The following supplement accompanies the article

Satellite tracking and stable isotope analysis highlight differential recruitment among foraging areas in green turtles

Phil J. Bradshaw, Annette C. Broderick, Carlos Carreras, Richard Inger, Wayne Fuller, Robin Snape, Kimberley L. Stokes, Brendan J. Godley*

*Corresponding author: b.j.godley@exeter.ac.uk

Marine Ecology Progress Series 582: 201–214 (2017)

Supplement 1. Satellite tracking

Table S1. Data from 28 green turtles satellite tracked from northern Cyprus.

| | | | | | | | | | | | Data |
|---------|-------|--------------|------|-----------------------------|-------------------|--------------|-------------|-------------|-------------|-------------|--------|
| ID | PTT | Release date | Days | Foraging area | Transmitter | Calibration | SI sample 1 | SI sample 2 | SI sample 3 | SI sample 4 | source |
| G077 | 4150 | 27/07/1998 | 294 | Libya (Gulf of Bomba) | Wildlife Computer | \checkmark | 2006* | 2011* | 2014 | | 1,2,3 |
| G078 | 4149 | 29/07/1998 | 222 | Libya (Gulf of Bomba) | Wildlife Computer | \checkmark | 2006* | 2014 | | | 1,2,3 |
| G082 | 4148 | 01/08/1998 | 289 | Turkey (Antalya Bay) | Wildlife Computer | × | | | | | 1,2,3 |
| G125 | 6598 | 19/07/1999 | 243 | West Libya (Gulf of Sirte) | Telonics ST18 | × | | | | | 1,2,3 |
| G059 | 4405 | 27/07/2002 | 403 | Turkey (Antalya Bay) | Telonics ST6 | \checkmark | 2012 | | | | 2,3 |
| G055 | 36638 | 12/07/2003 | 364 | West Libya (Gulf of Sirte) | Sirtrack 2003 | \checkmark | 2006 | 2010* | 2015 | | 2,3 |
| G008 | 36639 | 24/06/2004 | 395 | West Libya (Gulf of Sirte) | Sirtrack 2003 | \checkmark | 2012 | | | | 2,3 |
| G076 | 49815 | 08/07/2004 | 61 | Turkey (Antalya Bay) | Kiwisat 101 | × | | | | | 2,3 |
| G002 | 49816 | 23/07/2004 | 357 | Libya (Gulf of Bomba) | Kiwisat 101 | \checkmark | 2010 | 2014* | | | 2,3 |
| G044 | 49813 | 23/07/2004 | 311 | West Libya (Gulf of Sirte) | Kiwisat 101 | (C&N only) | 2007* | 2010* | 2013* | | 2,3 |
| Randall | 95099 | 08/06/2009 | 81 | Egypt (Lake Bardawil) | Kiwisat 101 | × | | | | | 4 |
| G015 | 95097 | 04/07/2009 | 486 | West Libya (Gulf of Sirte) | Kiwisat 101 | \checkmark | 2009 | 2013 | | | 3 |
| G157 | 95101 | 05/07/2009 | 715 | Libya (Gulf of Bomba) | Kiwisat 101 | \checkmark | 2009 | | | | 3 |
| G166 | 95098 | 15/07/2009 | 116 | Egypt (Gulf of Arab) | Kiwisat 101 | \checkmark | 2006 | | | | 3 |
| G189 | 95102 | 24/07/2009 | 110 | Egypt (Lake Bardawil) | Kiwisat 101 | \checkmark | 2009 | 2011 | | | 3 |
| G058 | 52820 | 16/06/2010 | 751 | Libya (Gulf of Bomba) | Kiwisat 101 | \checkmark | 2006* | 2010 | 2014 | | 3 |
| G009 | 86898 | 26/06/2010 | 475 | Libya (Gulf of Bomba) | Kiwisat 101 | \checkmark | 2010* | <u>2014</u> | | | 3 |
| G163 | 52846 | 28/06/2010 | 348 | West Libya | Kiwisat 101 | \checkmark | 2005* | 2007 | 2010* | 2013 | 3 |
| G080 | 52827 | 01/07/2010 | 407 | West Libya (Tunisia border) | Kiwisat 101 | \checkmark | 2010 | 2013* | | | 3 |
| G087 | 52949 | 07/07/2010 | 478 | Libya (Gulf of Bomba) | Kiwisat 101 | \checkmark | 2006* | 2010* | 2014 | | 3 |
| G006 | 86900 | 13/07/2010 | 412 | S. Cyprus | Kiwisat 101 | \checkmark | 2010* | 2015 | | | 3 |

P

| | | | | | | | | | | | Data |
|--------------|--------|--------------|------|-----------------------------|-------------------|--------------|-------------|-------------|-------------|-------------|--------|
| ID | PTT | Release date | Days | Foraging area | Transmitter | Calibration | SI sample 1 | SI sample 2 | SI sample 3 | SI sample 4 | source |
| G172 | 52888 | 21/07/2010 | 122 | West Libya (Tunisia border) | Kiwisat 101 | \checkmark | 2007* | 2010 | 2013 | | 3 |
| Pepsi Kibris | 52818 | 04/06/2011 | 134 | S. Cyprus | Kiwisat 101 | \checkmark | 2011 | | | | NA |
| G217 | 150429 | 30/06/2015 | 146 | Egypt (Lake Bardawil) | Wildlife Computer | \checkmark | 2009* | 2013 | 2015 | | NA |
| G252 | 150427 | 30/06/2015 | 68 | Egypt (Lake Bardawil) | Wildlife Computer | \checkmark | 2011 | 2015 | | | NA |
| G020 | 150430 | 01/07/2015 | 73 | Egypt (Lake Bardawil) | Wildlife Computer | \checkmark | 2009 | 2011 | 2013 | 2015 | NA |
| G201 | 150431 | 01/07/2015 | 58 | Egypt (Lake Bardawil) | Wildlife Computer | \checkmark | 2008* | 2011 | 2013* | 2015 | NA |
| G254 | 150428 | 01/07/2015 | 58 | Egypt (Lake Bardawil) | Wildlife Computer | \checkmark | 2011 | <u>2015</u> | | | NA |

ID = turtle's identification number (names in bold = male green turtles), PTT = numeric code for the platform terminal transmitter, Days = number of days the turtle was tracked for, Foraging area = conclusive end point where turtle was deemed resident, Calibration = turtles used to calibrate the discriminant analysis, SI samples 1 - 4 = which year tissue samples were collected for stable isotope analysis. Sample years underlined were specifically used to calibrate the discriminant analysis, * = only analysed for δ^{13} C or δ^{15} N. Data source legend, 1 = Godley et al. (2002), 2 = Broderick et al. (2007), 3 = Stokes et al. (2015), 4 = Wright et al. (2012) and NA = unpublished data.



Fig. S1. Post nesting green turtle satellite tracks recorded in 2015 from Cyprus to Lake Bardawil, Egypt. These 5 turtles were specifically selected for PTT attachment based on their δ^{13} C and δ^{15} N values.

Supplement 2. Lipid extraction

Samples from 20 green turtles that nested among years (2009 - 2014) were used to determine whether lipid extraction was necessary by subdividing the sample so that half were not lipid-extracted whilst the other half were lipid-extracted using a 2:1 chloroform:methanol ratio in a Soxlet apparatus and heated for 1 hour.

The selected samples had a pre-extraction C:N ratio of 2.68 (\pm SD = 0.06). No significant differences were found between untreated and lipid-extracted tissue samples for δ^{15} N (paired t-test, t₁₉ = 1.70, p = 0.11, S2a). Statistically significant differences were found for δ^{13} C (paired t-test, t₁₉ = - 4.0, p < 0.001, S2b) with a mean difference of -0.18 (range =-0.27 - 0.09). However, the differences in δ^{13} C values between lipid extracted and untreated samples were not substantially different considering the mean difference in δ^{13} C among foraging areas (1.68 ‰) and lipid extraction was not considered necessary for the whole dataset.



Fig. S2. Differences in stable isotope values between untreated and lipid extracted paired samples for $a = \delta^{15}N$ and $b = \delta^{13}C$ (n = 20). Grey dashed line = no difference (y = x).

Supplement 3. Storage concentration of ethanol

Paired epidermal tissue samples were collected simultaneously from 33 nesting females postoviposition and stored in a 96% and 70% ethanol concentration for up to 5 months. The concentration of ethanol had no significant effect on $\delta^{15}N$ (paired t-test, $t_{32} = 0.673$, p = 0.506, S3a) or $\delta^{13}C$ values (paired t-test, $t_{32} = -0.129$, p = 0.8981, S3b) as no consistent enrichment or depletion of $\delta^{15}N$ or $\delta^{13}C$ values was found among samples.



Fig. S3. Comparison of stable isotope values for paired green turtle epidermis samples (n = 33) stored in either 70% or 96% ethanol concentration, $a = \delta^{15}N$ values, $b = \delta^{13}C$ values. Grey dashed line = no difference (y = x).

Supplement 4. Predicting foraging area

This study adopted standard methods (Ceriani et al. 2012, Pajuelo et al. 2012, , Vander Zanden et al. 2015) to predict foraging areas using a discriminant function analysis evaluated by the leaveone-out cross validation method. Discrete differences in the combined isotopic values were assessed with a multivariate analysis of variance (MANOVA) with multiple pairwise comparisons conducted with Tukey's Honest Significant Difference to identify significant differences among foraging areas. Non-uniform priors were used based on the number of turtles tracked to each foraging area as they can improve the accuracy of assignment (Royle & Rubenstein 2004, Vander Zanden et al. 2015). We set a posterior probability of assignment at 80% or greater to maintain consistency among studies (Pajuelo et al. 2012, Seminoff et al. 2012, Vander Zanden et al. 2015) as this provides a 8 - 12 fold improvement in assignment over random odds considering 3 or 4 foraging areas, respectively (Wunder 2012, Vander Zanden et al. 2015).

| | Preli | minary DFA (cond | ucted prior t | io 2015) | Post 2015 DFA + secondary classification | | | | |
|--------------|-------|-------------------|---------------|------------|--|-------------------|----------|------------|--|
| Year sampled | Ν | Satellite tracked | Assigned | Unassigned | Ν | Satellite tracked | Assigned | Unassigned | |
| 2006 | 2 | 2 | | | 2 | 2 | | | |
| 2007 | 2 | 1 | 1 | | | | | | |
| 2008 | 2 | | 1 | | 1 | | 1 | | |
| 2009 | 24 | 1(1) | 12 | 10 | 13 | 1 | 10 | 2 | |
| 2010 | 14 | 4 | 10 | | 7 | 2 | 3 | 2 | |
| 2011 | 29 | 2 | 14 | 13 | 20 | 2 | 14 | 4 | |
| 2012 | 15 | 2 | 13 | | 6 | 2 | 1 | 3 | |
| 2013 | 64 | 3 (1) | 41 | 19 | 54 | 3 | 34 | 17 | |
| 2014 | 51 | 4(1) | 32 | 14 | 43 | 5 | 25 | 13 | |
| 2015 | | | | | 42 | 6 | 29 | 7 | |

Table S2. Sample sizes and the year that they were collected

| Total | 203 | 19 (22) | 124 | 56 | 188 23 | 117 | 48 |
|-------|-----|---------|-----|----|--------|-----|----|
| | | | | | | | |

Preliminary DFA was conducted using δ^{13} C and δ^{15} N prior to 2015, Post 2015 DFA + secondary classification was conducted using δ^{13} C, δ^{15} N and δ^{34} S. N = number of turtles sampled in each year, satellite tracked = number of turtles sampled that were satellite tracked to 1 of the 4 foraging areas, Assigned = number of turtles assigned to a foraging area, Unassigned = number of turtles that could not be assigned to a foraging area. Values in brackets under 'Satellite tracked' were the 3 turtles selected to characterise the unidentified foraging area and therefore their foraging area was not predicted

Supplement 5. Analysing foraging site fidelity

Foraging site fidelity was evaluated with a repeatability analysis employed in the R statistical package 'rptR' (Nakagawa & Schielzeth 2010) using a linear mixed-effects model based estimation for Gaussian data fitted with restricted maximum likelihood (REML). The identity of the turtle was the grouping factor and we controlled for the variance attributed to where a turtle forages by including this as a covariate, although 6 of the 45 turtles were unassigned to a foraging area. Confidence intervals (CI) were set at 95% and calculated through 1000 bootstrap statistics with asymptotic p-values calculated by 1000 permutations.

The differences in δ^{13} C and δ^{15} N values among serially collected samples were calculated using the first sample as a reference. The mean difference in δ^{15} N = 0.91‰ (upper & lower quantiles = 0.34 – 1.36‰, range = 0.02 – 2.50‰) and δ^{13} C = 0.61‰ (upper & lower quantiles = 0.27 – 0.81‰, range = 0.00 – 2.36‰, Fig.5 in the main text)

Supplement 6. Isotopic composition of the study population



Fig. S4. Pairwise collinearity plot for the year the sample was collected and the δ^{13} C, δ^{15} N and δ^{34} S values of the turtle epidermis for individuals included within the second discriminant analysis conducted after the 2015 satellite tracking (n = 188). All pairwise comparisons for isotopic values

were found to be significantly correlated (Pearson's product moment correlation coefficient, p < 0.001 in all cases $\delta^{13}C \& \delta^{15}N$, r = -0.26; $\delta^{15}N \& \delta^{34}S$, r = 0.23; $\delta^{13}C \& \delta^{34}S$, r = -0.75).

Supplement 7. Preliminary discriminant analysis to identify the origin of turtles from the foraging area not characterised through previous satellite tracking (1998 – 2011)

Stable isotope analyses conducted prior to satellite tracking can identify isotopic clusters to target foraging areas with specific isotopic profiles. When clusters are not evident, then groups of isotopic signatures can be selected to characterise the isotopic composition of the population. These groups can be used as pseudo-satellite tracked animals to calibrate a discriminant function analysis and obtain prior prediction probabilities for animals foraging in an area characterised by specific isotopic values.

For this study, we identified an area of isospace encompassing a large proportion of isotopic signatures which were not characterised by the pre-defined and calibrated foraging areas. To identify the origin of these isotopic values we selected 3 turtles from the 184 turtles of unknown origin that had temporal consistency in isotopic values over 2 breeding seasons and isotopically defined this region as the 'unidentified' foraging area. These turtles were used in addition to the 19 satellite tracked turtles to calibrate a discriminant analysis using $\delta^{15}N$ or $\delta^{13}C$ values. We predicted the putative foraging area for the remaining 181 turtles and produced a list of 48 turtles that were likely to forage at a greater than 80% probability in the 'unidentified' foraging area.

Supplement 8. Plots for final predictions of where turtles forage

The most discriminating isotopic criterion for turtles among foraging areas was visualised in a bivariate plot incorporating δ^{34} S and δ^{15} N (see main text Fig. 4). Here we present alternative plots incorporating the isotopic combination of δ^{13} C and δ^{15} N (S7a), δ^{13} C and δ^{34} S (S7b) and the full isotopic composition of the turtles (δ^{13} C, δ^{15} N and δ^{34} S) predicted to forage at each site (S7c).



Fig. S5. δ^{13} C and δ^{15} N values for green turtles predicted to forage in: closed circles = Bomba (n = 22), triangles = Egypt (n = 65), squares = Turkey-Cyprus (n = 11), diamonds = West Libya (n = 19), open circles = unassigned (n = 48). Ellipses set at 95% CI, (total n = 165).



Fig. S6 δ^{13} C and δ^{34} S for green turtles predicted to foraging in: closed circles = Bomba (n = 22), triangles = Egypt (n = 65), squares = Turkey-Cyprus (n = 11), diamonds = West Libya (n = 19), open circles = unassigned (n = 48). Ellipses set at 95% CI, (total n = 165).



Fig. S7. δ^{13} C, δ^{15} N and δ^{34} S for green turtles predicted to forage in: black circles = Bomba (n = 22), red circles = Egypt (n = 65), green circles = Turkey-Cyprus (n = 11), cyan circles = West Libya (n = 19) and blue circles = unassigned (n = 48), (total n = 165).

Supplement 9. Evaluating foraging area specific contributions to the breeding cohort

We employed linear and non-linear mixed effects modelling to evaluate foraging area specific contributions to the breeding cohort. We evaluated autocorrelation through generalised least squares estimation models (GLS) within the R statistical package nlme (Pinheiro et al. 2016) as a general bi-annual pattern in foraging area contributions was observed. However, only the GLS model for turtles foraging in Egypt was significantly more accurate when accounting for autocorrelation based on AICc model selection (R statistical package MuMin for multi-model selection based on information criteria). Therefore, we did not account for autocorrelation within the full model incorporating all foraging areas. We employed a generalised linear model with a quasibinomial error structure to determine if the proportion of nesters from each foraging area to the breeding cohort significantly differed among years. The model was fitted with a proportional dependent variable based on the number of nesters from each site (number of nesters from x / total number of nesters – number of nesters from x) with year (also fitted as a quadratic variable) and foraging area as interacting fixed effects. A Tukey test of Honest Significant Differences (HSD) revealed that 3 out of 6 pairwise comparisons were significantly different (Fig. S8) with Egypt exhibiting a strong positive trend in the proportion of nesters contributed to the rookery whereas the other 3 sites showed a negative trend.



Fig. S8. Prediction from the GLM for the proportions of the nesting cohort contributed from each foraging area from 1992 - 2015. Dot-dash line = Bomba, dashed line = Egypt, dotted line = Turkey-Cyprus and solid line = West Libya.

LITERATURE CITED

Broderick AC, Coyne MS, Fuller WJ, Glen F, Godley BJ (2007) Fidelity and over-wintering of sea turtles. Proc Biol Sci 274:1533–1538 PubMed doi:10.1098/rspb.2007.0211

Ceriani SA, Roth JD, Evans DR, Weishampel JF, Ehrhart LM (2012) Inferring foraging areas of nesting loggerhead turtles using satellite telemetry and stable isotopes. PLOS ONE 7:e45335 PubMed doi:10.1371/journal.pone.0045335

Godley BJ, Richardson S, Broderick AC (2002) Long-term satellite telemetry of the movements and habitat utilisation by green turtles in the Mediterranean. Ecography 25:352–362 doi:10.1034/j.1600-0587.2002.250312.x

Nakagawa S, Schielzeth H (2010) Repeatability for Gaussian and non-Gaussian data: A practical guide for biologists. Biol Rev Camb Philos Soc 85:935–956 PubMed

Pajuelo M, Bjorndal KA, Reich KJ, Vander Zanden HB, Hawkes LA, Bolten AB (2012) Assignment of nesting loggerhead turtles to their foraging areas in the Northwest Atlantic using stable isotopes. Ecosphere 3:art89doi:10.1890/ES12-00220.1

Pinheiro J, Bates D, Debroy S, Sarkar D R Core Team (2016) Linear and Nonlinear Mixed Effects Models.

Royle AJ, Rubenstein DR (2004) The role of species abundance in determining breeding origins of migratory birds with stable isotopes. Ecol Appl 14:1780–1788 <u>doi:10.1890/04-0175</u>

Seminoff JA, Benson SR, Arthur KE, Eguchi T, Dutton PH, Tapilatu RF, Popp BN (2012) Stable isotope tracking of endangered sea turtles: validation with satellite telemetry and δ^{15} N analysis of amino acids. PLOS ONE 7:e37403 <u>PubMed doi:10.1371/journal.pone.0037403</u>

Stokes KL, Broderick AC, Canbolat AF, Candan O and others (2015) Migratory corridors and foraging hotspots: critical habitats identified for Mediterranean green turtles. Divers Distrib 21:665–674 doi:10.1111/ddi.12317

Vander Zanden HB, Tucker AD, Hart KM, Lamont MM and others (2015) Determining origin in a migratory marine vertebrate: a novel method to integrate stable isotopes and satellite tracking. Ecol Appl 25:320–335 PubMed doi:10.1890/14-0581.1

Wright LI, Stokes KL, Fuller WJ, Godley BJ and others (2012) Turtle mating patterns buffer against disruptive effects of climate change. Proc Biol Sci 279:2122–2127 <u>PubMed</u> doi:10.1098/rspb.2011.2285

Wunder MB (2012) Determining geographic patterns of migration and dispersal using stable isotopes in keratins. J Mammal 93:360–367 doi:10.1644/11-MAMM-S-182.1