Food-web mediated export of biogenic carbon in oceans: hydrodynamic control

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ABSTRACT: This paper describes an approach to determine, using a small number of food-web or hydrodynamic variables, the partitioning of phytoplankton production among 3 carbon fluxes, i.e. remineralization within the euphotic zone, food-web transfer, and sinking to depth of organic particles. In order to do so, the flows of biogenic carbon in the marine pelagic environment are reduced to 5 broad pathways, which are ordered along a continuum of decreasing export relative to primary production. At one end of the continuum, ungrazed phytoplankton is exported to depth and, at the other, biogenic carbon is remineralized within the bacteria-microzooplankton loop. In the present paper, export of biogenic carbon from the euphotic zone includes food-web transfer of primary production and downward flux of particulate organic carbon (POC) into deep waters. A simple model and data from the literature lead to the conclusions that: (1) the export characteristics of pelagic ecosystems are largely determined by the size structure of primary production and the matching (or, conversely, segregation) between primary production and grazing and (2) total export from the euphotic zone is a function of the delivery of mechanical energy to the upper water column whereas the partitioning of total export between food-web transfer and sinking of POC is controlled by temporal variations in depth of the surface mixed layer. Food-web transfer is significant for marine resources and sinking of POC may contribute to the regulation of climate change (sequestration at depth of biogenic carbon).

KEY WORDS: Carbon flux · Oceans · Phytoplankton production · Size · Matching · Food web · Hydrodynamics

FLUXES OF BIOGENIC CARBON IN OCEANS: SIGNIFICANCE AND APPROACHES

In oceans, fluxes of biogenic carbon resulting from phytoplankton production are either remineralization (i.e. respiration) within the euphotic zone or export out of it. These fluxes are independent of horizontal advection because, advected or not, biogenic carbon can only be remineralized in the euphotic zone or exported out of it. Export, in the present paper, is defined as including both transfer to large animals and sinking of particulate organic carbon (POC) into deep waters. The paper does not consider the downward flux of dissolved organic carbon (DOC), in spite of its possible significance in some conditions (e.g. Carlson et al. 1994, Michaels et al. 1994), because this flux has not been quantified in most situations. These fluxes are not instantaneously coupled with phytoplankton production, as there often is transient accumulation of POC and DOC in the euphotic zone. The 2 particulate export fluxes have global socio-economic significance. Food-web export to large and long-lived animals sets an upper limit to marine renewable resources. This type of export is accompanied by respiratory losses (CO₂) of ingested carbon. Export towards deep waters is mainly through the production of fast-sinking faecal pellets and sedimentation of ungrazed algae, but it also includes other mechanisms such as vertical migrations of zooplankton (Longhurst et al. 1990), fall-out of organic debris and carcasses (e.g. Stockton & De Laa 1982, Noji 1991), and downward mixing of DOC produced by phytoplankton. In mesopelagic (Torres & Childress 1985, Cho & Azam 1988) and deep waters (Childress & Mickel 1985, Siebenaller & Somero 1989), part of the POC is respired by heterotrophic bacteria.
and metazoans. Some of the carbon that reaches the deep ocean may become sequestered, i.e. removed from the ocean-atmosphere system for at least 100 yr. Climate changes at various time scales, including the possible ongoing global warming, may be related to (or mediated by) variations in the pool of sequestered carbon.

There are at least 3 general approaches used to quantify biogenic carbon fluxes in oceans. The first consists in determining empirical relationships between phytoplankton production and some flux. For example, concerning fisheries, Iverson (1990) has shown that a linear relationship exists between the production of carnivorous fish and squid and that of phytoplankton, for open-ocean and coastal environments. Concerning the export of biogenic carbon to depth, several authors (e.g. Eppley & Peterson 1979, Betzer et al. 1984, Pace et al. 1987, Wassmann 1990) have proposed empirical models (equations) describing the downward POC flux as a function of total primary production in the euphotic zone. The second approach belongs to the general category of flow models. These models simulate the flows of one or several biogenic elements (e.g. C, N) among compartments that represent trophic levels or taxonomic groups (e.g. Fasham et al. 1990) or that are based on size classes (e.g. Moloney & Field 1991). Such models often include flows to large animals and out of the food web. The third approach aims at determining empirical relationships between some environmental variables and export of biogenic carbon. A fisheries example is the relationship determined by Curry & Roy (1989) between the annual recruitment of several fish species and Ekman-type upwelling.

The present paper proposes an approach to determine, using a small number of food-web or hydrodynamic variables, the partitioning of phytoplankton production among 3 fluxes, i.e. euphotic-zone remineralization, food-web transfer, and sinking of POC into deep waters. Equations are formulated and values of parameters are derived from observations.

**FLUXES, POOLS, AND FLOWS OF BIOGENIC CARBON IN OCEANS**

The above 3 fluxes of biogenic carbon in oceans generally correspond to the 3 carbon pools defined by Legendre & Le Fèvre (1992) on the basis of the turnover time of biogenic carbon (i.e. time elapsed between the photosynthetic uptake of carbon by phytoplankton and the return of this carbon as CO₂ to the surface waters or the atmosphere). The pools are: short-lived organic carbon (turnover time < 10⁻² yr = 3 to 4 d), long-lived organic carbon (turnover time ranging between 10⁻¹ and 10² yr), and sequestered biogenic carbon (turnover time > 10⁵ yr). Short-lived organic carbon consists of organisms with short turnover times as well as labile dissolved organic compounds and it mainly transits through microbial components of the food web. Long-lived organic carbon comprises renewable marine resources (e.g. fish, marine mammals) and also heterotrophic bacteria involved in the breakdown of organic matter derived from large heterotrophs. The latter stresses the fact that the turnover time of biogenic carbon is not the same as the doubling time of organisms which mediate the turnover, e.g. heterotrophic bacteria decomposing a whale carcass may have individual doubling times ≤1 d but they are part of a carbon pool whose turnover time is tens of years. Sequestered biogenic carbon may take various forms, among which are organic remains buried in sediments (including petroleum), inorganic deposits of biological origin (e.g. calcareous ooze, coral reefs, continental limestone), refractory dissolved organic matter, and dissolved CO₂ in deep waters resulting from the in situ oxidation (respiration) of organic compounds.

The sizes of marine phytoplankton cover almost 4 orders of magnitude, from ca 2 000 μm (diatom *Ethmodiscus rex*, in warm oceanic waters) down to 0.6–0.7 μm (ubiquitous prochlorophytes). In addition, photosynthetic products include dissolved organic matter exuded by phytoplankton. For simplicity, the size range of primary production is divided here into 3 classes: large phytoplankton (L: >5 μm), small phytoplankton (S: 0.2 to 5 μm), and DOC (<0.2 μm). Although marine microbiologists normally separate bacterioplankton from sea water using a 0.2 μm filter, the 'dissolved' fraction actually includes colloidal material and free viruses (≤0.1 μm). The choice here of 5 μm as the boundary between large and small phytoplankton, instead of the more usual picoplankton (<2 μm) and nanoplankton (2 to 20 μm), is because the present paper refers to trophic interactions and mesozooplankton generally do not efficiently graze particles <5 μm (e.g. Fortier et al. 1994). In addition, phytoplankton cells as small as 5 μm can aggregate upon collision and thus sink out of the euphotic zone, as shown experimentally by Kierboe et al. (1990) for 3 small diatoms, i.e. *Phaeodactylum tricornutum* (6.0 μm equivalent spherical diameter, ESD), *Thalassiosira pseudonana* (4.7 μm ESD), and *Skeletonema costatum* (5.5 μm ESD). Given that the size spectrum of photosynthetic products is continuous, from the largest phytoplankton to the smallest exuded organic molecules, the boundaries between large and small phytoplankton and DOC are somewhat arbitrary. The same is true for the turnover times of biogenic carbon, whose spectrum is continuous.
Fig. 1 summarizes the major flows of biogenic carbon in the euphotic zone of oceans, from the 3 size classes of primary production to the 3 carbon pools. Note that flows (defined here as exchanges of biogenic carbon among trophic compartments of the food web) are not the same as the 3 fluxes defined above (i.e. euphotic-zone remineralization, food-web transfer, and sinking to depth of organic particles). Following Legendre (1996), the Fig. 1 shows that oxidation of DOC by heterotrophic bacteria and respiration by microzooplankton generally correspond to short-lived organic carbon, because these flows rapidly convert organic carbon back to CO₂. The flows of organic carbon into the long-lived pool are mediated by mesozooplankton (mainly copepods) grazing on large cells, organic aggregates (e.g. marine snow) and microzooplankton, and by microphagous macrozooplankton (e.g. salps, dololids, appendicularians, and pteropods; these organisms can also feed on large particles) feeding on small particles and aggregates. Part of this carbon reaches large metazoans such as fish and marine mammals. Finally, sedimentation to deep waters of rapidly sinking aggregated algal cells and faecal pellets (mainly from microphagous macrozooplankton; see Fortier et al. 1994) may lead to the sequestration of biogenic carbon in deep ocean basins and, to a smaller extent, in sediments on continental shelves. As pointed out by Legendre & Le Fèvre (1991), long-lived (refractory) DOC can effectively sequester biogenic carbon without it necessarily being exported to depth.

**BIOGENIC CARBON PATHWAYS**

Legendre & Rassoulzadegan (1995) proposed that, in the pelagic environment, there is a continuum of trophic pathways, from the herbivorous food chain, to the multivorous and microbial food webs and to the microbial loop. By definition, the microbial loop is an almost closed system (a truly closed system could only be transient) of heterotrophic bacteria and zooflagellate grazers, in which the latter release dissolved organic matter (DOM) used as substrate by the former (e.g. Hågström et al. 1988). In contrast, the microbial food web includes small phytoplankton (which account for most primary production in oligotrophic oceans), so that it can export biogenic carbon (Rassoulzadegan 1993). The present paper focuses on the fate of biogenic carbon, and therefore considers an additional pathway, i.e. sinking of ungrazed phytoplankton. As illustrated in Fig. 1, the flows of biogenic carbon can be reduced to 5 broad pathways, which are ordered along a continuum of decreasing export relative to primary production: (1) sinking of ungrazed phytoplankton, (2) herbivorous, (3) multivorous and (4) microbial food webs, and (5) the microbial loop. At one end of the continuum (1), there is sinking of ungrazed phytoplankton and, at the other (5), biogenic carbon is remineralized within the bacteria-macrozooplankton loop. In (3), the multivorous food web, herbivorous and microbial trophic modes both play significant roles. In the 2 other, intermediate pathways, (2) herbivorous and (4) microbial food webs, are dominated by herbivorous and microbial grazing, respectively. Within the context of the present paper, it is much easier to determine relationships among a small number of biogenic carbon pathways and food-web or hydrodynamic variables than for a large number of interconnected flows.

The flows of biogenic carbon within ecosystems are largely determined by the size structure of primary producers and by grazing pressure. These 2 factors are controlled by the degree of segregation (or, conversely, matching = 1 - segregation) among trophic components. The concept of segregation was introduced by Margalef (e.g. 1978, 1985) who stressed that, when trophic components are segregated...
Fig. 2. Size-fractionated phytoplankton production (P) from various marine environments. Values in the large (P_L) and small (P_S) size fractions are plotted as functions of total primary production \( P_T = P_L + P_S \). The main diagonal represents \( P_T \) and the dashed lines provide visual references corresponding to 550 mg C m\(^{-2}\) d\(^{-1}\). For \( P_T \) higher than the latter reference value (68 data points), 56% (38/68) of the \( P_L \) points and 9% (6/68) of the \( P_S \) points are located above the horizontal line (i.e. >550 mg C m\(^{-2}\) d\(^{-1}\)). Total of 195 data points from 13 published papers (see Table 1).

Table 1: Characteristics of total and size-fractionated phytoplankton production data plotted in Fig. 2. Threshold between the 2 size fractions (\( \mu m \)), daily (D) or hourly (H) values, the latter multiplied by 10 in Fig. 2 to obtain estimates of daily primary production; ecological situation; number of data points; source of data.

<table>
<thead>
<tr>
<th>Threshold</th>
<th>D/H</th>
<th>Ecological situation</th>
<th>No. of data</th>
<th>Source</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>D</td>
<td>North Atlantic (39° to 44° N)</td>
<td>3</td>
<td>Glover et al. (1986)</td>
</tr>
<tr>
<td>1</td>
<td>D</td>
<td>Celtic Sea (50° N)</td>
<td>9</td>
<td>Joint et al. (1986)</td>
</tr>
<tr>
<td>1</td>
<td>D</td>
<td>North Atlantic (47° to 60° N)</td>
<td>28</td>
<td>Joint et al. (1993)</td>
</tr>
<tr>
<td>2</td>
<td>D</td>
<td>Indian Ocean (3° to 15° N)</td>
<td>1</td>
<td>Gomes et al. (1992)</td>
</tr>
<tr>
<td>2</td>
<td>D</td>
<td>Indian Ocean (18° to 23° N)</td>
<td>9</td>
<td>Jochem et al. (1993)</td>
</tr>
<tr>
<td>2</td>
<td>D</td>
<td>Tropical and subtropical Atlantic</td>
<td>15</td>
<td>Jochem et al. (1993)</td>
</tr>
<tr>
<td>2.5</td>
<td>D</td>
<td>Black Sea (43° N)</td>
<td>3</td>
<td>Krupatkin et al. (1991)</td>
</tr>
<tr>
<td>3</td>
<td>D</td>
<td>Baltic Sea (56° N)</td>
<td>12</td>
<td>Kuparinen (1987)</td>
</tr>
<tr>
<td>3</td>
<td>H</td>
<td>North Atlantic (35° to 44° N)</td>
<td>5</td>
<td>Glover et al. (1985)</td>
</tr>
<tr>
<td>3</td>
<td>H</td>
<td>Mediterranean Sea</td>
<td>4</td>
<td>Berman et al. (1984)</td>
</tr>
<tr>
<td>5</td>
<td>D</td>
<td>North Atlantic (39° to 44° N)</td>
<td>3</td>
<td>Glover et al. (1986)</td>
</tr>
<tr>
<td>5</td>
<td>D</td>
<td>Celtic Sea (50° N)</td>
<td>10</td>
<td>Joint et al. (1986)</td>
</tr>
<tr>
<td>5</td>
<td>D</td>
<td>North Atlantic and Arctic (75° N)</td>
<td>11</td>
<td>Legende et al. (1993)</td>
</tr>
<tr>
<td>5</td>
<td>D</td>
<td>Coastal North Pacific (38° N)</td>
<td>40</td>
<td>Cole et al. (1986)</td>
</tr>
<tr>
<td>5</td>
<td>D</td>
<td>North Atlantic (47° to 60° N)</td>
<td>28</td>
<td>Joint et al. (1993)</td>
</tr>
<tr>
<td>5</td>
<td>H</td>
<td>Coastal Norway (60° N)</td>
<td>14</td>
<td>Erga &amp; Heimdal (1984)</td>
</tr>
</tbody>
</table>

(i.e. exhibit low spatio-temporal covariance), exchanges of energy and materials within ecosystems are reduced. This concept was borrowed from colloid chemistry, where it is known that reactivity decreases upon compartmentation of the space and that the process can be reaccelerated by mixing (Smoluchowski 1918, Margalef 1993). Hence, high primary production reflects low spatio-temporal segregation between the 2 factors that govern phytoplankton, i.e. irradiance and nutrients. Fig. 2 shows that, in oceans, most primary production above ca 550 mg C m\(^{-2}\) d\(^{-1}\) is due to large phytoplankton (data taken from sources listed in Table 1). Hence, there exists an overall positive relationship between total phytoplankton production (\( P_T \)) and that of large phytoplankton (\( P_L \)). It follows that low segregation between irradiance and nutrients generally enhances \( P_L \). Concerning the second factor involved in the control of ecosystem flows, high grazing pressure reflects low spatio-temporal segregation between phytoplankton and their zooplankton grazers. To summarize, the flows of biogenic carbon in marine pelagic ecosystems are mainly governed by 2 food-web characteristics, i.e. the size structure of primary production and the segregation between primary production and grazing.

In the 2 extreme biogenic carbon pathways identified in Fig. 1, trophic components are highly segregated in space and time. At one end of the spectrum (1), massive sinking of ungrazed phytoplankton occurs when large cells are not grazed by mesozooplankton during the bloom, i.e. when there is strong segregation between the production of large phytoplankton and their grazing by zooplankton. At the other end of the spectrum (5), the microbial loop occurs when phyto-
plankton production is very low or even nil, because of severe irradiance or nutrient limitation, i.e. strong segregation between irradiance and nutrients. The 2 intermediate pathways exhibit intermediate levels of segregation between trophic components. (2) In the herbivorous food web, there is strong grazing pressure exerted on large phytoplankton by mesozooplankton. This is often the case near the end of a bloom, when the time lag between enhanced production of large cells and their grazing is relatively short. (4) In the microbial food web, the biomass of small phytoplankton is controlled by microzooplankton. This requires contact rates between phytoplankton and microzooplankton to be high. Finally, (3) in the multivorous food web, primary production is shared by large and small phytoplankton. Active grazing by large and small zooplankton indicates that there is little segregation between primary producers and their grazers.

In the next section, the above ideas will be expressed as equations. The size structure of primary production will then be quantified as the production of large to total phytoplankton \( \frac{P_L}{P_T} \), whereas the spatio-temporal coupling between primary production and grazing will be represented by the matching \( M; \) segregation = \( 1 - M \) between these 2 rate processes.

**FOOD-WEB CONTROL OF BIOGENIC CARBON PATHWAYS**

The 3 main fluxes of net (vs gross) phytoplankton production \( P \) are: (1) remineralization within the euphotic zone \( R \) (i.e. respiration) and export out of the euphotic zone \( E \) by (2) food-web transfer \( F \) and (3) downward POC flux \( D \), so that \( E = F + D \). Part of the carbon exported from the euphotic zone to the food web is rapidly egested, excreted or respired, so that only a small fraction of \( F \) contributes to the production of herbivores and other trophic compartments (secondary production is generally defined as a change in biomass per unit time). In order to apply the model locally, it is assumed that, over appropriate time and space scales, all phytoplankton production is respired or exported and horizontal advection is negligible relative to the other fluxes. The above terms are related as follows:

\[
P = R + E = R + F + D \tag{1}
\]

All terms in Eq. (1) have units of mass per unit volume (or area) and per unit time. Each term on the right-hand side may result from the combination of several food-web flows. For example, carbon respired within the euphotic zone which is fixed again by phytoplankton and ultimately transferred to the food web belongs to \( F \) and not \( R \).

Eq. (1) is valid not only for the total phytoplankton assemblage \( T \) but also for size fractions, i.e. large \( (L) \) and small \( (S) \) phytoplankton. The present development does not consider the third size class of primary production identified above \( (\text{DOC}) \); see Fig. 1. This is because the production of DOC by phytoplankton is not routinely determined at sea and little is known concerning the overall flows and fluxes of this component in the marine environment. However, it is generally agreed that, except under unusual circumstances, the production of DOC does not exceed ca 10% of total gross primary production \( (\text{e.g. Sharp 1984, Wood et al. 1992}) \). Hence, in Eq. (2), \( P_T \) does not include the exudation of DOC by phytoplankton. Each term of Eq. (1) can be algebraically partitioned between the large and small size fractions:

\[
P_T = P_L + P_S \tag{2}
\]

\[
E_T = E_L + E_S \tag{3}
\]

\[
R_T = R_L + R_S \tag{4}
\]

\[
F_T = F_L + F_S \tag{5}
\]

\[
D_T = D_L + D_S \tag{6}
\]

By using Eqs. (2) to (6), Eq. (1) is rewritten for the total phytoplankton assemblage and the 2 size fractions:

\[
P_T = R_T + E_T = R_L + F_L + D_L \tag{7}
\]

\[
P_L = R_L + E_L = R_L + F_L + D_L \tag{8}
\]

\[
P_S = R_S + E_S = R_S + F_S + D_S \tag{9}
\]

The model will first be developed using the following 2 simplifying assumptions: large phytoplankton are mostly not remineralized in the euphotic zone and small phytoplankton generally do not sink out of the euphotic zone. These 2 assumptions, which are discussed later (Eqs. 26 to 31), can be written as:

\[
R_L \ll F_L + D_L \tag{10}
\]

\[
D_S \ll R_S + F_S \tag{11}
\]

Combining Eqs. (8) & (10) and Eqs. (9) & (11), respectively, gives:

\[
P_L = F_L + D_L \tag{12}
\]

\[
P_S = R_S + F_S \tag{13}
\]

Additional assumptions are that the proportions of primary production channelled into \( F_L \) and \( F_S \) are direct functions of the degree of matching \( M \) between primary production and overall grazing. \( M \) is a dimensionless number, whose values range from 0 to 1. Linear equations are used here given that, as mentioned above, there is a linear relationship between primary production and food-web export \( (\text{e.g. Iverson 1990}) \). Hence:

\[
F_L/P_L = f(M) = k_L M, \text{ thus } F_L = P_L k_L M \tag{14}
\]

\[
F_S/P_S = f(M) = k_S M, \text{ thus } F_S = P_S k_S M \tag{15}
\]

where \( k_L \) and \( k_S \) are constants. Because \( F_L/P_L \leq 1, F_S/P_S \leq 1, \) and \( M \leq 1 \), it follows that \( k_L \leq 1 \) and \( k_S \leq 1 \).
Given Eq. (12), Eq. (6) is rewritten as:
\[ D_T - D_S = P_L - F_L \]

It will be shown below (Table 2) that, in situations where primary production is dominated by small phytoplankton \( (P_L/P_T \leq 0.35) \) and where, as a consequence, \( D_S \) is likely of significance, the total downward POC flux is very small \( (D_T/P_T \leq 0.1) \). Hence, it is assumed that, when \( D_T \equiv 0 \), \( D_S \) is small so that the above equation is simplified as follows:
\[ D_T = P_L - F_L \quad (16) \]

Combining Eqs. (14) & (16) gives:
\[ D_T = P_L - P_L k_L M = (1 - k_L M) P_L \quad (17) \]

In order to express \( D_T \) as a function of the size structure of primary production \( (P_L/P_T) \), Eq. (17) is divided by \( P_T \):
\[ D_T/P_T = (1 - k_L M) (P_L/P_T) \quad (18) \]

Similarly, given Eq. (13), Eq. (4) is rewritten as:
\[ R_T - R_L = P_S - F_S \quad (19) \]

It will be shown below (Table 2) that, in situations where primary production is dominated by large phytoplankton \( (P_L/P_T \geq 0.80) \) and where, as a consequence, \( R_L \) is likely of significance, total euphotic-zone remineralization is very small \( (R_T/P_T \leq 0.3) \). Hence, it is assumed that, when \( R_T \equiv 0 \), \( R_S \) is small so that the above equation is simplified as follows:
\[ R_T = P_S - F_S \quad (20) \]

Combining Eqs. (15) & (19) gives:
\[ R_T = P_S - P_S k_S M = (1 - k_S M) P_S \quad (21) \]

Which, after division by \( P_T \), becomes:
\[ R_T/P_T = (1 - k_S M) (P_S/P_T) \quad (22) \]

Or, using Eq. (2): 
\[ R_T/P_T = (1 - k_S M) (1 - P_L/P_T) \quad (23) \]

Eq. (7) provides the expression for \( F_T/P_T \):
\[ F_T/P_T = 1 - (R_T/P_T + D_T/P_T) \quad (24) \]

Using Eqs. (18) & (22), Eq. (23) is rewritten as a function of \( M \) and \( P_L/P_T \):
\[ F_T/P_T = M (k_S + (k_L - k_S) (P_L/P_T)) \quad (25) \]

It must be noted that, because of Eq. (2), Eq. (24) can be rewritten as follows:
\[ F_T/P_T = M (k_L P_L + k_S P_S) / P_T \]

which is identical to the sum \( F_L + F_S \) (Eqs. 14 & 15) = \( F_T \) (Eq. 5), divided by \( P_T \).

The ratio of total export from the euphotic zone, through the combined mechanisms of food-web transfers and downward POC flux (Eq. 1), to total primary production is computed using Eq. (22):
\[ E_T/P_T = F_T/P_T + D_T/P_T = 1 - R_T/P_T \]
\[ E_T/P_T = 1 - (1 - k_S M) (1 - P_L/P_T) \quad (26) \]

Eqs. (18), (22), (24) & (25) link \( D_T/P_T, R_T/P_T, F_T/P_T, \) and \( F_T/P_T \), respectively, to the 2 food-web variables identified in the previous section \( (P_L/P_T \) and \( M \)), using 2 constants \( (k_L \) and \( k_S \)).

Tremblay et al. (in press) have shown that there is a linear relationship, at sea, between \( P_L/P_T \) and the \( f \)-ratio, i.e. the ratio of nitrate uptake by phytoplankton to the total uptake of nitrogenous nutrients (Eppley & Peterson 1979). It follows that \( P_L/P_T \) provides a connection between the above equations and the uptake of nitrogenous nutrients in oceans.

The above 2 simplifying assumptions (Eqs. 10 & 11) may not be realistic under all circumstances. Concerning the assumption that \( R_L \leq (R_S + F_S) \) (Eq. 10), it is known that some heterotrophic dinoflagellates can eat diatoms as large as, or even larger than themselves (e.g. Hansen 1991, Lessard 1991). This, together with sloppy feeding by mesozooplankton (e.g. Roy et al. 1989), could lead to significant remineralization of large-size primary production within the euphotic zone. In situations with significantly high \( R_L \) (combined with low \( D_S \); Eq. 11), it can be shown that \( D_T/P_T \) (Eq. 18) would be diminished by \( R_L/P_T \) and that \( R_T/P_T \) (Eq. 21) would be augmented by the same quantity:
\[ D_T/P_T = (1 - k_L M) (P_L/P_T) - R_L/P_T \quad (26) \]
\[ R_T/P_T = (1 - k_S M) (1 - P_L/P_T) + R_L/P_T \quad (27) \]

Because \( F_T/P_T \) would be the same as in Eqs. (23) & (24), total export (Eq. 25) would be diminished by \( R_T/P_T \):
\[ F_T/P_T = 1 - (1 - k_S M) (1 - P_L/P_T) - R_L/P_T \quad (28) \]

Concerning the assumption that \( D_S \leq (R_S + F_S) \) (Eq. 11), there are cases where microphagous macrozooplankton package significant amounts of small phytoplankton into large, fast-sinking faecal pellets (see above and Fig. 1). In situations with significantly high \( D_S \) (combined with low \( R_S \); Eq. 10), it can be shown that \( D_T/P_T \) (Eq. 18) would be augmented by \( D_S/P_T \) and that \( D_T/P_T \) (Eq. 21) would be diminished by \( D_S/P_T \):
\[ D_T/P_T = (1 - k_L M) (P_L/P_T) + D_S/P_T \quad (29) \]
\[ R_T/P_T = (1 - k_S M) (1 - P_L/P_T) - D_S/P_T \quad (30) \]

Because \( F_T/P_T \) would be the same as in Eqs. (23) & (24), total export (Eq. 25) would be augmented by \( D_S/P_T \):
\[ E_T/P_T = 1 - (1 - k_S M) (1 - P_L/P_T) + D_S/P_T \quad (31) \]

The other 2 assumptions (Eqs. 14 & 15) will be discussed in the next section.

Finally, using different values of \( M \) for the large- and small-size fractions \( (M_L \) and \( M_S \)) would not change the...
above results. With 2 values of M, Eqs. (14) & (15) are rewritten:

\[ F_L = P_L c_L M_L \]
\[ F_S = P_S c_S M_S \]

However, values of coefficients \( c_L \) and \( c_S \) are not the same as \( k_L \) and \( k_S \):

\[ F_L = P_L k_L M = P_L c_L M_L \]
\[ F_S = P_S k_S M = P_S c_S M_S \]

so that:

\[ k_L M = c_L M_L \]
\[ k_S M = c_S M_S \]

Developing Eqs. (32) & (33) gives:

\[ D_T/P_T = (1 - c_L M_L) (P_L/P_T) \]
\[ R_T/P_T = (1 - c_S M_S) (1 - P_L/P_T) \]

Because of relations given by Eqs. (34) & (35) between \( k_L \) and \( c_L \) and \( k_S \) and \( c_S \), respectively, Eq. (36) is the same as Eq. (18) and Eq. (37) the same as Eq. (22). Hence, using 2 \( M \) values would not change the model, because of corresponding changes in the 2 constants. The model is thus based on a single \( M \) value.

**FITTING OF MODEL PARAMETERS TO DATA**

Table 2 shows values of \( P_L/P_T \), \( R_T/P_T \), \( F_T/P_T \), \( D_T/P_T \), and \( E_T/P_T \) for the 5 biogenic carbon pathways identified above, based on data from the literature. Values of \( M \) are derived from the authors' experience and are thus open to discussion (details concerning the derivation of \( M \) values are provided in the next section).

(1) Massive sinking of phytoplankton, following ungrazed blooms of large cells, has been reported in a number of papers (cited in Legendre 1990). Under such circumstances, primary production is strongly dominated by large cells (for simplicity, \( P_L/P_T = 1.0 \), even if there is always some production by small phytoplankton; see Fig. 2) and most of it sinks to depth \( (D_T/P_T = 1.0) \).

(2) Herbivorous grazing often occurs at times of blooms. In the Gulf of St. Lawrence (eastern Canada; Rivkin et al. 1996), during the spring phytoplankton bloom, the >5 pm size fraction accounted for 0.49 to 1.00 of the primary production (hence \( P_L/P_T = 0.8 \) in Table 2). Dividing the \( f \)-ratio (new/total phytoplankton production: \( P_N/P_T \)) by the ratio of \( P_L \) to the downward POC flux (POC was then mostly algal material, so that \( P_N/POC = P_N/D_T \), assuming no allochthonous input of POC) provides values for \( D_T/P_T = 0.02 \) to 0.09 (hence \( D_T/P_T = 0.1 \) in Table 2). The value \( R_T/P_T = 0.3 \) was chosen as intermediate between those for the sinking and multivorous pathways. By difference, \( F_T/P_T = 1 - (D_T/P_T + R_T/P_T) = 0.6 \).

(3) The paper of Rivkin et al. (1996) also provides values for the multivorous food web (post-bloom conditions): \( P_L/P_T = 0.09 \) to 0.43 and, given that most of the sinking POC flux was then of non-algal origin (their Fig. 2), the same calculation as above (there for \( D_T/P_T \)) gives \( F_T/P_T < 0.33 \) to 0.60. Additional estimates can be found in Moloney et al. (1991), who modelled pelagic ecosystems off western South Africa. Their results for phytoplankton combine sinking of ungrazed cells together with 'senescence' (and lysis; according to their Table 1, the senescence rate increases with decreasing cell size and increasing temperature). Two of the studied systems correspond to the multivorous food web, i.e. the Agulhas Bank and the Benguela Upwelling. According to their Fig. 21, \( P_L/P_T \) (gross) = 0.13 to 0.28 (proportion of total gross phytoplankton production in the >5 pm size fraction), \( D_T/P_T < 0.13 \) to 0.18 (proportion of net primary production, including exuded DOC, channelled into phytoplankton sinking and senescence), \( F_T/P_T = 0.30 \) to 0.31 (proportion of net primary production and exuded DOC exported as sinking faecal pellets), \( E_T/P_T = D_T/P_T + F_T/P_T < 0.42 \) to 0.48, and \( R_T/P_T = 1 - (E_T/P_T > 0.52 \) to 0.58. Finally, Legendre & Rassoulzadegan (1995) used the subarctic Pacific Ocean as an example for the multivorous food web Miller et al. (1991) published estimates for grazing on phytoplankton by micro- and mesozooplankton, based on several cruises in the area. Part of the carbon

<table>
<thead>
<tr>
<th>Biogenic carbon pathway</th>
<th>( P_L/P_T )</th>
<th>( M )</th>
<th>( R_T/P_T )</th>
<th>( F_T/P_T )</th>
<th>( D_T/P_T )</th>
<th>( E_T/P_T )</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Sinking of ungrazed cells</td>
<td>1.00</td>
<td>0.00</td>
<td>0.0</td>
<td>0.0</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>(2) Herbivorous food web</td>
<td>0.80</td>
<td>0.55</td>
<td>0.3</td>
<td>0.6</td>
<td>0.1</td>
<td>0.7</td>
</tr>
<tr>
<td>(3) Multivorous food web</td>
<td>0.35</td>
<td>0.65</td>
<td>0.6</td>
<td>0.3</td>
<td>0.1</td>
<td>0.4</td>
</tr>
<tr>
<td>(4) Microbial food web</td>
<td>0.10</td>
<td>0.25 or 0.75</td>
<td>0.8</td>
<td>0.2</td>
<td>0.0</td>
<td>0.2</td>
</tr>
<tr>
<td>(5) Microbial loop</td>
<td>0.00</td>
<td>0.00 or 1.09</td>
<td>1.0</td>
<td>0.0</td>
<td>0.0</td>
<td>0.0</td>
</tr>
</tbody>
</table>
grazed by small zooplankton is passed on to large zooplankton, so that micrograzing provides maximum estimates for $R_T$, whereas large zooplankton can graze both large algae and microzooplankton, so that macrograzing on phytoplankton provides minimum estimates for $F_T$. In Table 3 of Miller et al. (1991), values for the ratio of micro- to total grazing range from 0.70 to 0.93, hence $R_T/(R_T + F_T) \leq 0.70$ and $F_T/(R_T + F_T) \geq 0.30$. These are consistent with values of $R_T/P_T$ and $F_T/P_T$ in our Table 2, i.e. $R_T/(R_T + F_T) = 0.67$ and $F_T/(R_T + F_T) = 0.33$.

(4) Estimates for the microbial food web can be derived from model results of Moloney et al. (1991) for oceanic waters: $P_L/P_T = 0.03$ and $D_T/P_T = 0$ (because of dominance by small phytoplankton and high temperature, sinking is very low and senescence is very high). Proportions of net primary production (including exuded DOC) channelled into phytoplankton senescence and exported as sinking faecal pellets are 0.27 and 0.17, respectively. Assuming that ca 10% of the organic carbon from lysed phytoplankton is exported through the food web, $F_T/P_T = 0.17 + 0.03 = 0.2$, hence $R_T/P_T = 0.8$. Legendre & Rassoulzadegan (1995) used a study of Lancelot et al. (1993) on the ice-edge water column of the northwestern Weddell Sea to exemplify the microbial food web. In this system, phytoplankton 2 to 10 \( \mu \)m account for >80% of the autotrophic carbon biomass (their Fig. 3.3c). The model of Lancelot et al. (1993) provides the following estimates (their Table 6.11) for the marginal ice zone (MIZ) and the adjacent permanently open ocean zone (POOZ): $F_T/P_T < 0.34$ to 0.36 (proportion of net primary production available for grazing by large zooplankton), hence $R_T/P_T > 0.64$ to 0.66. In oceans, the microbial food web occurs under 2 drastically different types of conditions, i.e. well-stratified surface layer (e.g. oligotrophic tropical waters) or deeply mixed water column (e.g. winter at mid-latitudes), hence $M = 0.75$ or 0.25.

(5) For the microbial loop, values in Table 2 correspond to the extreme case of a closed remineralizing system of bacteria and microzooplankton. In a truly closed loop, there would be almost no autotrophic production ($P_T = 0$), so that $P_L/P_T = P_T/PT = F_T/P_T = D_T/P_T = E_T/P_T = 0$. The microbial loop can be seen as the limit case of the microbial food web (previous paragraph), hence $M = 1.00$ or 0.00.

Values of parameters $k_L$ and $k_S$ were computed by least-squares linear regression, using Eq. (24) and values in Table 2:

$$\left( \frac{F_T}{P_T} \right) / M = k_S + (k_L - k_S) \left( \frac{P_L}{P_T} \right)$$

Table 2 provides 7 pairs of values for $(F_T/P_T)/M$ and $R_T/P_T$. In the 2 cases where $F_T/P_T$ and $R_T/P_T$ are both equal to zero, it was decided that $(F_T/P_T)/M = 1$. The resulting values are $k_L = 1.0$ and $k_S = 0.4$.

Table 3 shows estimates of $R_T/P_T$, $F_T/P_T$, $D_T/P_T$, and $E_T/P_T$ for the 5 biogenic carbon pathways, computed using Eqs. (18), (22), (24) & (25) and values of $P_L/P_T$ and $M$ from Table 2. The 21 values for $R_T/P_T$, $F_T/P_T$, and $D_T/P_T$ account for $r^2 = 0.83$ of the variance of corresponding values in Table 2 ($r = 0.91$, $p < 0.001$). The high value of constant $k_S$ (1.0) suggests that, when phytoplankton production is dominated by the large size fraction ($P_L$), food-web transfer ($F_T$, mostly through herbivorous grazing) may be strongly influenced by the degree of matching ($M$) between primary production and grazing (Eq. 14). In contrast, the relatively low value of constant $k_L$ (0.4) suggests that, when the small size fraction is dominant ($P_S$), grazing on small phytoplankton ($F_S$) is not much influenced by $M$ (Eq. 15).

It is possible to increase the fit of model to observed data by increasing the number of forecasting variables. For example, Eqs. (26) & (27) could be used instead of Eqs. (18) & (22), in order to include in the model remineralization of large-sized primary production within the euphotic zone (RL). This should be especially significant for biogenic carbon pathways 2 and 3, where mesozooplankton grazers play a major role. Exact values for $R_T/P_T$ are not known, since part of the carbon released as DOC by grazing is ultimately exported after being taken up by bacteria. Given that ingestion and digestion of phytoplankton can release as

<table>
<thead>
<tr>
<th>Biogenic carbon pathway</th>
<th>$P_L/P_T$</th>
<th>$M$</th>
<th>$R_T/P_T$</th>
<th>$F_T/P_T$</th>
<th>$D_T/P_T$</th>
<th>$E_T/P_T$</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Sinking of ungrazed cells</td>
<td>1.00</td>
<td>0.00</td>
<td>0.00</td>
<td>0.00</td>
<td>1.00</td>
<td>1.00</td>
</tr>
<tr>
<td>(2) Herbivorous food web</td>
<td>0.80</td>
<td>0.55</td>
<td>0.16</td>
<td>0.48</td>
<td>0.36</td>
<td>0.84</td>
</tr>
<tr>
<td>(3) Multivorous food web</td>
<td>0.35</td>
<td>0.65</td>
<td>0.48</td>
<td>0.40</td>
<td>0.12</td>
<td>0.52</td>
</tr>
<tr>
<td>(4) Microbial food web</td>
<td>0.10</td>
<td>0.25</td>
<td>0.81</td>
<td>0.11</td>
<td>0.08</td>
<td>0.19</td>
</tr>
<tr>
<td>Microbial food web</td>
<td>0.10</td>
<td>0.75</td>
<td>0.63</td>
<td>0.34</td>
<td>0.03</td>
<td>0.37</td>
</tr>
<tr>
<td>(5) Microbial loop</td>
<td>0.00</td>
<td>0.00</td>
<td>1.00</td>
<td>0.00</td>
<td>0.00</td>
<td>0.08</td>
</tr>
<tr>
<td>Microbial loop</td>
<td>0.00</td>
<td>1.00</td>
<td>0.60</td>
<td>0.40</td>
<td>0.00</td>
<td>0.40</td>
</tr>
</tbody>
</table>
Table 4 Values of ecosystem characteristics for the 5 biogenic carbon pathways, computed here using Eqs. (24), (26) & (27), the values of $P_{1}/P_{T}$ and $M$ from Table 2, the values of $R_{1}/P_{T}$ defined in the text, and constants $k_{L} = 1.0$ and $k_{S} = 0.4$. Coefficient of determination $r^2 = 0.86$ between the 2 sets of 21 $R_{1}/P_{T}$, $F_{1}/P_{T}$, and $D_{1}/P_{T}$ values in Tables 2 & 4. Abbreviations as in Table 2.

<table>
<thead>
<tr>
<th>Biogenic carbon pathway</th>
<th>$P_{1}/P_{T}$</th>
<th>$M$</th>
<th>$R_{1}/P_{T}$</th>
<th>$R_{T}/P_{T}$</th>
<th>$F_{1}/P_{T}$</th>
<th>$D_{1}/P_{T}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Sinking of ungrazed cells</td>
<td>1.00</td>
<td>0.00</td>
<td>0.00</td>
<td>0.00</td>
<td>0.00</td>
<td>1.00</td>
</tr>
<tr>
<td>(2) Herbivorous food web</td>
<td>0.80</td>
<td>0.55</td>
<td>0.20</td>
<td>0.36</td>
<td>0.48</td>
<td>0.16</td>
</tr>
<tr>
<td>(3) Multivorous food web</td>
<td>0.35</td>
<td>0.65</td>
<td>0.10</td>
<td>0.58</td>
<td>0.40</td>
<td>0.02</td>
</tr>
<tr>
<td>(4) Microbial food web</td>
<td>0.10</td>
<td>0.25</td>
<td>0.03</td>
<td>0.66</td>
<td>0.34</td>
<td>0.00</td>
</tr>
<tr>
<td>Microbial food web</td>
<td>0.10</td>
<td>0.75</td>
<td>0.03</td>
<td>0.84</td>
<td>0.11</td>
<td>0.05</td>
</tr>
<tr>
<td>(5) Microbial loop</td>
<td>0.00</td>
<td>0.00</td>
<td>0.00</td>
<td>0.50</td>
<td>0.40</td>
<td>0.00</td>
</tr>
<tr>
<td>Microbial loop</td>
<td>0.00</td>
<td>1.00</td>
<td>0.00</td>
<td>1.00</td>
<td>0.00</td>
<td>0.00</td>
</tr>
</tbody>
</table>

Table 5. Values of ecosystem characteristics for the 5 biogenic carbon pathways, computed here as in Table 3, after perturbation of $P_{1}/P_{T}$ and $M$ by ±20%. Coefficients of determination between the 21 $R_{1}/P_{T}$, $F_{1}/P_{T}$, and $D_{1}/P_{T}$ values in Table 2 and each set of corresponding values in the present table: $r^2 = 0.83 (P_{1}/P_{T} - 20\%)$, 0.79 ($P_{1}/P_{T} + 20\%$), 0.85 ($M - 20\%$), 0.77 ($M + 20\%$). Abbreviations as in Table 2.

<table>
<thead>
<tr>
<th>Biogenic carbon pathway</th>
<th>$P_{1}/P_{T}$</th>
<th>$M$</th>
<th>$R_{1}/P_{T}$</th>
<th>$R_{T}/P_{T}$</th>
<th>$F_{1}/P_{T}$</th>
<th>$D_{1}/P_{T}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>±20% perturbation of $P_{1}/P_{T}$</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1) Sinking of ungrazed cells</td>
<td>0.80–1.00</td>
<td>0.00</td>
<td>0.20–0.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td>0.80–1.00</td>
</tr>
<tr>
<td>(2) Herbivorous food web</td>
<td>0.64–0.96</td>
<td>0.55</td>
<td>0.28–0.03</td>
<td>0.43–0.54</td>
<td>0.29–0.43</td>
<td></td>
</tr>
<tr>
<td>(3) Multivorous food web</td>
<td>0.28–0.42</td>
<td>0.65</td>
<td>0.53–0.43</td>
<td>0.37–0.42</td>
<td>0.10–0.15</td>
<td></td>
</tr>
<tr>
<td>(4) Microbial food web</td>
<td>0.08–0.12</td>
<td>0.75</td>
<td>0.64–0.62</td>
<td>0.34–0.35</td>
<td>0.02–0.03</td>
<td></td>
</tr>
<tr>
<td>Microbial food web</td>
<td>0.08–0.12</td>
<td>0.75</td>
<td>0.64–0.62</td>
<td>0.34–0.35</td>
<td>0.02–0.03</td>
<td></td>
</tr>
<tr>
<td>(5) Microbial loop</td>
<td>0.00–0.00</td>
<td>0.00</td>
<td>1.00–1.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td></td>
</tr>
<tr>
<td>Microbial loop</td>
<td>0.00–0.00</td>
<td>1.00</td>
<td>0.60–0.60</td>
<td>0.40–0.40</td>
<td>0.00–0.00</td>
<td></td>
</tr>
<tr>
<td>±20% perturbation of $M$</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1) Sinking of ungrazed cells</td>
<td>1.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td>1.00–1.00</td>
</tr>
<tr>
<td>(2) Herbivorous food web</td>
<td>0.80</td>
<td>0.44–0.66</td>
<td>0.16–0.15</td>
<td>0.39–0.58</td>
<td>0.45–0.27</td>
<td></td>
</tr>
<tr>
<td>(3) Multivorous food web</td>
<td>0.35</td>
<td>0.52–0.78</td>
<td>0.51–0.45</td>
<td>0.32–0.48</td>
<td>0.17–0.08</td>
<td></td>
</tr>
<tr>
<td>(4) Microbial food web</td>
<td>0.10</td>
<td>0.20–0.30</td>
<td>0.83–0.79</td>
<td>0.09–0.14</td>
<td>0.08–0.07</td>
<td></td>
</tr>
<tr>
<td>Microbial food web</td>
<td>0.10</td>
<td>0.60–0.90</td>
<td>0.68–0.58</td>
<td>0.28–0.41</td>
<td>0.04–0.01</td>
<td></td>
</tr>
<tr>
<td>(5) Microbial loop</td>
<td>0.00</td>
<td>0.00–0.00</td>
<td>1.00–1.00</td>
<td>0.00–0.00</td>
<td>0.00–0.00</td>
<td></td>
</tr>
<tr>
<td>Microbial loop</td>
<td>0.00</td>
<td>0.80–1.00</td>
<td>0.68–0.52</td>
<td>0.32–0.48</td>
<td>0.00–0.00</td>
<td></td>
</tr>
</tbody>
</table>

DOC between 0.1 and 0.5 of net particulate phytoplankton production (Jumars et al. 1989), values of $R_{1}/P_{T}$ (Table 4) were set at 0.20, 0.10, and 0.03 for pathways 2 through 4, respectively. Resulting estimates of $R_{1}/P_{T}$, $F_{1}/P_{T}$ and $D_{1}/P_{T}$ are given in Table 4. These account for $r^2 = 0.86$ of the variance of corresponding values in Table 2. Given that the model without $R_{1}/P_{T}$ ($t^2 = 0.83$, previous paragraph) accounts for almost as much variance as the model with $R_{1}/P_{T}$, the rule of parsimony dictates to use Eqs. (18), (22), (24) in the remainder of this paper. In other words, addition of euphoticzone remineralization of large-size primary production does not significantly improve the model, whose purpose is to determine the partitioning of phytoplankton production among 3 broad carbon fluxes.

In order to assess the effects of natural variability on the model, estimates of $R_{1}/P_{T}$, $F_{1}/P_{T}$, $D_{1}/P_{T}$ were computed for values of $P_{1}/P_{T}$ and $M$ 20% smaller and larger than those used above. Results are given in Table 5. The relatively small variability in carbon flux and $r^2$ values, in spite of the 40% perturbation of $P_{1}/P_{T}$ and $M$, shows that the model is quite robust with respect to natural variability.

Fig 3 illustrates the responses of $R_{1}/P_{T}$, $F_{1}/P_{T}$ and $D_{1}/P_{T}$ to the combined effects of $P_{1}/P_{T}$ and $M$. As already evident from Tables 2 & 3, the 5 biogenic carbon pathways are located on the figure according to their coordinates (Tables 2 & 3). In other words, addition of euphoticzone remineralization of large-size primary production does not significantly improve the model, whose purpose is to determine the partitioning of phytoplankton production among 3 broad carbon fluxes.
HYDRODYNAMIC CONTROL OF BIOGENIC CARBON PATHWAYS

The 2 axes of Fig. 3 (P/L and M) are under hydrodynamic control. Concerning the abscissa (P/L), hydrodynamic conditions that favour high P/L also lead to high P/L (Fig. 2). Primary production is generally enhanced at spatio-temporal transitions between 2 types of segregation (e.g. Legendre et al. 1986). Such transitions correspond to transient matching of factors whose segregation otherwise limits phytoplankton production. In oceans, strong vertical mixing generally causes light limitation, whereas prolonged stratification results in nutrient limitation. Thus, the mechanism that generally controls total phytoplankton production is the alternation, in time or space, from destabilized to vertical stratification of the water column (e.g. Mann & Lazier 1991). Legendre (1981) proposed to characterize the phytoplankton production potential of marine ecosystems by their frequency of destabilization-stabilization of the water column, defined as the number of sequences of vertical mixing/stabilization per unit time. According to Legendre (1981), a near-zero frequency identifies systems seldom destabilized or stabilized, whereas upwelling conditions tend to be near the upper limit of destabilization-stabilization (i.e. spatio-temporal coexistence of the 2 hydrodynamic conditions). It follows that the abscissa of Fig. 3 (P/L) likely corresponds to a positive gradient in the frequency of destabilization-stabilization of the water column (Fig. 4a).

Concerning the ordinate of Fig. 3, actual values of M and the hydrodynamic conditions that control the matching between primary production and grazing processes are not well known. (1) Massive sinking of ungrazed phytoplankton may occur following blooms of large cells, when grazing pressure is very low (hence M = 0). This is observed, for example, when the bloom is caused by rapid stabilization of a water column that has been deeply mixed for a long period (e.g. first stages of the spring bloom in the North Atlantic, at a time when mesozooplankton are still overwintering at depth; Parsons & Lalli 1988) or sudden injection of nutrients in nutrient-depleted surface waters (e.g. episodic blooms of large phytoplankton at the bottom of the euphotic zone in oligotrophic oceanic waters, where large grazers are mostly absent; Goldman 1988, 1993). Thus, the sudden release of factor(s) that limited phytoplankton production may cause low matching between enhanced production of large and small phytoplankton and herbivorous grazing, hence sinking of ungrazed cells. (2) The herbivorous food web requires spatio-temporal coincidence between enhanced production of large phytoplankton and the presence of herbivores, hence relatively high M. This occurs when conditions that favour the production of large phytoplankton persist on a time scale which is compatible with the response time of their grazers (e.g. late stages of the spring bloom in the North Atlantic, which coincide with presence of mesozooplankton in surface waters; Cushing 1989). (3) In the multivorous food web, there is production of both large and small phytoplankton and coexistence of grazing by large and small zooplankton, hence higher M than in the herbivorous web. A favourable condition is the alternation between vertical stability and moderate mixing (e.g. subarctic North Pacific, where it has been suggested that, as a consequence of shallow winter mixing, micro- and mesozooplankton grazers never mix...
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The brief review of grazing in the 5 biogenic carbon pathways suggests that there is a window in temporal variations in depth of the surface mixed layer within which the matching between primary production and grazing (M) is high and outside which segregation (1 - M) prevails. This is similar to the 'optimal environmental window' proposed by Cury & Roy (1989) for the relationship between annual fish recruitment (Peruvian anchoveta, Pacific sardine, West African sardines and sardinellas) and Ekman-type upwelling, where vertical advection, inputs of allochthonous nutrients and turbulence are related to wind speed. They found that recruitment is low during both weak upwelling (nutrient limited primary production) and strong upwelling (low matching between food and larval patches) and is optimal under moderate upwelling (wind speeds ca 5 to 6 m s⁻¹). Thus, in Fig. 3, the matching between primary production and grazing would be high under conditions of prolonged moderate mixing (pathways 2, 3, and 4-5 with high M). Conversely, low matching would occur in areas and at times (pathway 1) when rapid stratification of a deeply mixed water column does not allow mesozooplankton to track the enhanced phytoplankton production, hence accumulation of biomass (called a bloom; see Legendre 1990) followed by massive sinking of large cells. Low matching would also occur (pathways 4 and 5 with low M) in waters where contacts between autotrophic or heterotrophic producers and their microzooplankton grazers are reduced because of very high turbulence (e.g. deep winter mixing). The responses of biogenic carbon pathways to temporal variations in depth of the surface mixed layer are therefore nonlinear (Fig. 4b), all the more so because the characteristics of zooplankton grazers vary among groups.

Fig. 4. General relationships between the food-web and hydrodynamic factors that control biogenic carbon pathways. (a) Size structure of primary production (Pc/Pc, or Pc/Pc; abscissa in Fig. 3) plotted as a function of the frequency of stabilization-stabilization of the water column. (b) Matching (M) or segregation (1 - M) between primary production and grazing (ordinate in Fig. 3) plotted as a function of temporal variations in depth of the surface mixed layer (zm), expressed in terms of the ratio of minimum to maximum mixed layer depth (ratios zmin/max/zmax = 1 characterize steady conditions and ratios < 1 correspond to alternation between mixing and stability) and the ratio of minimum mixed layer depth to critical depth (when ratios zmin/max/zc < 1, there is net growth of phytoplankton in zm and, when ratios > 1, there is no net growth). The 5 pathways (identified by numbers 1 to 5, as in Figs. 1 & 3 and Tables 2 & 3) are positioned according to the values of (a) Pc/Pc and (b) M listed in Table 2.

The microbial food web is generally encountered, as mentioned above, under conditions of either prolonged stability (e.g. oligotrophic tropical waters) or continuous deep mixing (e.g. winter at mid-latitudes), these 2 types of environment being thus characterized by steady hydrodynamics. Under the first condition, primary production operates on a low supply of regenerated nutrients, whereas the second leads to light limitation of phytoplankton. The 2 types of limitation generally favour the production of small instead of large phytoplankton (e.g. Kirk 1986, Raven 1986). Under stable conditions, there is often tight coupling between the production of small phytoplankton and their grazing by microzooplankton (M = 0.75), because the latter have doubling rates that are similar to those of small phytoplankton (e.g. Sheldon et al. 1992, Riegman et al. 1993). This is presumably not the case when mixing is strong (M = 0.25). (5) In the microbial loop, net (vs gross) phytoplankton production is very low or even null. The first situation has been observed in oligotrophic tropical waters (M = 1), where the dominance of heterotrophic bacteria over phytoplankton has been interpreted as either a typical (Cho & Azam 1990) or a transient condition (Rassoulzadegan 1990). The second case occurs above the polar circle during winter (no solar radiation during several months), where all processes in the deeply mixed water column (M = 0) are heterotrophic.
from organisms that multiply very fast and thus often track phytoplankton (e.g. protozoan microzooplankton and large microphages) to mesozooplankton copepods that overwinter in deep waters and reproduce once or twice a year. It follows that the ordinate of Fig. 3 (M) likely reflects nonmonotonic responses to temporal variations in depth of the surface mixed layer (Fig. 4b). The latter is defined here by the ratios of minimum to maximum mixed layer depth ($z_{\text{mix}, \text{min}}/z_{\text{mix}, \text{max}}$) and of minimum mixed layer depth to critical depth ($z_{\text{mix}, \text{min}}/z_{c}$). Sverdrup 1953; $z_{c} = E_0/k E_{s}$, where $E_{s}$ is the irradiance at the sea surface, $k$ is the coefficient of diffuse light attenuation, and $E_{s}$ is the compensation irradiance for phytoplankton photosynthesis).

In Fig. 5, the 5 biogenic carbon pathways and the isopleths for $R_T/P_T$, $F_T/P_T$, and $D_T/P_T$ are positioned by reference to the above 2 hydrodynamics axes (frequency of destabilization-stabilization and temporal variations in depth of the surface mixed layer). The figure indicates that temporal variations in depth of the surface mixed layer do not influence $R_T/P_T$ much (isopleths almost parallel to the ordinate) but that they largely govern $F_T/P_T$ and $D_T/P_T$. The ordering of pathways in Fig. 5 is almost monotonous along both axes, which was not the case on the food-web axes ($P_I/P_T$ and $M$; Fig. 3). This suggests that the ultimate control of biogenic carbon export from the euphotic zone is exerted by hydrodynamic factors, through the proximal agency of food-web characteristics (Fig. 4).

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Fig. 6 illustrates the response of $E_T/P_T$ to the combined effects of (Fig. 6a) $P_I/P_T$ and $M$, computed from Eq. (25) with constants $k_L = 1.0$ and $k_S = 0.4$, and (Fig. 6b) the frequency of destabilization-stabilization...
and temporal variations in depth of the surface mixed layer. Because $E_T/P_T = 1 - R_T/P_T$, the isopleths for $E_T/P_T = 0.25, 0.50$ and $0.75$ in Fig. 6a, b are the same as those for $R_T/P_T = 0.25, 0.50$ and $0.75$ in Figs. 3 & 5, respectively. Fig. 6 shows that total export relative to primary production ($E_T/P_T$) is almost independent from $M$ or from temporal variations in depth of the surface mixed layer (isopleths almost parallel to the ordinates). This is because, in Eq. (25), $M$ is multiplied by the constant $k_s = 0.4$, which is relatively small. Thus, the degree of matching ($M$) between primary production and grazing, which is controlled by temporal variations in depth of the surface mixed layer (Fig. 4b), does not play a major role in determining total export from the euphotic zone. In contrast, $E_T/P_T$ is a direct function of $P_T/P_T$ or of the frequency of destabilization-stabilization (regular gradient along the abscissae) and, hence, of total primary production (Fig. 2).

The proportion of total export channelled through the food web ($F_T/E_T$) is:

$$F_T/E_T = (F_T/P_T)/(E_T/P_T)$$

which, using Eq. (24), gives:

$$F_T/E_T = [M/(E_T/P_T)]\left[k_L + (k_s - k_L)(P_T/P_T)\right]$$

and, because $E_T = F_T + D_T$, the proportion of total export channelled through the downward POC flux ($D_T/E_T$) is:

$$D_T/E_T = 1 - F_T/E_T$$

Eq. (38) shows that, for a given value of $P_L/P_T$, $F_T/E_T$ is a direct function of $M$ and an inverse function of $E_T/P_T$. The reverse is true for $D_T/E_T$ (Eq. 39). Thus, temporal variations in depth of the surface mixed layer, which are instrumental in determining $M$ (Fig. 4b), may play a major role in partitioning total export from the euphotic zone between food-web transfer and sinking of POC. Export to the food web is significant for marine resources (which are part of the long-lived organic carbon) and export to depth may contribute to the regulation of climate change (by sequestering sinking biogenic carbon).

Combining Figs. 1 & 3 shows that the pathways from marine primary production to carbon pools are largely determined by 2 food-web characteristics, i.e. the size structure of primary production ($P_T/P_T$) and the degree of matching between primary production and grazing ($M$). These 2 characteristics are under hydrodynamic control, i.e. $P_T/P_T$ responds to the delivery of mechanical energy to the euphotic zone (Fig. 4a) and $M$ reflects temporal variations in depth of the surface mixed layer as defined in Fig. 4b. The latter mechanism has little influence on total export from the euphotic zone (Fig. 6), but it may play a major role in partitioning total export between the food web and downward export fluxes (Eqs. 38 & 39). In addition, temporal variations in depth of the surface mixed layer: appear to strongly influence both food-web and downward POC export (Fig. 5). It follows that total export from the euphotic zone is a function of the delivery of mechanical energy and that the type of export (i.e. food-web vs sinking) is also determined by temporal variations in depth of the surface mixed layer.

**Determining Biogenic Carbon Fluxes Using Food-Web or Hydrodynamic Characteristics**

The equations in the present paper can be used to compute the partitioning of phytoplankton production among 3 biogenic carbon fluxes, i.e. euphotic-zone remineralization ($R$), food-web transfer ($F$), and downward POC flux ($D$). In order to do so, estimates of 2 food-web characteristics ($P_L/P_T$ and $M$) and of 2 constants ($k_L$ and $k_s$) are required. Values of $k_L$ and $k_s$ were derived, above, from published data. $P_L/P_T$ is relatively easy to determine at sea, but estimation of $M$ under field conditions has not been attempted yet. Alternatively, the 2 food-web characteristics can be replaced, in Eqs. (18), (22), (24), (25), (38) & (39), by hydrodynamic factors for which values may be more readily available or easier to estimate or model, i.e. the frequency of stabilization-destabilization of the water column (Fig. 4a) and temporal variations in depth of the surface mixed layer (Fig 4b). In this respect, future work should aim at quantifying the 2 relationships in Fig. 4. This quantification should include not only equations that relate food-web characteristics to hydrodynamic factors but also the spatio-temporal scales at which these equations apply. Relationships described in the present paper provide a simple framework for investigating the food-web and hydrodynamic controls of biogenic carbon fluxes in oceans. These controls were shown to be significant for both marine resources and climate change.

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