diffusional transport. These are regulated by a flow-dependent DBL. In the dark, coral respiration is limited by the diffusive supply of O₂ from the surrounding water through the DBL, while in the light the zooxanthellae photosynthesis results in hyperoxic conditions in the coral tissue and thus provides an internal supply of O₂ for coral respiration. Coral respiration in the light is much higher than dark respiration, although photorespiration might account for a part of the observed difference. The photosynthetic performance of the zooxanthellae is highly dependent on the coral tissue light field. Scattered light is important for the zooxanthellae and results in a higher light intensity and a different spectral composition of light in the coral tissue than expected from the incident irradiance. Scalar irradiance is thus the most relevant light intensity parameter in relation to zooxanthellae photosynthesis.

We found microsensors to be ideal tools for studies of the microenvironmental controls of coral physiology. Many of the experimental techniques traditionally used in studies of coral photosynthesis and respiration rely on assumptions about the microenvironment of the coral tissue, which can now be tested by direct measurements with microsensors.

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duction of microalgae in sediments measured by oxygen microprofile, $^{14}\text{C}O_2$ fixation, and oxygen exchange methods. Limnol. Oceanogr. 717–730

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