Relationships between fertilization of the Southeastern Pacific sea urchin *Arbacia spatuligera* and environmental variables in polluted coastal waters

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ABSTRACT: Fertilization bioassays with the sea urchin *Arbacia spatuligera* were used to assess the water quality of 9 marine coastal sectors in the Biobío Region (Southern Chile) and to investigate the relationship between fertilization and environmental variables (*in situ* pH, turbidity, dissolved oxygen and salinity), heavy metals (copper, cadmium and mercury) and organic compounds (oil and grease, lindane, aldrin, dieldrin, DDE, DDT and phenols). The fertilization (mean ± standard deviation) of *A. spatuligera* eggs in sectors with high organic discharges from fishmeal industries such as Rocuant (14.1 ± 6.0%) and San Vicente (22.5 ± 10.0%) was significantly lower (*p* < 0.05) than in the other sectors (Chivilingo, Coronel, Lenga, Talcahuano, Penco, Tomé, Dichato) (70.5 ± 13.0%). Percent saturation of oxygen (*r* = 0.839), pH (*r* = 0.830), turbidity (*r* = -0.683) and oil and grease (*r* = -0.888) showed significant correlations (*p* < 0.001) with fertilization. Principal Component Analysis (PCA) using standardized variables grouped fertilization with dissolved oxygen and pH (total variance of 2 first components explained 73.0%) and Linear Multiple Regression Analysis produced the following model: Fertilization = 30.001 + 0.785(Percent saturation of oxygen) - 0.059(Oil and grease) (*r*² = 0.837; *p* < 0.001). Rocuant and San Vicente stations were grouped and separated from the rest by inverse PCA and then discriminated by Discriminant Analysis (Hotelling-Lawley Trace, *p* < 0.001). It was concluded in laboratory bioassays that waters of coastal sectors from Rocuant and San Vicente, with the highest levels of oil and grease and lowest oxygen concentrations, had a significant effect on the fertilization of *A. spatuligera* eggs.

KEY WORDS: Environmental assessment, Bioassays, Sea urchin fertilization, Organic pollution

INTRODUCTION

Organic pollution (i.e., eutrophication) of marine coastal waters is becoming a widespread problem throughout the world (Rosenberg 1985, Crema et al. 1991). Commonly, this phenomenon leads to high levels of organic matter in sediments, with subsequent occurrences of hypoxia or anoxia both in shallow and in bottom waters (Cederwall & Elmgren 1990). It has been found that these altered environmental conditions may cause adverse biological effects, such as the mass mortality of benthic organisms (Elmgren 1989, Baden et al. 1990, Rosenberg 1990, Sandberg 1994).

Several sectors within a large coastal area of the Biobío Region (36°40' S, 73°10' W) of southern Chile are characterized by low oxygen concentrations and large amounts of organic matter in their sediments (Ahumada & Rudolph 1989). This region, including the Gulf of Arauco and Bays of Coliumo, San Vicente and Concepción (Fig 1), consists of semi-enclosed shallow areas with depths of about 20 m and very low water exchange with adjacent seawater (Ahumada et al., 1989a). Discharges from industries, coupled with active urban development, produce point and non-
point sources of marine pollution in the region (Chuecas 1989). In particular, Concepcion and San Vicente Bays have been identified as the most polluted coastal zones in Chile as a consequence of port activities and raw sewage discharges of domestic and industrial origin.

Rudolph et al. (1984) have shown that organic pollutants from domestic sewage from the city of Talcahuano and the effluent discharges from fishmeal factories on Rocuant Island produce strong coastal pollution in Concepcion Bay. Likewise, San Vicente Bay receives untreated domestic sewage and industrial discharges from fishmeal and steel mills, the liquid discharges of industrial fishing ships and untreated effluents from several chemical industries in Lenga. Various authors have reported that the severe pollution of this bay (Gallardo 1984, Ahumada et al. 1989a, Chuecas 1989, Encina 1993) causes important ecological effects, mainly benthic ecosystem alterations resulting from anoxic conditions.

Potential sources of pollution in the Gulf of Arauco include organic compounds from paper mill effluents, agricultural runoff of organochlorine pesticides through the Biobio river and release of sewage from fishmeal factories in Coronel. This port has been severely affected by high contents of organic matter in its sediments (Ahumada & Rudolph 1989).

In spite of the intensive activities in the coastal areas within the Biobio Region, there are no organized environmental monitoring efforts and few studies have been conducted to determine the causes and effects of the pollution within this coastal zone (Chuecas 1989).

Methods to assess effects of conditions related to organic pollution on marine organisms have not yet been fully explored and little is known about the relationships between indicative parameters of organic pollution (e.g. dissolved oxygen, pH, hydrogen sulphide and organic matter), and their biological effects. Rapid and accurate laboratory toxicity tests may be used to assess biological effects of single or combined factors related to organic pollution, monitor water quality, and to predict the potential harmful effects of contaminated environments (Richardson & Martin 1994).

Several authors have reported the effects of single variables associated with organic pollution (e.g. hypoxia, organic enrichment) on the benthic environment (Rosenberg et al. 1991, Baden et al. 1994, Sandberg 1994, Van den Thillart et al. 1994), but studies integrating effects of multiple variables are desirable. Correlations between toxicity test results and multiple variables have only been established for sediments. Swartz et al. (1984) have shown that more than one variable from organically enriched sediments affects the survival of the amphipod *Rhepoxynius abronius*. Likewise, Meador et al. (1990) have shown that several sediment contaminants are related to embryo abnormalities in the sand dollar *Dendraster excentricus*.

The main goal of this study was to detect spatial differences in water quality from polluted and non-polluted coastal areas within the Biobio Region (ca 120 km of coastline) using a standardized short-term sperm toxicity test (i.e. fertilization bioassay) with the sea urchin *Arbacia spatuligera*. We also wished to determine the relationships between fertilization success of sea urchins and environmental factors (dissolved oxygen, salinity, pH, turbidity), heavy metals (copper, cadmium and mercury) and organic compounds (oil and grease, lindane, aldrin, dieldrin, DDT [1,1-dichloro-2,2-bis(chlorophenyl)ethylene], DDE [1,1,1-trichloro-2,2-bis(p-chlorophenyl)ethane] and phenols).

**MATERIALS AND METHODS**

**Sampling.** Four seawater samples of 200 ml were collected by hand in the low intertidal zone at 17 coastal stations within the Biobio Region (Fig. 1). The samples were transported to the laboratory and used for bioassays within 24 h. At each station, temperature (°C), salinity (%), pH, turbidity (nephelometric units, NTU) and oxygen concentration (mg l⁻¹) were measured *in situ* with a Horiba Water Quality Analyzer.
Dissolved oxygen was transformed to percent saturation of oxygen according to Weiss (1970).

Water samples for copper and cadmium analyses were placed in 1 l capped plastic bottles and stored at 4°C. Samples for mercury analysis were placed in 0.5 l capped glass bottles, acidified with concentrated nitric acid and then stored at 4°C. Samples for organochlorine pesticides (lindane, aldrin, dieldrin, DDE, DDT), phenols, and oil and grease were taken in 2.5 l brown glass bottles and stored at 4°C prior to analysis.

Heavy metal analyses. Methods for heavy metal analyses are described in Capelli et al. (1983). To determine copper and cadmium concentrations, seawater samples were filtered with 0.45 μm Millipore filters. A 300 ml volume of the filtrated sample was extracted by mechanical shaking over the course of 3 d with 0.1500 g of Chelex-100 resin. The water was drained and the resin dried at 35°C. The ions were then liberated from the resin by adding 2 ml of 1 N HNO₃. After 1 h 8.0 ml of distilled water was added, the solution homogenized and the resin discarded. Finally, the samples were analyzed on a Perkin Elmer Atomic Absorption Spectrophotometer (AAS) with graphite furnace. Mercury analyses were taken directly by AAS coupled to a dragging cold vapor system with stannous chloride (MHS-20).

Organic compound analyses. Methods for organic compound analyses are described in UNEP/IAEA (1982) and UNEP/IOC/IAEA (1986). The samples for organochlorine pesticides, phenols and oil and grease analyses were acidified to a pH of 4.0 with concentrated H₂SO₄ and then extracted 3 times with 10 ml of n-hexane (Merck p.a.). The extracts were mixed and dried with anhydrous Na₂SO₄ and evaporated to 5 ml in a rotary evaporator. Excess hydrogen sulphide was eliminated with copper and excess grease with concentrated H₂SO₄. Finally, phenol concentrations were estimated with a Shimadzu UV-190 spectrophotometer at 273 nm. Pesticides were analyzed on a Varian 2700 gas chromatograph with electron capture detection (60Ni) with N₂ as the carrier gas. Quantification of pesticides was accomplished by comparing peak heights for the sample with those produced by reference standards. For the analysis of oil and grease an aliquot of the extracted sample was dried to a constant weight. Two ml of n-hexane and 1 ml of concentrated H₂SO₄ were then added to eliminate greases. The acid phase was discarded and the organic phase was dried once again to a constant weight. The difference between the 2 weights corresponded to oil and grease in the sample.

Bioassay procedures. The methods for sperm toxicity tests were described by Weber et al. (1989) and modified by Zúñiga et al. (1995). Arbacia spatuligera adults were collected on November 1993 by SCUBA divers in Coliumo Bay (36°32'S, 72°56'W) and transported to the Marine Biology Station of Dichato, where they were acclimated in aerated flowing seawater for 1 wk. Seawater samples, collected on November 1993, were tested undiluted (i.e. as ambient seawater) with 4 replicates per station in a randomized experimental design. In addition, a reference toxicant test (serial dilution of CuSO₄ ranging from 100 to 3.125 μg l⁻¹ of Cu²⁺) was conducted in conjunction with each series of experiments to document the performance of the bioassays. The seawater used for test controls and reference toxicant dilutions was obtained near the sea urchin collection site at Dichato. Gametes were obtained by injecting 1 ml of 0.5 M KC1 into the perivisceral fluid of 10 individuals. Undiluted sperm from 5 males was pooled and kept on ice. The eggs of 5 females were collected with a pipette, pooled in 250 ml of seawater and held at room temperature before use in the experiments. The exposure vessels were test tubes containing 5 ml of the seawater or reference toxicant sample. The newly collected sperm was diluted to 5 x 10⁷ cells ml⁻¹ with control seawater. A 0.1 ml volume of this diluted sperm suspension was added to the seawater in each test tube and exposed for 1 h. Approximately 2000 eggs were added to the test tube, following the sperm cell exposure period, and held for an additional 15 min to allow fertilization to occur. Finally, fertilization was stopped by the addition of 1 ml of a concentrated formaldehyde solution. Temperature during conduction of bioassays was ambient (16.5°C). The percentage of fertilized eggs was determined in each replicate by microscopic examination of 100 eggs for the presence or absence of a fertilization membrane.

An additional seawater sampling was made to validate the assumption that variations in physicochemical parameters between in situ and laboratory experimental conditions do not affect the results. Samples, handled in the same manner as test samples, were used to measure the physico-chemical parameters before and after experiments. Variations (mean ± standard deviation) between in situ and experimental measures were 10.0 ± 6.4% for percent saturation of oxygen, 0.09 ± 0.09 for pH, 0.6 ± 0.4% for salinity, 0.8 ± 0.6°C for temperature and 12 ± 1 NTU for turbidity.

Data analysis. Toxstat (Gulley et al. 1988) and Systat (Wilkinson 1990) programs were used for statistical analyses. Parametric analyses were validated by testing assumptions of normality and homogeneity of variances of the error terms using the arcsine (square root) transformation of fertilization data. Analysis of variance was followed by Dunnett's test in order to detect the differences between the control and treatments. The Spearman-Karber program (Hamilton et al. 1977) was used to calculate the 50% effective concentration (EC₅₀) for the reference toxicant test. Relationships
between environmental variables and laboratory fertilization bioassays were validated by simple regression analyses. Principal component analysis (PCA) was used with standardized variables to detect grouping between environmental variables and fertilization bioassays. Inverse PCA and discriminant analysis (DA) were used to detect groups of stations with different fertilization success. Stepwise Linear Multiple Regression Analysis was used to find the relationship between fertilization and environmental variables and organic compounds.

Control samples showed the highest fertilization success (88.8 ± 5.8%), indicating that sea urchin gametes were in good physiological conditions for bioassays. The EC_{50} obtained with the copper standard was 8.21 μg l^{-1} with 95% confidence intervals between 6.91 and 9.74 μg l^{-1}.

Mean fertilizations in sperm toxicity tests for seawater samples from the different sectors are presented in Fig. 2. Stations at Rocuant (14.1 ± 6.0%) and San Vicente (22.5 ± 10.0%), with the lowest fertilizations, were significantly different from stations with intermediate and high fertilizations, represented by Dichato, Tomé, Penco, Talcahuano, Lenga, Coronel and Chivilingo (70.5 ± 13.0%) (Dunnett's test, p < 0.05).

Mean values (± standard deviation) of environmental variables by sector are shown in Table 1. Mean salinity values showed little variability between sectors (33.3 ± 2.0%), well within the normal range for survival and reproduction of the sea urchins (Stickle & Diehl 1987). Temperature also shows low variability between sectors (15.0 ± 1.6°C). Fertilization did not show any relationship with salinity.

Mean oxygen concentrations from Rocuant (0.49 ± 0.62 mg l^{-1}; 6.3 ± 6.8% saturation), San Vicente (1.24 ± 1.06 mg l^{-1}; 14.9 ± 10.8% saturation) and Talcahuano (1.59 ± 1.39 mg l^{-1}; 19.5 ± 12.8% saturation) were significantly lower than in the rest of sectors combined (4.56 ± 0.88 mg l^{-1}; 54.9 ± 9.9% saturation; Dunnett's test, p < 0.05; Table 1). The poor environmental conditions of Rocuant, San Vicente and Talcahuano were correlated with low oxygen concentrations. Fertilization was positively correlated with in situ oxygen concentration (r = 0.839; p < 0.001; Fig. 3a). Most sectors with low oxygen concentrations (<2.5 mg l^{-1}; 30.1% saturation) had fertilization success below 40%, while water samples with oxygen concentrations between 3.0 and 5.8 mg l^{-1} (i.e. between 36 and 70% saturation) had fertilizations above 50% (Fig. 3a).

Mean pH from Rocuant (7.16 ± 0.04) and San Vicente (7.22 ± 0.07) were significantly lower than in the rest of
sectors (7.59 ± 0.24; p < 0.01; Table 1). Fertilization was positively correlated with in situ pH (r = 0.830, p < 0.001; Fig. 3b). Water samples with pH values below 7.3 showed fertilization success below 35%; those with higher pH values showed increased fertilization success (Fig. 3b).

Mean turbidity in Rocuant (156 ± 80 NTU), Penco (125 NTU), Coronel (81 ± 4 NTU) and San Vicente (94 ± 30 NTU) were significantly higher than in the rest of sectors (25 ± 17 NTU; p < 0.05; Table 1). Fertilization was negatively correlated with in situ turbidity (r = -0.683, p < 0.001; Fig. 3c). Most water samples with turbidity values above 50 NTU showed fertilization success lower than 30%, while those with turbidity below 50 NTU had higher fertilization success (Fig. 3c).

**Fertilization vs heavy metals**

Heavy metal concentrations for the different sectors within the Biobio Region are shown in Table 2. In general, the 3 metals showed no significant variations between sectors. Mean seawater concentrations of copper (0.61 ± 0.47 μg l⁻¹) were similar to unpolluted seawater concentrations (0.64 μg l⁻¹) and below US EPA (1976, 1982) water quality criteria (10 to 43 μg l⁻¹). Concentrations of cadmium (0.05 ± 0.03 μg l⁻¹) and mercury (0.05 ± 0.04 μg l⁻¹) were slightly above unpolluted seawaters.

<table>
<thead>
<tr>
<th>Sampling sector</th>
<th>Copper</th>
<th>Cadmium</th>
<th>Mercury</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dichato</td>
<td>0.16</td>
<td>&lt;0.02</td>
<td>0.03</td>
</tr>
<tr>
<td>Tomé</td>
<td>1.41</td>
<td>0.08</td>
<td>&lt;0.02</td>
</tr>
<tr>
<td>Penco</td>
<td>0.69</td>
<td>&lt;0.02</td>
<td>0.03</td>
</tr>
<tr>
<td>Rocuant</td>
<td>0.38</td>
<td>0.06</td>
<td>0.09</td>
</tr>
<tr>
<td>Talcahuano</td>
<td>0.51</td>
<td>0.05</td>
<td>0.03</td>
</tr>
<tr>
<td>San Vicente</td>
<td>0.60</td>
<td>0.02</td>
<td>0.04</td>
</tr>
<tr>
<td>Lenga</td>
<td>0.51</td>
<td>&lt;0.02</td>
<td>0.03</td>
</tr>
<tr>
<td>Coronel</td>
<td>0.36</td>
<td>0.03</td>
<td>0.03</td>
</tr>
<tr>
<td>Chivilingo</td>
<td>1.09</td>
<td>0.10</td>
<td>0.03</td>
</tr>
<tr>
<td>Unpolluted seawaters*</td>
<td>0.64</td>
<td>0.025</td>
<td>0.012</td>
</tr>
<tr>
<td>Water quality criteria**</td>
<td>10–43</td>
<td>5–10</td>
<td>0.1–1.0</td>
</tr>
</tbody>
</table>

*aFurness & Rainbow (1990)
**US EPA (1976, 1982) and Kobylinski et al. (1993)
luted seawater concentrations (0.025 μg l⁻¹ for Cd and 0.012 μg l⁻¹ for Hg) but were also below US EPA (1976, 1982) water quality criteria (5 to 10 μg l⁻¹ for Cd and 1 to 10 μg l⁻¹ for Hg). No significant correlations were found between fertilization success and copper, cadmium or mercury concentrations.

**Fertilization vs organic compounds**

The concentrations of organic compounds within the Biobio Region are shown in Table 3. Seawater phenol concentrations varied between 0.01 and 0.745 μg l⁻¹ with the highest values in San Vicente (0.745 μg l⁻¹) and Rocuant (0.126 μg l⁻¹), both below US EPA (1982) water quality criteria (1.0 μg l⁻¹).

In some sectors organochlorine pesticide concentrations fell below detectable levels (<0.001 μg l⁻¹). The highest concentrations of lindane were found in San Vicente (0.060 μg l⁻¹), Rocuant (0.046 μg l⁻¹) and Talcahuano (0.036 μg l⁻¹). These concentrations are above both unpolluted (Antarctic) seawaters (0.126 ng l⁻¹) and the US EPA (1982) water quality criteria (0.004 μg l⁻¹). The highest concentrations of aldrin-dieldrin were detected in Rocuant (0.180 μg l⁻¹), San Vicente (0.148 μg l⁻¹) and Talcahuano (0.148 μg l⁻¹). These concentrations are also greater than both unpolluted (Antarctic) seawaters (0.086 ng l⁻¹) and US EPA (1982) water quality criteria (0.006 μg l⁻¹). Water concentrations of DDT-DDE in Talcahuano (0.061 μg l⁻¹), San Vicente (0.057 μg l⁻¹), Coronel (0.030 μg l⁻¹) and Rocuant (0.024 μg l⁻¹) were many times above unpolluted waters (0.42 ng l⁻¹) and US EPA (1982) water quality criteria (0.002 μg l⁻¹). However, no significant correlations were found between fertilization and phenols and organochlorine pesticides.

Oil and grease concentrations show high variability between stations, with values in Rocuant (260.3 ± 148.7 mg l⁻¹) and San Vicente (240.1 ± 396.7 mg l⁻¹) significantly higher than in the remaining sectors considered together (1.1 ± 1.6 mg l⁻¹) [data transformed to ln(x+1); Dunnett's test, p < 0.05; Table 3]. Concentrations of oil and grease both in Rocuant and San Vicente were above US EPA (1982) water quality criteria (25.0 mg l⁻¹). Fertilization was negatively correlated with oil and grease (r = −0.888, p < 0.001; Fig. 3d). Most water samples with oil and grease concentrations above 5 mg l⁻¹ showed fertilizations lower than 45%, while those with concentrations below 5 mg l⁻¹ had higher fertilizations (Fig. 3d).

**Multivariate analyses**

PCA applied over environmental factors, heavy metals and organic contaminants showed that the first 2 principal components explained 73% of the total variance. Principal Component 1 was explained mainly by fertilization (r = −0.947). This analysis showed that fertilization was positively correlated with variables located to the right (dissolved oxygen and pH) and negatively correlated with variables located to the left (turbidity, aldrin, total organochlorine pesticides, phenols and oil and grease) (Fig. 4).

When a Stepwise Linear Multiple Regression Analysis was applied, considering fertilization as the dependent variable, the following model was obtained: Fertilization = 30.001 + 0.785(Percent saturation of oxygen) + 0.059(Oil and grease) (r² = 0.837, p < 0.001). The high multiple regression correlation coefficient indicates that fertilization is influenced more by seawater oxygen and by oil and grease concentrations than by the other variables examined. The first 2 principal components from inverse PCA explained 89% of total variance as shown in Fig. 5. Component axis 2 was well correlated with fertilization (r = −0.873). Stations located at the top of axis 2 (Rocuant and San Vicente) share very low fertilizations. Discriminant Analysis (Fig 6) confirmed that stations at Rocuant and San Vicente were significantly different from the others (Hotelling-Lawley Trace, p < 0.001), locating stations with lowest fertilization success to the right of the graph (i.e. Rocuant and San Vicente). Factor 1 was well correlated with component axis 2 (r = 0.894) and fertilization (r = −0.996).

Table 3. Mean seawater concentrations of pesticides (μg l⁻¹), phenols (μg l⁻¹) and oil and grease (mg l⁻¹) in the Biobio Region.

<table>
<thead>
<tr>
<th>Sampling sector</th>
<th>Phenols</th>
<th>Lindane</th>
<th>Aldrin + Dieldrin</th>
<th>DDT + DDE</th>
<th>Oil and grease</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dichato</td>
<td>0.010</td>
<td>0.006</td>
<td>0.004</td>
<td>0.002</td>
<td>0.0</td>
</tr>
<tr>
<td>Tomé</td>
<td>0.030</td>
<td>0.008</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>0.0</td>
</tr>
<tr>
<td>Penco</td>
<td>0.030</td>
<td>0.006</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>0.0</td>
</tr>
<tr>
<td>Rocuant</td>
<td>0.126</td>
<td>0.046</td>
<td>0.180</td>
<td>0.024</td>
<td>260.3</td>
</tr>
<tr>
<td>Talcahuano</td>
<td>0.028</td>
<td>0.036</td>
<td>0.148</td>
<td>0.061</td>
<td>2.8</td>
</tr>
<tr>
<td>San Vicente</td>
<td>0.745</td>
<td>0.060</td>
<td>0.148</td>
<td>0.037</td>
<td>240.1</td>
</tr>
<tr>
<td>Lenga</td>
<td>0.010</td>
<td>0.002</td>
<td>0.006</td>
<td>0.002</td>
<td>0.0</td>
</tr>
<tr>
<td>Coronel</td>
<td>0.098</td>
<td>&lt;0.001</td>
<td>0.063</td>
<td>0.039</td>
<td>1.2</td>
</tr>
<tr>
<td>Chilquingo</td>
<td>0.020</td>
<td>&lt;0.001</td>
<td>0.006</td>
<td>&lt;0.001</td>
<td>0.9</td>
</tr>
<tr>
<td>Unpolluted seawaters*</td>
<td>-</td>
<td>1.26×10⁻¹</td>
<td>0.86×10⁻¹</td>
<td>4.24×10⁻¹</td>
<td>-</td>
</tr>
<tr>
<td>Water quality criteria*</td>
<td>1.000</td>
<td>0.004</td>
<td>0.006</td>
<td>0.002</td>
<td>25.0</td>
</tr>
</tbody>
</table>

*Becerra et al. (1992); *US EPA (1982)
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**DISCUSSION**

**Bioassay assessment**

In the last decade, sea urchin sperm toxicity tests (i.e. fertilization bioassays) have been used to detect differences in seawater quality and to give quantitative information about unpolluted and polluted areas (Dinnel et al. 1988, Kobayashi 1991). In an extensive water quality assessment, we tested differences in fertilization success of the sea urchin *Arbacia spatuligera* between several unpolluted and polluted coastal waters within the Biobio Region and focused on detection of the environmental factors and contaminants significantly related to these differences. Fertilization in sea urchins includes sperm activation following contact with seawater after its release from the gonads. Many physiological events, including movement of the sperm flagellum, may be affected by poor environmental conditions or the presence of chemical pollutants during toxicity tests (Dinnel et al. 1989). For example, metabolic events during the activation of the sperm may be affected by changes in dissolved oxygen and pH (Giudice 1986).

In a previous report Zúñiga et al. (1995) found that water samples from Rocuant Island and San Vicente significantly decreased fertilization success in *Arbacia spatuligera*. Our results from water quality assessment in a more extensive area within the Biobio Region confirm that the Rocuant and San Vicente sectors produced the lowest percent fertilization in the sea urchin bioassays. When fertilization was related to environmental factors, it was found that fertilization success was positively correlated with dissolved oxygen and pH and negatively correlated with turbidity. Therefore, the poorest seawater quality situations in coastal environments, as reflected by the lowest fertilizations in *A. spatuligera*, were associated with low oxygen levels, low pH and high turbidity. We found that anoxic conditions are present in sectors with the highest inputs of organic matter and that oxygen concentration is directly related to pH \( r^2 = 0.741, p < 0.001 \). One explanation for this relation is that aerobic and anaerobic metabolism of high contents of organic matter consume dissolved oxygen and produce high levels of carbon dioxide and hydrogen sulfide that finally reduce pH (Riley & Chester 1971, Aubert 1990).
Relationships between fertilization and environmental variables

It has been suggested that gametes and embryos are the stages in the life cycle of marine invertebrates which are most sensitive to anoxia and hypoxia (Lutz et al. 1994). Rosenberg et al. (1991) have reported that many marine benthic species are very sensitive to low oxygen concentrations, with a lower tolerance limit of 1 ml l⁻¹ (ca 1.43 mg l⁻¹). Accordingly, our results showed that in situ oxygen concentrations of 1 to 2 mg l⁻¹ reduced fertilization success in Arbacia spatuligera to below 50% (between 46 and 26% in the linear model).

The relationship between fertilization success and in situ pH indicated that seawater samples with a pH of 7.1 to 7.3 reduced fertilization success to below 40% (in the linear model between 36 and 17%). These low fertilization levels might be due to the inactivation of sperm caused by low pH. Christen et al. (1983) pointed out that pH variations affected the motility of sperm, suggesting that low internal pH (7.0) prevents motility of sperm within the gonad, by inhibiting the dynein ATP-ase, which is rate-limiting for the respiration of tightly coupled mitochondria. Alkalization of the internal pH (8.0) acts as an activator of the ATP-ase, increasing sperm respiration rate and motility.

Turbidity has been used as a parameter indicative of suspended particulates and it has been established that such particulates may adsorb and transport high levels of organic pollutants (e.g. organochlorine pesticides and polycyclic aromatic hydrocarbons; Boulo-bassi & Saliot 1991). Water samples from polluted sectors within the Biobio Region showed high levels of suspended particulates (i.e. turbidity) which are associated with high concentrations of organochlorine pesticides (r = 0.814). This could contribute to the significant reduction of fertilization success in the laboratory bioassays.

Additionally, low oxygen concentrations, low pH and high levels of suspended solids might induce additive or synergistic negative effects on physiological processes during fertilization of Arbacia spatuligera in seawater.

Relationships between fertilization and inorganic and organic contaminants

In most stations, concentrations of copper, mercury and cadmium were near those of unpolluted seawaters or below US EPA (1976, 1982) water quality criteria. This could explain the lack of correlation between heavy metals and fertilization success in Arbacia spatuligera. Potential sources of heavy metal within the region are scarce. The main discharges originate in steel mills in San Vicente (Gallardo 1984). Additionally, the area might be subjected to aerial deposition of cadmium and mercury from fossil fuel combustion, as shown by Taylor (1993) elsewhere. However, the low metal concentrations found in coastal seawaters in this study indicate little or no heavy metal pollution in the Biobio Region.

The correlation between fertilization success and concentration of oil and grease was highly significant. It has been suggested that oil and grease are an unidentified group of hydrophobic compounds, associated with high levels of organic contaminants such as phenols and organochlorine pesticides (US EPA 1976). We found that stations with the highest levels of oil and grease (Rocuant and San Vicente sectors) showed the lowest fertilization success. Similarly, Swartz et al. (1984) found that relative toxicities of sewage sludges for the infaunal marine amphipod Rhepoxynius abronius were correlated with multiple factors, mainly with the concentration of oil and grease.

Organochlorine pesticides (e.g. DDT, dieldrin, lindane) are considered to be major environmental contaminants because they are accumulated in fatty tissues, produce high toxicity to many invertebrate species and are persistent in aquatic ecosystems (WHO 1989, Pridmore et al. 1991). Chuecas et al. (1991) have reported that concentrations of organochlorine pesticides in coastal seawaters within the Biobio Region are above EPA (1976) water quality criteria, in spite of governmental restrictions on their import, manufacture, sale and agricultural use. Our results show that levels of organochlorine pesticides are still above these criteria. The interactions between these contaminants might have strong negative effects in processes as sensitive as fertilization in marine invertebrates. However, in our study we did not find significant correlations between fertilization and the concentrations of phenols and different organochlorine pesticides (aldrin, dieldrin, lindane, DDT). The highest concentrations of these organic contaminants were detected in Rocuant Island and San Vicente sectors.

If the station with the highest concentrations of oil and grease (located in San Vicente Bay) is excluded from the analysis, fertilization shows a better correlation with phenols (r = -0.633, p < 0.001, Fig. 7a) and total organochlorine pesticides (r = -0.633, p < 0.001; Fig. 7b). When a backward linear multiple regression analysis was applied to these data, we found the following model: Fertilization = 41.7933 - 315.689(Phenols) - 0.348(Oil and greases) + 0.572(Saturation % of oxygen) + 480.038(Organochlorine pesticides) (r² = 0.919, p < 0.001). The results of these analyses indicate that high levels of oil and grease are associated with high levels of organic contaminants and that both are
related to decreased fertilization success in Arbacia spatuligera. On the basis of these data we suggest that Rocuant Island and San Vicente Port are the most polluted sectors within the Biobio Region.

Bioassay assessment and organic pollution within the Biobio Region

In many enclosed coastal marine areas, high organic sewage discharges producing extremely low oxygen concentrations in bottom waters have been identified with organic pollution processes (Rosenberg 1985, 1990). For many years, this process has been observed in semi-enclosed bays within the Biobio Region (Ahumada & Rudolph 1989, Ahumada et al. 1989b). According to our study, the most important seawater variables related to toxicity (i.e., decreased fertilization) were dissolved oxygen (positively related) and oil and grease (negatively related), both highly related to sewage pollution in the Region (Pradenas & Rudolph 1989). Oil and grease have been related to high loads of organic matter of untreated discharges of fishmeal factories (Rudolph et al. 1984). Low dissolved oxygen might be a consequence of high accumulation rates of this organic matter in the sediments.

Little is yet known about the potential toxic effects of chemical substances contained in the discharges of organic sewage (Swartz et al. 1984). It has been suggested that in the long term these discharges produce organic pollution, with important effects on the structure and energetic flows of benthic communities. Rosenberg (1985) and Baden et al. (1990) have associated organic pollution with mortality of benthic infauna and demersal fishes in waters of west Sweden; Croma et al. (1991) recorded immature communities with abundance of species of early successional stages in a highly organic polluted area within the Northern Adriatic Sea; Zmarzly et al. (1994) have reported that organic enrichment from wastewater discharges may increase relative abundances of small species and reduce temporal persistence of soft-bottom assemblages on the southern San Diego shelf.

In the Biobio Region, Oyarzun et al. (1987) have detected high density and biomass of opportunistic species associated with high organic matter enrichment in coastal sediments in Concepcion Bay. Carbaljal-Villalta (1994) mentioned that the increase of organic enrichment in coastal sediments could be responsible for long-term perturbations of the structure and dynamics of benthic macrofaunal communities within San Vicente Bay.

Although direct relationships between specific pollutants in organically polluted environments and biological responses in the laboratory have not been firmly established, our results suggest that multiple interactions between physico-chemical variables and organic pollutants can have significant effects on fertilization success in sea urchins. This might also have negative effects on the reproduction of other species of invertebrates belonging to these coastal ecosystems.

CONCLUSIONS

The results of sea urchin fertilization bioassays and the relationship between fertilization and environmental variables provided confirmation of organic pollution in the bays of the Biobio Region. However, according to Swartz et al. (1984), single-species toxicity test results should not be extrapolated uncritically to field situations.

We conclude that semi-enclosed coastal marine environments in the Biobio Region, mainly Rocuant Island and San Vicente Bay, are subject to organic pollution, which is demonstrated by the presence of high levels of organic compounds (mainly oil and grease)
and low oxygen concentrations. These extreme conditions were related to biological effects, as shown by decreased fertilization in *Arbacia spatuligera*.

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