

Copepod foraging in patchy habitats and thin layers using a 2-D individual-based model

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ABSTRACT: Evidence has shown that thin, horizontally extending phytoplankton layers may be comprised of smaller high-concentration aggregations of phytoplankton, rather than a homogeneous high-concentration sheet. A 2-D (horizontal and vertical dimensions) individual-based model of copepod foraging was developed, in order to examine whether the foraging success of a copepod would be significantly affected by phytoplankton patchiness. The foraging rules for the simulated copepods were to decrease speed and increase turning angle when high food concentrations are encountered. The underlying distributions of phytoplankton used in the model were, for the patchy layer scenario, representations of raw 2-D field fluorescence obtained using the Optical Serial Section Tomography (OSST) device, and for the homogeneous layer scenario, distributions created by simulated vertical sampling of the OSST distributions with a CTD/Fluorometer. In both the patchy and homogeneous layer scenarios, the copepods always had higher net foraging efficiency than randomly behaving controls, suggesting that the simple behavioral rules adopted are advantageous for copepod-like organisms. Foraging efficiency was significantly greater for the patchy layer scenarios than for the homogeneous layer scenarios when patches were small (i.e. one step length in width) and intense (i.e. near ingestion-saturating concentrations). Ingestion was up to 30% higher in the most patchy case versus its paired homogeneous case, suggesting that the existence of patchiness is critical to copepod survival, and that sampling scales should not exceed the step length of a copepod.

KEY WORDS: *Acartia* · Foraging · Area-restricted search · Thin layers · Copepods · Individual-based model

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INTRODUCTION

Free-living planktonic herbivorous copepods depend upon phytoplankton for food. Finding phytoplankton is not necessarily easy, as it is patchily distributed throughout the water column at many scales: $O(10\text{ m})$ (Cullen & Eppley 1981), $O(1\text{ m})$ (Bjornsen & Nielsen 1991), $O(0.1\text{ m})$ (Owen 1989, Cowles et al. 1993), $O(1\text{ cm})$ (Cassie 1959, McAlice 1970, Derenbach et al. 1979, Mitchell & Fuhrman 1989, Owen et al. 1992, Tiselius et al. 1994). In some cases, subsurface chlorophyll maxima consist of smaller-scale features of several localized minima and maxima on a scale of

$O(10\text{ cm})$ in the vertical dimension (Cowles et al. 1993). These smaller features may span a range of chlorophyll concentrations that may be limiting through to satiating for copepods. The patches themselves extend horizontally for $O(10\text{ km})$ and can last for $O(10\text{ h})$, and have been labeled 'thin layers' (Cowles et al. 1993). Some species of copepods, which, when vertically migrating, can easily travel $O(10\text{ m})$ in a $O(10\text{ min})$, and $O(100\text{ m})$ over the course of 1 d, should encounter these thin layers repeatedly within their normal foraging ambit. The impact of these features on the feeding and growth of copepods, and the foraging strategies that copepods might use to exploit thin layers are presently unknown.

The difficulty in assessing the impact of thin layers on copepod growth arises from the current inability to

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sample copepods *in situ* on the same scale as the patchiness of the phytoplankton, or to follow individuals as they move within and between patches of food. This leaves modeling or laboratory studies as alternative methods for studying the impacts of microscale patchiness on copepod growth. Laboratory work has indicated that the existence of thin layers of food (i.e. patches) may significantly increase the growth of *Acartia tonsa*, as measured by either egg or fecal production rate (Tiselius 1992, Saiz et al. 1993). However, it is difficult to 'scale up' laboratory results obtained in small vessels to the field because the size of the vessels usually restricts the movement of the copepods over anything but the shortest time scales of observation, i.e. $O(10\text{ s})$ to $O(10\text{ min})$. Modeling studies can provide a way to examine the motions of copepods over any spatial or temporal scale, although they have the drawback that they are only as realistic as their inputs. In a modeling study, Tiselius et al. (1993) simulated the foraging of copepods for distributions of food that were horizontally homogeneous and vertically heterogeneous, i.e. thin layers. They found that as the degree of patchiness—defined as the relative difference between the concentration of food within a layer versus the average background concentration—changed, there was an impact on both growth and predation risk for the copepods. It was concluded that having a strong behavioral reaction to patchiness in the copepods' swimming behavior was more beneficial for avoiding predation than increasing ingestion. This may have resulted because the swimming behavior of their model copepods reacted in the same way to all patches of food above a minimum threshold concentration, although their ingestion rates would vary between these patches. Thus it is unclear from their study whether thin layers have an appreciable effect on growth rates, or whether their result was specific to the foraging strategy that was chosen.

New evidence concerning the structure of thin layers themselves is further confounding discussion of the importance of these layers on the feeding of planktonic copepods. In a field study, Jaffe et al. (1998), used a non-invasive camera system with a visual field in the vertical plane of $\sim 70 \times 70\text{ cm}$ and a resolution of $\sim 7 \times 7\text{ mm}$; they found that what appeared to be thin, continuous horizontal layers as measured by a fluorometer-equipped CTD, were actually comprised of small

$O(1\text{ cm})$ isotropic patches (Fig. 1). Thus, there may be times when thin layers are not of uniform concentration in the horizontal direction. If these images are accurate representations of thin layers, there could be important implications for both the grazing and foraging behavior of planktonic herbivorous copepods. First, copepods must find depths at which these high-concentration patches (i.e. the thin layers) exist, and then they must forage both horizontally and vertically to find the smaller individual patches which comprise the horizontal 'layer'. This implies that copepods need foraging strategies that will function over multiple spatial scales, if patchy thin layers are a regular feature of their planktonic environment. It is unclear how often layers are not horizontally homogeneous, and additional sampling of microscale phytoplankton distributions is needed.

In the present study, an individual-based simulation model (Leising 2000) was used to examine the impact of different dimensionalities of thin layer patchiness on the foraging success of planktonic copepods. The simulated copepods were given a correlated random walk as their foraging behavior, which is essentially an 'area-restricted search' (Tinbergen et al. 1967), where a forager takes shorter step lengths and turns at greater angles between steps as the density of prey increases. This behavior is somewhat different than the behavior used by Tiselius et al. (1993) as in the cur-

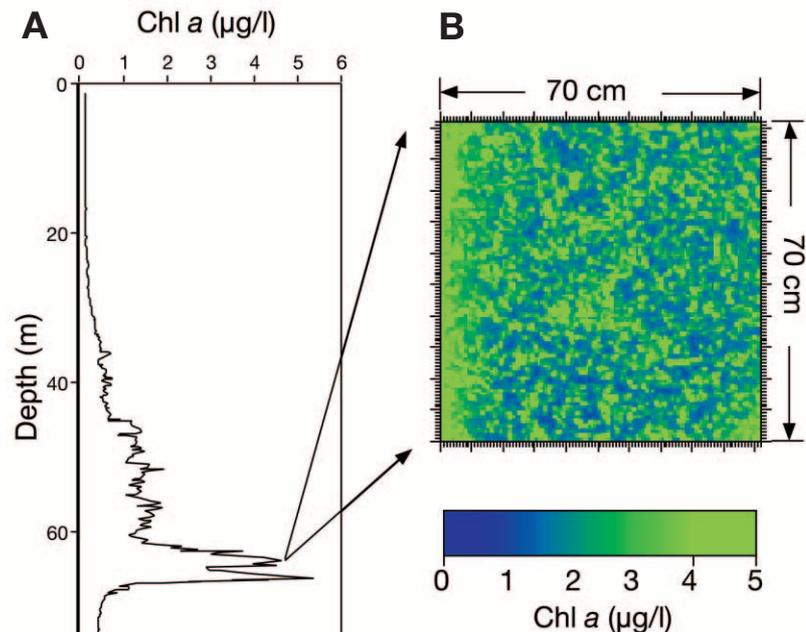


Fig. 1. (A) Chlorophyll vs depth (Seabird SBE-19 CTD with Wetlabs WetStar fluorometer). (B) Chlorophyll at depth $\approx 64\text{ m}$ (Optical Serial Section Tomography data). Image size $\approx 70 \times 70\text{ cm}$, resolution 100×100 pixels (Jaffe et al. 1998, Leising 2000)

rent model the behavioral response of the copepods varies in proportion with the strength of the stimulus, i.e. the concentration of food. The main questions that these simulations attempt to address are: (1) Is there a large impact on copepod feeding when thin horizontal layers are themselves made up of smaller patches? and (2) What effect, if any, does the choice of a particular functional (i.e. swimming) response have on the foraging success of the copepods in these various thin-layer environments?

MATERIALS AND METHODS

Model description. The model used for this study was a FORTRAN77 version of a MATLAB simulation model called SEARCH (Simulator for Exploring Area-Restricted search in Complex Habitats) (Leising 2000). In general, the copepods move around in a 2 dimensional grid that has reflecting boundaries at the top and bottom, and wrap-around boundaries on the sides. The 'area-restricted search' behavior, defined in the 'Introduction', was chosen because it has been seen to result in aggregation into areas of higher food resources in other animals, including insects (Karieva 1982, Karieva & Odell 1987, Ferran & Dixon 1993), birds (Tinbergen et al. 1967, Smith 1974, Zach & Falls 1977), and mammals (Benedix 1993, Cassini & Föger 1995, Haskell 1997). Some copepods have been observed to decrease their swimming speed or decrease their frequency of 'hop' or 'jump' behavior when encountering higher food concentrations, and may also turn at greater angles, a characteristic of area-restricted search behavior (*Pseudocalanus minutus*, Buskey 1984; *Oithona davise* females, Uchima & Hiramio 1988; *Temora longicornus* and *Pseudocalanus elongans*, Tiselius & Jonsson 1990; *Acartia tonsa* females, Tiselius 1992).

The model is only briefly summarized here, as a detailed description of how the model works has been published elsewhere (Leising 2000). At each time-step, each individual copepod moves a particular step length S_i , defined as the straight line distance traveled before being allowed the possibility of turning. It may differ from swimming speed, i.e. the time taken to travel a set distance, depending on the distance over which swimming speed is calculated. Step length S_i is determined by the concentration of food C within the grid space where the animal is located:

$$S_i = S_{\max}(1 - C/[k_{sv} + C]) \quad (1)$$

where k_{sv} is the 'half-saturation constant' of step length response, and S_{\max} is the maximum step length allowed (Fig. 2A). This function is asymptotic towards zero at high concentrations, and is essentially a modi-

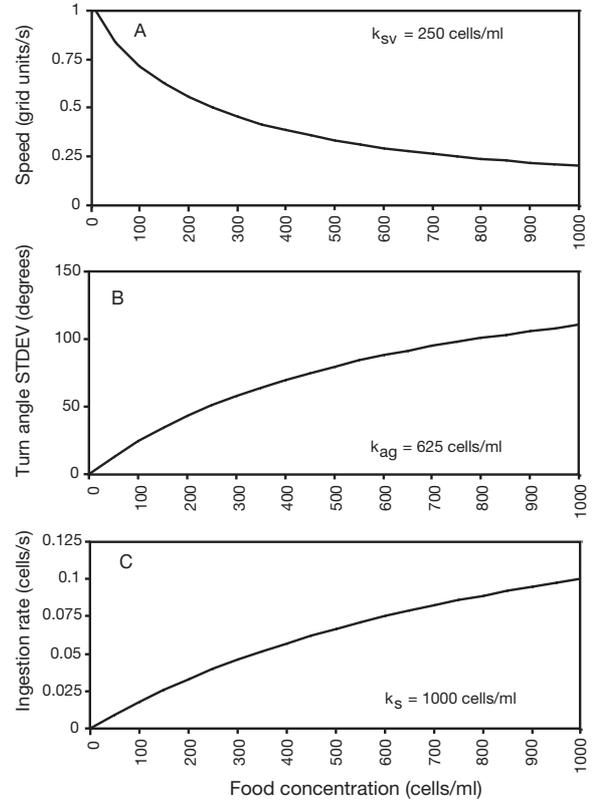


Fig. 2. Variation of forcing functions with food concentration: (A) step length; (B) standard deviation of turn angle probability density function; (C) ingestion rate

fied Holling Type-II response (Holling 1966). This formulation was chosen since it reflects what is known from the small amount of data concerning copepod step lengths versus food concentration (Piontkovskii & Petipa 1976). Before moving forward at each time-step, the simulated copepod turns. The probability of each particular turn angle is taken from a Gaussian distribution with the previous heading providing the mean, and the standard deviation A_{std} , varying with the underlying concentration of food C :

$$A_{std} = A_{\max}C/(k_{ag} + C) \quad (2)$$

where k_{ag} is the half-saturation constant for turning angle, and A_{\max} is the maximum allowed standard deviation of the probability density function for turning angle (PDF; Fig. 2B). For animals with anterior-posterior polarization, bilateral symmetry, and which therefore exhibit a tendency to continue in the direction they are already heading (e.g. copepods), this function should mimic such movement (Bovet & Benhamou 1988). This function has the property that at food concentrations that are low compared to the turning angle half-saturation constant, the copepod will have a high probability of continuing in a straight line. Conversely,

when A_{\max} is large (e.g. 180° for the current study), and the concentration of food is large compared to the turning angle half-saturation constant, turn angles after each step will be virtually uncorrelated with the previous heading; thus complete reversals of direction are possible between each time-step.

Once a copepod has turned and moved, it will eat phytoplankton at its new location, with an ingestion rate described by a Michaelis-Menten style equation:

$$I = I_{\max}C/(k_s + C) \quad (3)$$

where I_{\max} is the maximum ingestion rate of cells, k_s is the half-saturation constant for feeding, and C is the concentration of food (Fig. 2C). Laboratory data on the feeding of copepods suggests that this equation is a fair representation of ingestion rate (e.g. Mullin et al. 1975). No satiation response was included in the model, although over longer time periods, the copepods might be expected to fill their guts.

Foraging environment. The foraging arena was set to 50 units wide, by 500 units high. For copepods, the units will correspond to cm, making the foraging arena 50 cm wide by 5 m high. Horizontally, the copepods and food patches wrap around (i.e. copepods that swim off of one side, re-enter the environment on the other side, with no change in heading from their point of view), while vertically, the boundaries are reflecting (i.e. copepods reaching the top or bottom 'bounce' off the surface at 90° to their previous heading). These boundary conditions were chosen to reflect the idea that in nature, copepods can affect their vertical position relative to the small-scale distribution of their food at a scale of $O(10\text{ m})$; however, they cannot easily affect their horizontal position relative to the size of a horizontal layer with a potential scale of $O(10\text{ km})$. Thus the copepods will experience the smallest scale patchiness of $O(1\text{ cm})$ in both the vertical and horizontal direction, but will experience larger scale patchiness of $O(1\text{ m})$ in the vertical dimension only.

The phytoplankton patch structure was created as a vertical series of horizontally extending layers, in which the layers themselves were comprised of smaller patches. These smaller individual patches had a radially symmetric Gaussian distribution of phytoplankton concentration in cross section. The density of patches throughout the entire foraging arena (n_{patches}) was set at 375 patches per 10^4 cm^{-2} , while the maximum density of phytoplankton within the individual patches that comprise a layer varied as a function of depth. This patch density is close to $1/2$ of the patch density shown in the 2-D OSST image of Fig. 1, and is similar to the patch density that was found at areas adjacent to the chlorophyll maximum during that study (Jaffe et al. 1998). The maximum concentration of phytoplankton versus depth within these smaller patches was set by a

Gaussian probability density function with its mean at the vertical center of the water column and its standard deviation as $1/4$ the vertical height of the arena, such that the maximum concentration within patches was near the vertical center of the water column. The horizontally-extending layers were then created by adding a sine wave, with wavelength 100 cm and amplitude 100 cells ml^{-1} , to the vertical control of the maximum concentration of phytoplankton within individual patches. The wavelength of 100 cm was chosen because it results in 5 layers extending for approximately 50 cm in the vertical, with 50 cm spaces in between containing little, if any, phytoplankton. These spatial scales are similar to the range of thin layer scales that have been previously measured (~ 10 to 50 cm, reviewed in Cowles et al. 1998). The resulting maximum concentration of phytoplankton within an individual patch versus depth is shown in Fig. 3.

Two different patchy environment cases were then created by altering the standard deviation of the Gaussian cross-sectional density of the individual patches. In case A (Fig. 4A), the standard deviation of the smaller individual patches ($pstdev$) was set to 0.5 cm, and in case B (Fig. 5A), it was set to 2 cm. With $pstdev = 0.5$, the distance between ± 1 standard deviation of the Gaussian cross-sectional density of a patch

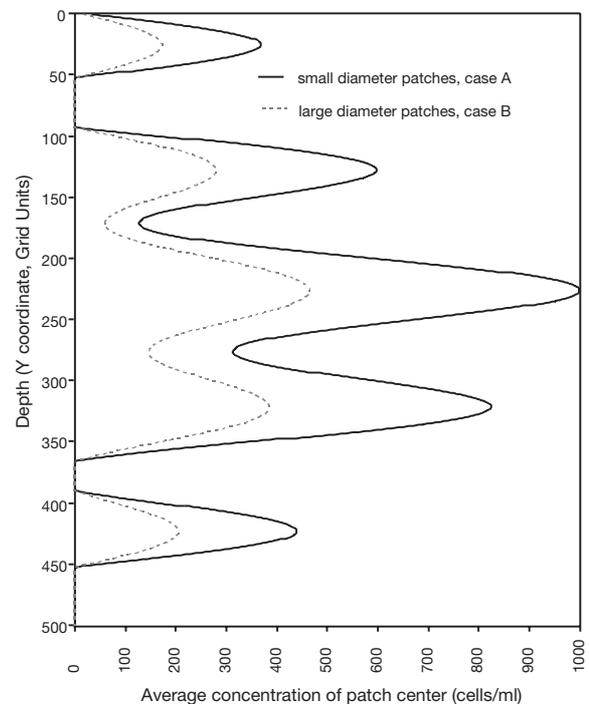


Fig. 3. Effect of vertical variation of $conc$ parameter on average cell concentration at center of patch. Structure created by combining a sine wave (amplitude 100 $conc$ units, wavelength 100 grid units) with a Gaussian distribution (mean $y = 250$, $SD = 125$ grid units). Solid line, $pstdev = 0.5$ grid units (case A); dashed line $pstdev = 2$ grid units (case B)

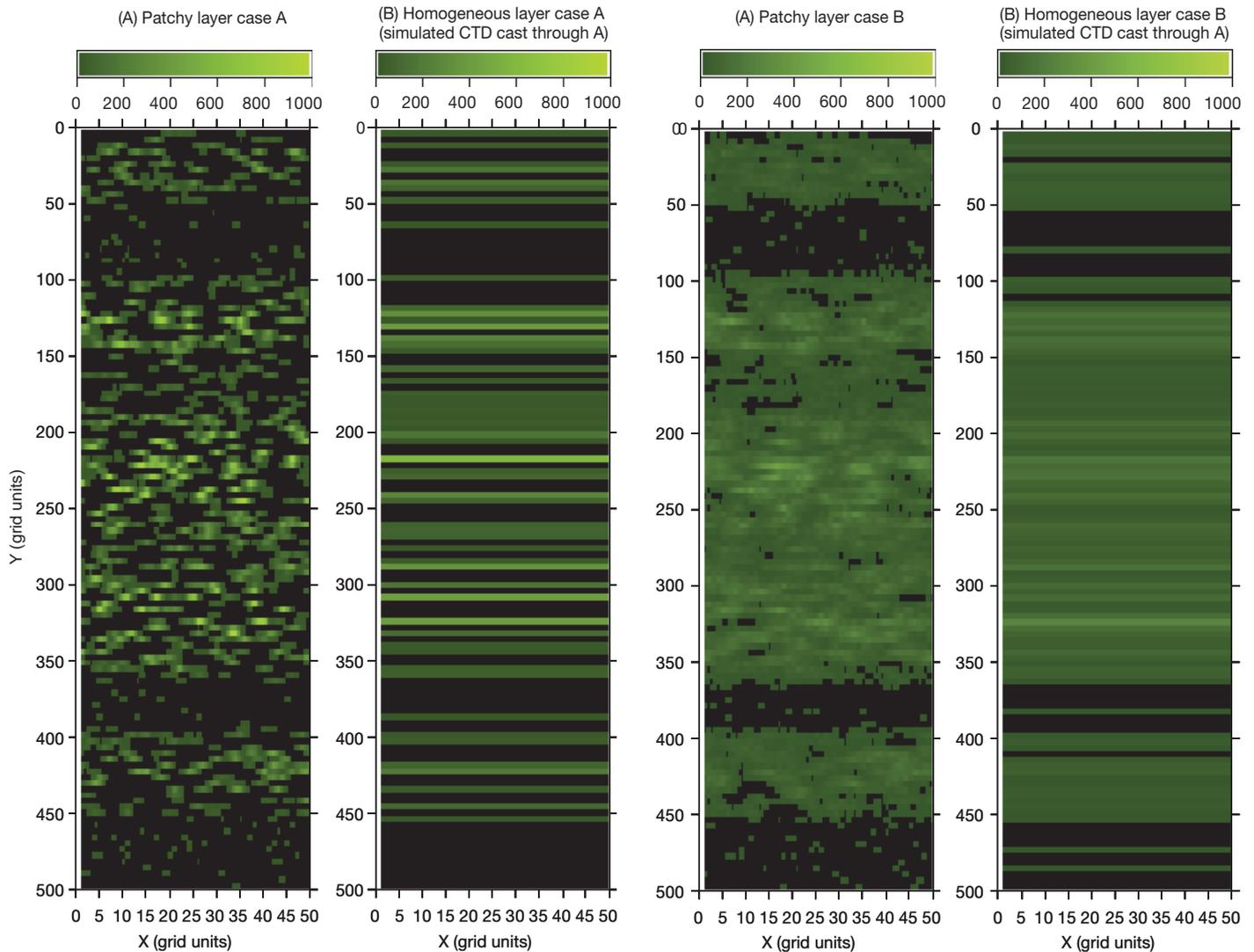


Fig. 4. Case A: (A) heterogeneous layer scenario ($n_{patches} = 375$ per 10^4 grid units², $ps_{tdev} = 0.5$ grid units), and (B) homogeneous layer scenario (patches in the vertical dimension only)

Fig. 5. Case B: (A) heterogeneous layer scenario ($n_{patches} = 375$ per 10^4 grid units², $ps_{tdev} = 2.0$ grid units), and (B) homogeneous layer scenario (patches in the vertical dimension only)

(which will be referred to as the standard width of a patch) is the same as the maximum step length of the simulated copepods, S_{max} , of 1 cm per time-step. In case B, the standard patch width was $4 \times S_{max}$. The overall result of these settings is that the ‘intensity’ of individual patches is greater in case A because the individual patches are narrower, although the total spatially integrated amount of food available is the same in both cases. These patch widths were chosen because they produce patches on the same scale as those observed by Jaffe et al. (1998); patches are 1 to 5 cm wide, with similar sized spaces in between containing little or no phytoplankton.

To estimate the effects of patchy thin layers versus homogeneous thin layers, horizontally homogeneous layers were created separately for both case A and

case B. This was accomplished by taking the average value of the 3 center-most grid squares horizontally from the above patch cases, and then replacing all grid squares horizontally, for each value of Y , with this average. The resulting distribution has structure only in the vertical direction, with a resolution of 1 grid unit (Figs. 4B & 5B). This structure is intended to be representative of what might be sampled with a CTD equipped with a high-resolution fluorometer. The total integrated number of cells available to be eaten was slightly different in these horizontally homogeneous scenarios due to the averaging procedure; spatially averaged concentration for case A was 50.3 cells ml^{-1} , and 49.8 cells ml^{-1} for its homogeneous approximation (i.e. 1% lower), and average concentration for case B was 50.4 versus 50.6 cells ml^{-1} for its homogeneous

approximation (i.e. <1% higher). This small difference in total average cell concentration should have little effect, however, since it has a nearly negligible effect on the average concentration within any particular grid square, which is what ultimately affects the responses of the copepods.

Specific foraging behavior. The simulated copepods were given a maximum speed of 1 cm s^{-1} . While this seems like a high speed for a copepod, they will only travel at this maximum speed when there is no food present, and this maximum speed is easily within 'hop' or 'jump' speeds that have been observed in the laboratory (Tiselius & Jonsson 1990, Saiz & Alcaraz 1992, Leising & Yen 1997). The maximum ingestion rate was set to 0.2 cells s^{-1} , and the half-saturation constant of the ingestion response, k_s , was set to $1000 \text{ cells ml}^{-1}$, which gives ingestion rates similar to those reported by Durbin & Durbin (1992) for *Acartia hudsonica* feeding on *Thalassiosira constricta*. Initial starting locations for copepods were random; however, all copepods started with a heading straight upwards, so that they would all encounter at least 1 patch within the simulation time of 21 600 s. As a measure of the foraging success of the copepods, the average number of cells eaten over the length of the simulation and the total distance traveled over the length of the simulation were recorded for each individual copepod. This allowed calculation of the ratio of number of 'cells eaten: distance traveled' (CEDT) for each copepod. The average and standard deviation of CEDT were also calculated for the population. It is hypothesized that as the distance traveled increases, the net gain from food eaten should decrease, if travel costs are not negligible (Morris et al. 1985, Morris et al. 1990) and predation risk increases (Gerritsen & Strickler 1977).

Model runs. Each run was for 100 copepods and simulated 6 h with a 1 s time-step. To examine the effect of varying the parameters k_{sv} and k_{ag} controlling the swimming response of the copepod, multiple runs were made for each patchy-layer environment (Figs. 4A & 5A). For the first set, k_{ag} was held constant at $250 \text{ cells cm}^{-2}$, and k_{sv} was either 250, 625 or $1000 \text{ cells cm}^{-2}$. For the second set, k_{sv} was held constant at $250 \text{ cells cm}^{-2}$, and k_{ag} was either 250, 625, or $1000 \text{ cells cm}^{-2}$. These two sets were then repeated for the homogeneous layer scenarios (Figs. 4B & 5B). Finally, random speed control simulations were run, to compare with each behavioral run from the patchy layer scenarios. For these control simulations, step length was drawn at random from a uniform distribution at each time-step, regardless of underlying food concentration and the maximum step length was set such that the average distance traveled by the individuals is the same as the average distance traveled in the behavioral simulations; any subsequent difference in CEDT is therefore due to differential feeding. A complete listing of the parameters used to force the SEARCH model are given in Table 1.

RESULTS

Individuals foraging within case A, whose step length controlling parameter had the highest sensitivity to food concentration (k_{sv} of $250 \text{ cells cm}^{-2}$), ate, on average, the highest number of cells compared with any other case (Fig. 6A). For both cases A and B, the average number of cells eaten decreased as step length sensitivity decreased for both patchy and homogeneous layer scenarios (Fig. 6). For patchy layers in both cases A and B,

Table 1. Parameter values used for the SEARCH model

Variable	Description	Value	Units
<i>time</i>	Number of time-steps for a run	21600	s
<i>gridx</i>	Grid size in horizontal dimension (x)	50	Grids
<i>gridy</i>	Grid size in vertical dimension (y)	500	Grids
<i>pstdev</i>	Standard deviation of patch diameter	0.5–2	Grids
<i>npatches</i>	Density of patches	375	10^4 grids^{-2}
<i>conc</i>	Number of grid cells filled per patch	200	–
<i>spotc</i>	Concentration required to fill grid spaces at each iteration	10	Cells ml^{-1}
c_{\max}	Maximum concentration of any single grid space	1000	Cells ml^{-1}
<i>wlength</i>	Wavelength of sine wave	100	Grids
<i>amp</i>	Amplitude of sine wave	100	Cells ml^{-1}
S_{\max}	Maximum step length of copepod	1	Grids
k_{sv}	Half-saturation constant of step length	250–1000	Cells ml^{-1}
A_{\max}	Maximum standard deviation of turning angle	180	Degrees
k_{ag}	Angle half-saturation constant	250–1000	Cells ml^{-1}
<i>number</i>	Number of copepods	100	–
I_{\max}	Maximum ingestion rate	0.2	$\text{Cells ind.}^{-1} \text{ time-step}^{-1}$
k_s	Half-saturation constant of ingestion response	1000	Cells ml^{-1}

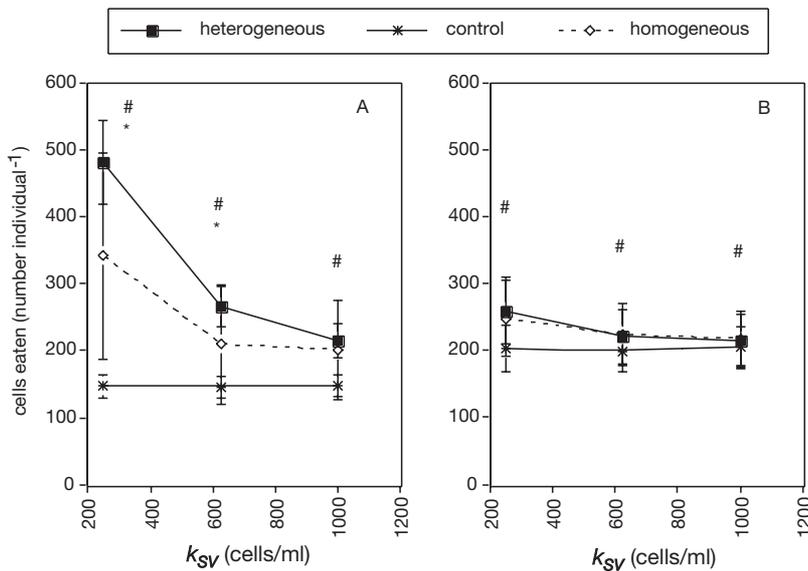


Fig. 6. Mean total cells eaten per copepod ($n = 100$) versus k_{sv} . Error bars ± 1 SD. $k_{ag} = 250$ cells per grid square. *Significant differences ($p < 0.05$, paired t -test with unequal variances) between heterogeneous and homogeneous layers and # between heterogeneous layer and control

the average number of cells eaten was significantly higher for area-restricted search runs than control runs for any combination of k_{sv} and k_{ag} that was run; having a behavioral rule was always better than having none ($p < 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 6 & 7).

For case A, the average number of cells eaten was significantly greater for the patchy layer scenario than the homogeneous layer scenario for any combination of k_{sv} and k_{ag} , except at the highest k_{sv} of 1000 cells cm^{-2} ($p < 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 6A & 7A). For case B, copepods in the patchy layer scenarios did not eat significantly more cells on average than copepods in the homogeneous layer scenarios ($p > 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 6B & 7B). It appears that high intensity patches (case A) led to a large difference between the amount of food ingested between the patchy and homogeneous layer scenarios, and little difference when individual patches were not as intense (case B). Within the high intensity patch environment (case A), copepods that had the highest step-length sensitivity ate the most, while varying their turn-angle sensitivity had little effect.

The trends of the results for the calculated CEDT ratio were nearly identical to those for the average number of cells eaten, as was expected since there is an inverse relationship between cells eaten and distance traveled in the model formulation. Distance traveled is not reported here, however, it can be estimated by dividing average CEDT by the average amount eaten for each run. The patchy layer environment (case A), with a k_{sv} of 250 cells cm^{-2} (i.e. highest sensitivity to food concentration) led to the highest average CEDT (Figs. 8A & 9A). For both cases A and B, CEDT was less as their step lengths became less sensitive to the underlying concentration of food (i.e. increased k_{sv}) (Fig. 8). For both cases A and B, the average CEDT was significantly higher for copepods using area-restricted search within the patchy layer scenario than in any control run, regard-

less of step-length or turn-angle sensitivity ($p < 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 8 & 9). For case A, CEDT was significantly greater for the patchy layer scenario than the homogeneous layer scenario, for any combination of k_{sv} and k_{ag} , except at the highest k_{sv} of 1000 cells cm^{-2} ($p < 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 8A & 9A). For

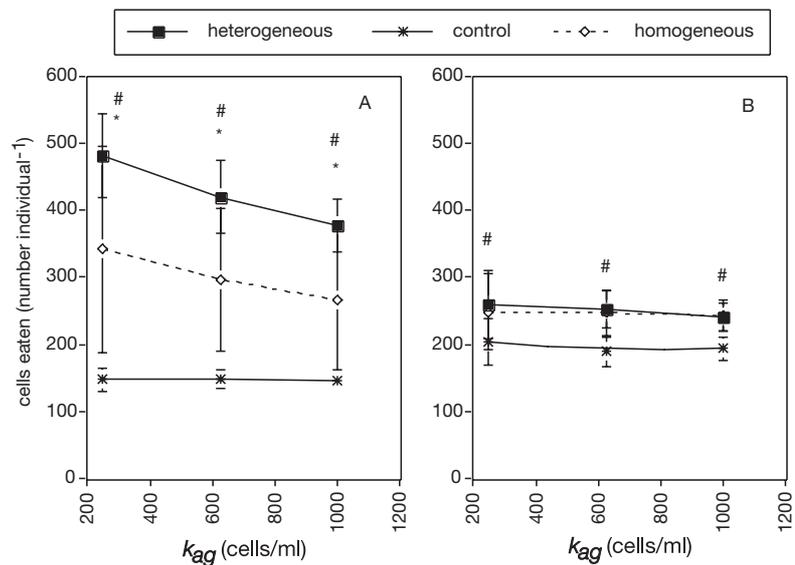


Fig. 7. Mean total cells eaten per copepod ($n = 100$) versus k_{ag} . Error bars ± 1 SD. $k_{sv} = 250$ cells per grid square. *Significant differences ($p < 0.05$, paired t -test with unequal variance) between heterogeneous and homogeneous layers and # between heterogeneous layer and control

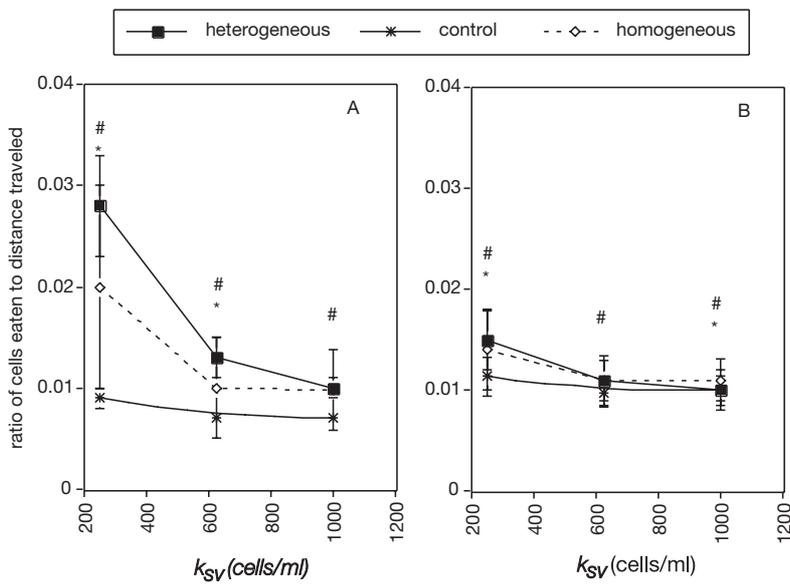


Fig. 8. Mean CEDT (cells eaten: distance traveled) (n = 100) versus k_{sv} . Error bars ± 1 SD. $k_{ag} = 250$ cells per grid square. *Significant differences ($p < 0.05$, paired t -test with unequal variance) between heterogeneous and homogeneous layers and # between heterogeneous layer and control

case B, CEDT was only significantly higher for the patchy layer scenario than the homogeneous layer scenario when both k_{sv} and k_{ag} were at their lowest (250 cells cm^{-2} cells/grid unit²) ($p > 0.05$, 2-sample t -test for samples with unequal variance—Satterthwaite's method; Figs. 8B & 9B). In fact, in case B, at high k_{sv} , the copepods ate significantly more in the homogeneous layer scenario than in the patchy layer scenario ($p < 0.05$, 2-sample t -test for samples with un-equal variance—Satterthwaite's method; Fig. 7B). Copepods generally ate more and traveled shorter distances in the patchy layers than the homogeneous layer approximations when the patchy layers were intense and when the copepods' step-length sensitivities were high. Turn-angle sensitivity had little effect regardless of the level of patchiness.

Comparing results between patch environments, the the highest values of food eaten and CEDT were found in case A, the 'patchiest' environment. The less patchy case B led to the lowest amount of food eaten, and therefore lower CEDT. With lower maximum concentrations available in case B (see Fig. 3), ingestion rates are also lower; copepods need to spend more time within these lower concentration

patches to ingest the same number of cells compared to a patch with a higher concentration. Conversely, patches should be easier to find, since they are 4 times wider, and copepods will therefore spend more time within patches than between patches.

The differences in time spent within and between patches, and the effects of the swimming parameters sensitivities, can best be seen by examining example tracks of individual copepods foraging through the simulated environments (Fig. 10). Fig. 10 shows examples of 4 individual copepods (1 copepod per panel), tracked for 600 s. Fig. 10A,C represent an enlargement of a 50 × 50 cm portion of patchy layer in case A (Fig. 4A), while Fig. 10B,D are 50 × 50 cm portions of patchy layer in case B (Fig. 5A). For illustration purposes, the boundaries wrap around, such that a copepod exiting the left side reappears on the right side of the panel. Fig. 10A,B show copepods that are very sensitive

in their swimming functional response ($k_{sv} = 250$ and $k_{ag} = 250$) while Fig. 10C,D show copepods that are less sensitive ($k_{sv} = 1000$ and $k_{ag} = 1000$). It is important to note that in Fig. 10, the phytoplankton concentration is not the same scale for all panels; white corresponds to a maximum concentration of 1000

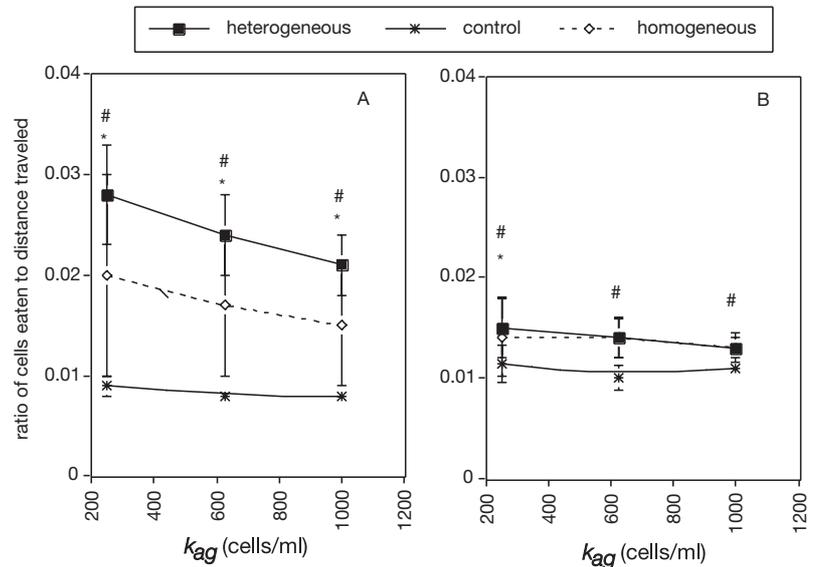


Fig. 9. Mean CEDT (cells eaten: distance traveled) (n = 100) versus k_{ag} . Error bars ± 1 SD. $k_{sv} = 250$ cells per grid square. *Significant differences ($p < 0.05$, paired t -test with unequal variance) between heterogeneous and homogeneous layers and # between heterogeneous layer and control

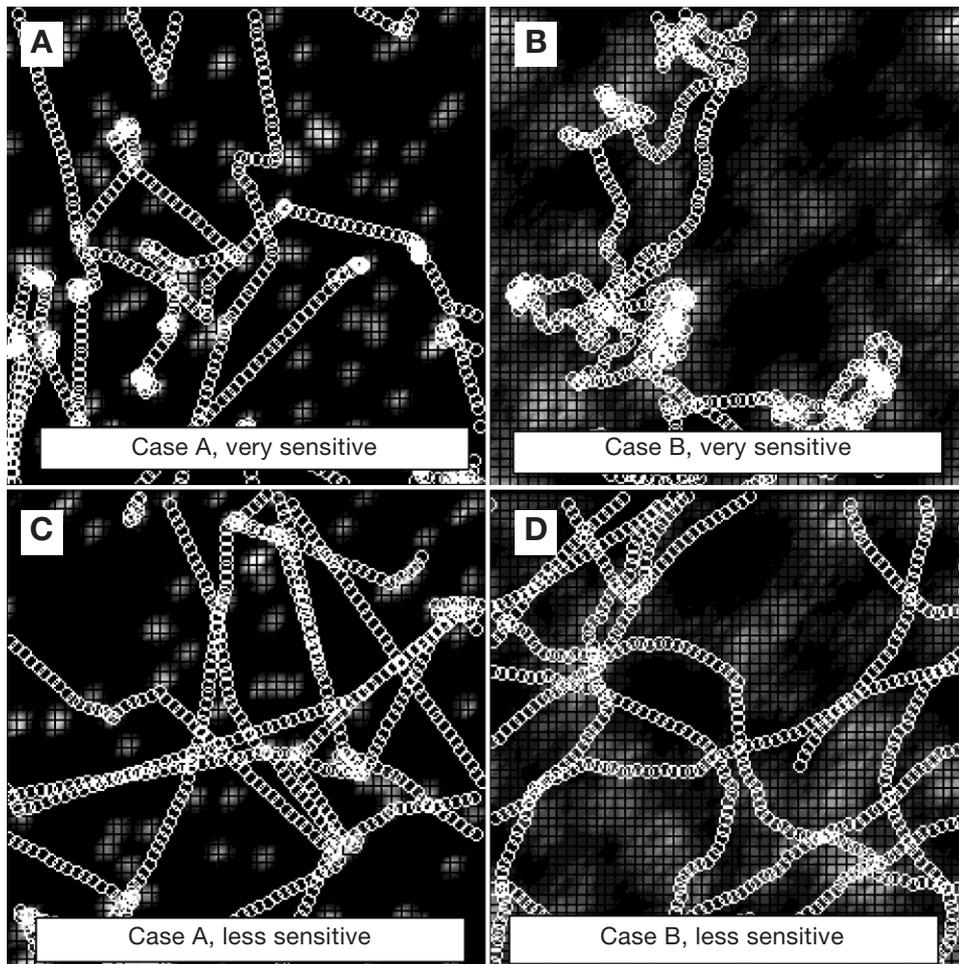


Fig. 10. Tracks for 4 individual copepods (1 track per panel) over 50×50 cm area, overlaid on phytoplankton distribution. Each track for 600 s (~ 600 iterations). White circles, position of copepod at 1 s intervals. Maximum phytoplankton concentration (white): case A (A,C) $1000 \text{ cells ml}^{-1}$; case B (B,D) $450 \text{ cells ml}^{-1}$. (A,B) Very sensitive swimming response ($k_{sv} = 250$, $k_{sv} = 250$), (C,D) less sensitive response ($k_{sv} = 1000$, $k_{ag} = 1000$)

cells ml^{-1} in Fig. 10A,C, but only $450 \text{ cells ml}^{-1}$ in Fig. 10B,D. In all 4 examples, copepods travel within and between several different individual patches within the 600 s; a short time compared to the 21 600 s total simulation time. When patches are intense (Fig. 10A,C) the copepods spend more time 'lost' between the patches. However, when they do encounter a patch, they stay there longer and feed at much higher rates, because the concentration of food within the patch is so much higher—patch center concentrations are between 800 and $1000 \text{ cells ml}^{-1}$ when patches are intense, versus 400 to $450 \text{ cells ml}^{-1}$ when they are diffuse. In addition, the copepods take more time to penetrate the center of a patch when the patch is more diffuse because the width of the patch is large compared to the step length of the copepod, such that it is more likely that the copepod may randomly change direction and exit the patch.

DISCUSSION

When thin horizontal layers are not homogeneous but are composed of smaller patches there can be a large difference in the foraging success of herbivorous copepods that actively move around as they search for food. The biggest difference in foraging efficiency between the patchy and homogeneous layer scenarios occurred when patches were small and intense. The results of this modeling exercise suggest that if we assume patchy thin layers are homogeneous, as we might when reconstructing the environment by averaging single vertical profiles of fluorescence, we will greatly underestimate the feeding rates of certain planktonic copepods. For the patchiest case A, the homogeneous layer approximations led to $\sim 30\%$ decrease in the average amount of food ingested. Alternatively, it is possible that thin layers are not

always patchy but are homogeneous at the microscale. When this is the case, the large decrease in ingestion rate could potentially be fatal to smaller copepods and copepods that do not store lipid reserves, such as the genus *Acartia* (Dagg 1977). Copepods that are omnivorous could switch to microzooplankton prey when phytoplankton is limiting in this fashion. Patchiness of food may be absolutely necessary for the survival of some copepods when food is limiting, if they cannot switch to an alternative feeding mode. There was also a large difference in the foraging success of the copepods between the 2 different heterogeneous cases—the average number of cells eaten was nearly double in case A, the environment with more intense patches—although the total available food was similar between the 2 environments. These 2 patch environments differed in the maximum concentration of food within a patch, which was a function of the width of the patch; patch widths varied by only a few centimeters between the 2 cases. In combination, these results suggest that patchiness needs to be sampled close to the same scale as the copepod's maximum step length, because using spatially averaged phytoplankton concentrations to estimate copepod feeding can be highly misleading.

Even if starvation does not occur, the egg production rate of a copepod will decrease when the environment is not patchy and food is limiting. Kleppel (1992) used field data to correlate egg production with daily ingestion rate for *Acartia tonsa*. The correlation coefficient was poor since the data were highly scattered, but based on that relationship, the 30% decrease in ingestion rate seen in the current study could lead to as much as a 50% decrease in egg production. As a lower boundary, a laboratory study by Kiørboe et al. (1985) showed that the egg production of *A. tonsa* decreased by at least as much as 20% given a 30% decrease in ingestion rate (measured in units of μgC). These levels of reduction of egg production are quite large, and further support the idea that without food being concentrated into small, more intense patches, some copepods may not produce enough eggs to persist. This being the case, foraging characteristics that allow copepods to take advantage of food patches should be highly selected for, while other copepods that are not adapted to deal with patchiness may be out-competed. Alternatively, copepods may switch their foraging mode completely, perhaps to 'ambush predation'—as seen in some *Acartia* species (Kiørboe et al. 1996)—in order to avoid problems related with actively foraging in patchy environments.

When food is not patchy, a copepod experiences a decreased ingestion rate, and would therefore have to increase the time it spends foraging and feeding in order to achieve the same total daily ingestion as

when food is patchily distributed. This may be an even greater problem for copepods than decreased egg production for 2 reasons, both related to predation risk. First, there may be an increase in predation risk if the swimming activities associated with feeding bouts make a copepod a more attractive target to a predator (Buskey et al. 1993, Buskey 1994). If this is true, then it would be advantageous for a copepod to quickly fill its gut, cease feeding activity, and perhaps even become passive so as not to attract either visual or mechanosensory predators. In a laboratory study, Saiz et al. (1993) found that *A. tonsa* spent less time within food patches when predators were present, suggesting that feeding activity is linked to predation risk. The second way in which decreased ingestion rate can lead to higher predation risk is through an increase in the encounter rate with predators. In the current model, ingestion rate and swimming speed are coupled, such that when swimming speed is high, ingestion rate is low. Based on the difference in the average swimming speed of individuals between the patchy and homogeneous layer scenarios in case A, and assuming a predator speed approximately twice that of the copepod, encounter rate would be around 5% higher for copepods in the homogeneous layer scenario (calculated using the encounter model of Evans 1989). This may seem like a small increase in predation risk relative to the large decrease in ingestion rate; however, a decrease in ingestion rate may not always be fatal, whereas a single encounter with a predator can be. Thus, predator avoidance can be an important factor controlling the behavior of an organism, and may, or may not, select for the same traits as those for optimally finding food.

In the current study, there was little difference in foraging efficiency when the parameter controlling the turn angle versus food concentration was changed (Figs. 7 & 9). On the other hand, changing the step length parameter (k_{sv}) had a large effect on the model results in both cases A and B (Figs. 6 & 8). For a 400% increase in k_{ag} there was a decrease of between 6 and 28% in CEDT, whereas for a 400% increase in k_{sv} , the decrease in CEDT was anywhere between 33 and 65%; changing step length sensitivity is more than twice as important as turn angle sensitivity for increasing foraging efficiency. This supports the results of an earlier modeling study by Doucet & Drost (1985), which found that klinokinesis—varying turn angle as a function of resource density—had little if any effect on the distribution of individuals, while orthokinesis—varying step length as a function of resource density—could lead to significant concentration of individuals within high resource areas. Their model, however, was run with only one particular 2-D patch environment (i.e. a perfect 'checkerboard' pattern). In this study, case A

showed a slight decreasing trend in CEDT as k_{ag} increased (Fig. 9A). From their modeling work examining the diffusion of predators through a patchy environment, Davis et al. (1991) suggested that changing turn angle as a function of food concentration should affect the rate of dispersion through the environment, but would not by itself lead to aggregation. If this is true, then it may be that for case A, where there was the largest distance between patch edges, changing the sensitivity of the turn angle parameter affected the rate of movement through patches, and therefore affected the retention rate within a patch and ultimately CEDT. For case B, the effects may not have been noticeable because the distance between patch edges was shorter.

Unlike the turning response, changing the sensitivity of the copepods' step length response had a large effect on foraging efficiency. For the different cases A and B, there was a general decrease in foraging efficiency as the sensitivity of a copepod's step length response to food concentration decreased (i.e. k_{sv} increased). This was also found in a modeling study by Leising & Franks (2000), which used the same parameterization of step length versus food concentration as the current model, although the movement of the simulated copepods was limited to the vertical dimension. Leising & Franks' (2000) results showed that while decreasing the sensitivity of a copepod's step length to food concentration led to a decrease in foraging efficiency, it led to an increase in the rate at which the copepod moved through its environment. This can clearly be seen by comparing Fig. 10A,B (sensitive copepods) with Fig. 10C,D (less sensitive copepods); the less sensitive copepods travel much more extensively through the environment. Although it was not explicitly tested in the 2-D model used here, the conclusion from Leising & Franks' (2000) 1-D model should hold; there is a trade-off between net foraging efficiency and movement rate through an environment. Moving through the environment rapidly to sample a larger amount of the available patches is important when first entering a new environment—it prevents the copepod from becoming 'stuck' in low-concentration patches near the periphery. This may be especially relevant to herbivorous copepods that migrate vertically each night into the upper water column to feed, and will therefore always encounter lower phytoplankton concentrations before reaching the sub-surface chlorophyll maximum.

Even though decreasing the sensitivity of a copepod's functional response, or assuming that layers were homogeneous, led to a decrease in foraging efficiency, it was always higher than the random controls. This is an important result, because it suggests that even when an environment is not very patchy, it is still advantageous to have a set behavior of some kind,

rather than to behave randomly. For the current study, a type of area-restricted search was chosen as the behavior. This behavior has been examined in many different taxa and appears to be a robust way to find and remain within high resource areas. According to the results presented here, it should also work well for herbivorous copepods that live in predictably patchy environments. In their 1-D model, Leising & Franks (2000) found that there was not a large increase in the *mean* foraging efficiency of copepods using area-restricted search versus random behavior, but that there was a large increase in the *variance* of foraging efficiency—more than 30% of the population had a higher CEDT ratio than any of the randomly behaving individuals. When expanding the model to 2-D, variance of foraging efficiency for the area-restricted searching copepods increased over the 1-D model, but there was also a significant increase in the mean for the population over the controls.

The increased difference in foraging efficiency between the copepods using area-restricted search and the controls when going from 1-D to 2-D can best be explained by considering the rate of motion of the copepods through space. For the control copepods in both cases, their motion can be modeled as a random walk. Following arguments in Berg (1983), the *rms* displacement $\langle r^2 \rangle^{1/2}$ over time t of a particle exhibiting a random walk will be a factor of 2 greater in 2-D than in 1-D space. This increase in displacement over time for the 2-D controls means that each copepod samples more of the environment, thus differences between individuals in ingestion rate due to the spatial variability of food concentration will be averaged out. As seen in the current 2-D model, with the subsequent decrease in the variance of food eaten in the control population, it becomes easier to statistically differentiate between the mean value of food ingested by the copepod populations using random and set behavior. Besides this difference in the control populations between 2-D and 1-D space, the copepods using area-restricted search in 2-D decrease their *rms* displacement—effectively their 'diffusivity'—to a greater extent than their 1-D counterparts, because in 2-D the copepods change not only their step lengths, but increase the tortuosity of their paths as well. Taking these facts into account, if the model were expanded to 3-D space there should be a further increase in the difference in foraging success between control and area-restricted searching copepods. For the controls, the *rms* displacement will increase by an additional factor of 1.2 for the 3-D case over the 2-D case (Berg 1983), while the area-restricted searching copepods will decrease their diffusivity because they have higher-dimensionality paths.

An appealing aspect of choosing area-restricted search behavior for a modeling study such as the cur-

rent one, is that while it can lead to complicated distributions which mimic other theoretically derived optimal distributions, such as the Ideal-Free Distribution (Fretwell & Lucas 1970), it does not require complicated actions by the copepod (Leising & Franks 2000). In fact, for some copepods, which feed and swim using some of the same appendages, there is no choice but to slow down when higher concentrations of food are encountered. It also appears that area-restricted search confers a greater advantage as the dimensionality and complexity of the environment increases. As mentioned earlier, there have been some laboratory studies showing that copepods exhibit movements characteristic of an area-restricted search. However, further studies are necessary to quantify the exact responses of swimming speeds and turning angles versus food concentration for particular species of copepods. Another important issue that was not addressed in the current model was satiation of the copepods feeding response. Many species of copepods could fill their guts in a much shorter time than the 6 h simulated here, and would probably change their feeding and swimming behavior at that point. This could have a large impact on the results of the current model, and will hopefully be addressed in future modeling work as more laboratory measurements of such effects are examined. With such information, it should be possible to use models such as SEARCH to make quantitative predictions of copepod growth and micro-scale distributions given real field-sampled phytoplankton distributions.

Acknowledgements. This work was supported by the US National Science Foundation OPP-9525803, Office of Naval Research N00014-95-1-0189 and N00014-95-1-0764 to Peter J. S. Franks, and a Postdoctoral fellowship from the University of Washington, School of Fisheries and Ocean Science to A.W.L. I would like to thank Peter J. S. Franks for advice and comments on this manuscript, and also the other members of the Franks Lab group for discussions concerning this work. Comments from 3 anonymous reviewers also helped to improve this contribution considerably. Special thanks to Jules Jaffe and Dave Zawada for letting me speculate wildly with their data from both the LUMIS and OSST systems.

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Editorial responsibility: Thomas Kjørboe (Contributing Editor), Charlottenlund, Denmark

Submitted: September 9, 1999; Accepted: October 11, 2000
Proofs received from author(s): May 30, 2001