

Denitrification in a soft bottom lake: evaluation of laboratory incubations

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ABSTRACT: Annual benthic denitrification in a shallow soft bottom lake was estimated in laboratory-incubated sediment cores, using the isotope pairing technique. Rates were <30% of the estimate calculated from nitrogen mass balance of the lake. To investigate this discrepancy, *in situ* measurements were performed in flexible enclosures to which $^{15}\text{NO}_3^-$ was added. Accumulation of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ in the sediment and water column was measured and the loss to the atmosphere was estimated from depletion of argon, added in excess to the water of the enclosures. The calculated denitrification activities in the enclosures were 6 to 26 times higher than the activity in the incubated cores. We suggest that denitrification *in situ* was enhanced by wave forces, increasing the transport of oxygen and nitrate into these soft sediments. Such transport is not simulated by conventional stirring of the water column in laboratory incubations, which consequently may underestimate the *in situ* activity.

KEY WORDS: Rigid core tubes · Flexible enclosures · *In situ* measurements · Nitrogen mass balance

INTRODUCTION

Denitrification activity is typically measured from either large-scale nitrogen mass balances or from rates of nitrogen removal or nitrogen gas production in sampled sediments.

The nitrogen mass balance approach is commonly used in lakes and other systems, where input and output of combined nitrogen can be estimated (e.g. Messer & Brezonik 1978, Jensen et al. 1992, Windolf et al. 1996). Denitrification is calculated as the input from streams, groundwater, precipitation and nitrogen fixation minus the loss through outlets and sediment burial.

Measurements of denitrification in sampled sediments include techniques like NO_3^- depletion (Andersen 1977), acetylene block (Sørensen 1978), N_2 flux (Seitzinger et al. 1980), ^{15}N -tracer techniques (Nishio et al. 1983, Nielsen 1992) or changes in N_2/Ar ratios (Kana et al. 1994). The techniques are based on incubations of representative sediment samples, isolated in core tubes or benthic chambers (e.g. Devol 1991, Risgaard et al. 1995, Kana et al. 1998, Trimmer et al.

1998). Mixing of the water column inside these containers is typically provided by stirring devices like rods or discs. Such mixing is assumed to approach *in situ* hydrodynamic conditions.

Only a few studies compare denitrification rates obtained in sampled sediment with rates obtained from large-scale nitrogen mass balances. Good correspondence was found in a small estuary (Nielsen et al. 1995) and in 2 deep lakes (Mengis et al. 1997). In both of these studies, the isotope pairing technique (Nielsen 1992) was used to estimate the activity in laboratory-incubated sediment, and the nitrogen mass balance approach was applied to the entire study area. Andersen (1977), on the other hand, found that denitrification, measured from NO_3^- depletion in the water overlying laboratory-incubated lake sediment cores, was only about half of the values obtained by the mass balance approach. Van Luijn et al. (1996) found a similar difference when nitrogen gas production of sediment from a shallow lake was measured with the isotope pairing technique. In their study, rates obtained with the N_2 flux method, however, agreed well with the nitrogen mass balance estimate. The studies of Andersen (1977) and van Luijn et al. (1996) were conducted in shallow lakes with a mean depth ranging between

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1.6 and 5 m. Although the inconsistency between laboratory measurements and the mass balance approach might be due to shortcomings of the applied denitrification assay (cf. Seitzinger et al. 1993), it is possible that laboratory measurements in general fail in such lakes, because of inadequate simulations of the *in situ* conditions.

In the present study we wanted to investigate this possibility. By means of the isotope pairing technique we measured annual nitrogen gas production in laboratory-incubated sediment, collected in a shallow soft bottom lake. Rates obtained with that technique were compared with rates obtained from the nitrogen mass balance of the whole lake. The 2 methods produced different results, but apparently they measured actual denitrification activity in the laboratory-incubated cores and in the lake, respectively. We assumed that the laboratory incubation technique failed to simulate factors that controlled denitrification *in situ*, and the incubation technique was therefore evaluated by comparing rates of nitrogen gas production obtained in the laboratory with rates obtained in the field in flexible enclosures. Hansen et al. (1991) previously found that NO_3^- disappearance inside and outside such enclosures was similar during a 3 mo study in the same lake, which suggests that the enclosures did not change the nitrogen removal capacity of the lake sediment. In this paper we report and discuss data from both the annual measurements and from the comparison experiment.

MATERIALS AND METHODS

Study site. The study was performed in Lake Søbygaard, a small (0.39 km²), shallow (mean depth: 100 cm; maximum depth: 190 cm) hypertrophic lake in central Jutland, Denmark (56.15° N, 9.48° E). The lake is exposed to prevailing southwestern winds, but otherwise protected by slopes and forest. *Phragmites* sp. grows sparsely along the banks, while submerged vegetation is absent due to light limitation. Phytoplankton production is 275 ± 9 mmol C m⁻² d⁻¹ (Jeppesen et al. 1997) and Secchi depth ranges between 55 and 85 cm (Jeppesen et al. 1990).

Up to 90% of the water is supplied from a nitrogen-rich creek flowing into the east end of the lake. Groundwater fed springs are responsible for the remainder. The hydraulic residence time is 18 to 27 d (Jensen et al. 1992), and the nitrogen load for the 1985–1995 period was 11.4 ± 1 mmol N m⁻² d⁻¹ (Jeppesen et al. 1998). The sediment of the lake is organic-rich (25 to 35% of sediment dry weight is organic matter in the upper centimeter) soft mud with a porosity of 0.92 (vol./vol.).

Nitrogen gas production in sampled sediment cores. Rates of denitrification were measured with the

¹⁵N isotope pairing technique in intact sediment cores collected 8 separate times during the May 1992 to May 1993 period. Measurements were only performed in darkness, as no light reached the sediment surface *in situ*. The cores (n = 4 or 5) were sampled at the central part of the lake (depth: 150 cm) in Plexiglas tubes (inner diameter: 3.6 cm, height: 20 cm). Filtered (glass fiber GF/C filters) water samples for $\text{NO}_3^- + \text{NO}_2^-$ analysis were collected at the field site in 20 ml polyethylene vials and frozen (–20°C) on return to the laboratory. Lake water for the incubations was collected in 50 l polyethylene jars.

In the laboratory the height of the sediment was adjusted to ~5 cm, and the tubes were hereafter placed in an open tank with lake water, held at *in situ* temperature. A 1 cm long Teflon-coated rotating (60 rpm) magnet, mounted inside the tubes, kept the water column above the sediment stirred. ¹⁵N-labeled NO_3^- (50 to 100 μM) was then added to the water in the tank, and this ¹⁵N-enriched lake water was carefully mixed with the water above the sediment cores. The resulting ¹⁵N enrichment of the NO_3^- pool in the water column ranged from 20 to 99 ¹⁵N atom% due to seasonal variations of the *in situ* NO_3^- concentration (see Fig. 5).

Approximately 10 min after tracer addition, the core tubes were sealed with rubber stoppers. The incubation of the individual cores was terminated at regular time intervals within 1.7 h (summer) or 9 h (winter) of incubation. Total incubation time, determined after preliminary measurements of the sediment O₂ demand, was set to ensure that change in O₂ never exceeded 20% of the initial O₂ concentration. The incubation of the cores was terminated by adding 0.5 ml 7 M ZnCl₂ and subsequently mixing the sediment and the water column with a PVC rod. A sample of the resultant sediment-water suspension was collected with a syringe in order to determine the amount of ¹⁵N₂ that accumulated in both the sediment and the water column during the incubation. This sample was transferred to 12 ml glass vials (Exetainers, Labco, High Wycombe, UK), preserved with an additional 250 μl 7 M ZnCl₂ and stored at room temperature for later ²⁹N₂ and ³⁰N₂ determinations.

During June, August and September 1992 denitrification was measured at 2 different ¹⁵NO₃⁻ concentrations in the water overlying the sediment. These measurements were performed to verify the fundamental requirements of the isotope pairing technique, i.e. first order dependency of denitrification with respect to NO_3^- concentration in the water column and uniform mixing of ¹⁴NO₃⁻ and ¹⁵NO₃⁻ in the denitrification zone (Nielsen 1992).

To verify that the station selected for annual denitrification measurements was representative for the

whole lake, denitrification was measured at 3 different sites during October 1992: the east end (depth: 65 cm), the west end (depth: 100 cm), and at the station described above.

Nitrogen gas production in field enclosures. During May and September 1996, denitrification activity was measured with the isotope pairing technique both in air-exposed enclosures installed in the lake and in laboratory-incubated cores. Field and laboratory incubations were conducted under similar nutrient, O_2 and temperature conditions. The temperature of the water column was 13°C , O_2 was close to saturation ($297 \mu\text{M}$), and NO_3^- concentration in the water column was below $1 \mu\text{M}$.

The enclosure (inner diameter: 150 cm; height: 120 cm) was constructed from a transparent flexible PVC bag, fixed to a metal cylinder (Fig. 1). The day before incubation was initiated, the enclosure was placed in the western part of the lake at a water depth of ~ 85 cm. The metal cylinder was forced into the sediment and fixed to wooden poles, while the PVC bag was left below the water surface, to allow free exchange with ambient lake water. Just before the incubation began, the upper rim of the bag was extended ~ 40 cm above the surface of the water and tightened to the poles. Wave amplitude and frequency in the enclosure and the lake were similar, as judged from visual observations.

Argon-saturated lake water (15 l) with $^{15}\text{NO}_3^-$ was gently mixed into the water of the enclosure without re-suspending the sediment. The ^{15}N isotope was added to a final concentration of $100 \mu\text{M}$ in May and to $30 \mu\text{M}$ in September. The high initial working concentration of $^{15}\text{NO}_3^-$ in May was chosen to prevent phytoplankton from exhausting the tracer. In order to determine rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ production, it was necessary to determine both the amount of $^{15}\text{N}_2$ that accumulated in the enclosed water column plus the sediment and the amount of $^{15}\text{N}_2$ that was lost to the atmosphere during the incubation. Evolution of $^{29}\text{N}_2$ and $^{30}\text{N}_2$, and depletion of Ar in the enclosure were therefore followed for 24 h, by collecting samples from the entire water column and subsamples of the sediment plus the water column, for later gas determinations.

Water samples from the entire water column of the enclosure were collected in duplicate with a 100 cm long 0.3 cm i.d. glass tube. The glass tube was vertically immersed in the water column so gently that the meniscus always stayed at the lake surface. Just before the tip of the tube reached the bottom of the lake, the upper end was closed with a finger, and the tube, containing a sample of the intact water column, was redrawn from the enclosure. Additional samples of the bottom water were collected with the upper tube end sealed until a few cm above the bottom. Sampling of the bottom water was performed to investigate if the

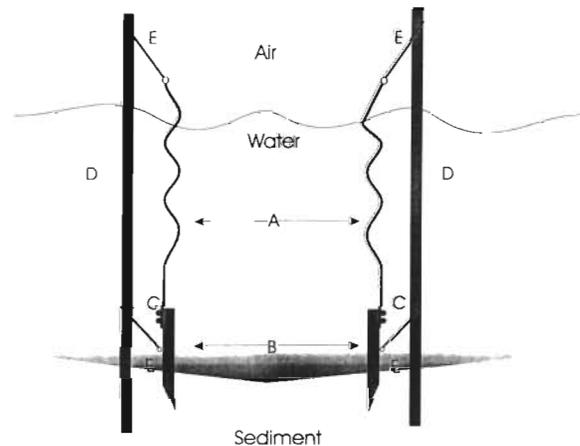


Fig. 1. Enclosure used for field incubations. A PVC bag (A) is fixed to a metal cylinder (B) with nylon strips (C). The enclosure is fixed to wooden poles (D) by wires (E)

water column was stratified or mixed during the incubation. All water samples were transferred to 12 ml glass vials (Exetainer, Labco) and preserved with $250 \mu\text{l}$ 7 M ZnCl_2 . The samples were stored for less than 5 d in the laboratory before the dissolved gases were determined.

Subsamples of sediment plus the water column were collected as intact cores in triplicate with a Plexiglas tube (inner diameter: 5.5 cm; height: 100 cm). The tube was forced ~ 10 cm into the sediment, sealed with a rubber stopper, and then redrawn from the enclosure. After adding 1 ml 7 M ZnCl_2 to the cores, the water and the sediment were mixed and a sample of the slurry was transferred to 12 ml glass vials (Exetainer, Labco) and preserved with additional $250 \mu\text{l}$ 7 M ZnCl_2 . Like the samples from the water column, the sediment-water samples were stored for less than 5 d in the laboratory before the gas concentration was determined.

Intact cores ($n = 6$), for the laboratory measurements, were collected in Plexiglas tubes (inner diameter: 5.2 cm; height: 30 cm) close to the enclosure. The cores were allowed to equilibrate in the laboratory for 12 h in a darkened tank with air-saturated lake water, before incubation was initiated. The water column above the sediment was stirred with a 2.5 cm Teflon-coated magnet (60 rpm), positioned about 5 cm above the sediment surface. Denitrification measurements were performed as described in the previous section.

Nitrogen mass balance of the lake. Denitrification rates, calculated from the nitrogen mass balance of the lake from May 1992 to May 1993, were provided by J. P. Jensen (National Environmental Research Institution, Silkeborg, Denmark, pers. comm.). The nitrogen mass balance was based on frequent (6 h) measurements of nitrogen concentrations (DIN and DON) in the inlet and outlet of the lake, daily nitrogen measure-

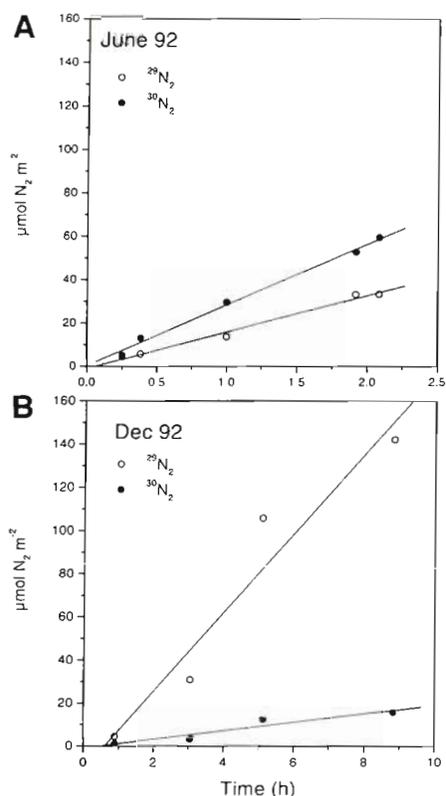


Fig. 2. Linear accumulation of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ in the sediment and the water column of laboratory-incubated samples during a summer (A: June 1992, $r^2 = 0.99$) and a winter (B: December 1992, $r^2 > 0.90$) incubation. Each paired $^{29}\text{N}_2/^{30}\text{N}_2$ point represents data from a single core. Note different scales on the time axes in June and December

ments in the mid lake, and approximately monthly measurements in the springs. The transport of nitrogen was calculated by multiplying values of nitrogen concentration with measured values for daily water discharge. Quarterly denitrification was estimated from differences between nitrogen load, washout, storage and burial. Storage was calculated from differences in nitrogen content of the lake water during yearly or quarterly periods, and burial was calculated from iron mass balances and the iron to nitrogen ratio in the sediment. For further details about the mass balance of the lake see Jensen et al. (1992)

Analyses. Gas samples of 250 μl for Ar, $^{29}\text{N}_2$ and $^{30}\text{N}_2$ determinations were extracted from the water samples in the glass vials as described by Risgaard-Petersen & Rysgaard (1995), and subsequently analyzed on a GC-MS system (Sira Series II, VG Isotech, Middlewich, UK), described by Nielsen (1992). Gas samples extracted from air-saturated deionized water samples were used as reference.

Lake water $\text{NO}_3^- + \text{NO}_2^-$ concentrations were determined on a Flow Injection Analyzer (Tecator, Hö-

ganäss, Sweden), using the sulphanilamide-naphthylene-diamide method after reduction of NO_3^- to NO_2^- by a cadmium column (Grasshoff et al. 1983).

Calculations. Rates of denitrification of *in situ* NO_3^- ($^{14}\text{NO}_3^-$) were determined using the principle of isotope pairing (Nielsen 1992). It was assumed that the added $^{15}\text{NO}_3^-$ mixed homogeneously with the $^{14}\text{NO}_3^-$ pool in the sediment and that the 2 NO_3^- species were reduced at a rate proportional to their respective mole fractions. Denitrification of *in situ* NO_3^- could then be calculated from the production rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ using the equation of Nielsen (1992):

Denitrification of *in situ* $\text{NO}_3^- =$

$$\frac{F^{29}}{2 \times F^{30}} \times (F^{29} + 2 \times F^{30}) \quad (1)$$

where F^{29} and F^{30} are the production rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$.

Rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ production in the core tubes were calculated from the slope of the regression line, obtained from plots of $^{29}\text{N}_2$ or $^{30}\text{N}_2$ ($\mu\text{mol m}^{-2}$) versus time (h); see Fig. 2.

Production rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ in the enclosure, installed in the lake, were calculated from $^{15}\text{N}_2$ accumulation in the sediment and the water column plus the amount lost to the atmosphere (Fig. 3). The water column in the enclosure was well mixed in May and September 1996, as indicated by identical concentrations of $^{15}\text{N}_2$ or Ar in the bottom and surface water (*t*-test, hypothesized mean difference = 0, $p \gg 0.05$, data not shown). The loss of $^{15}\text{N}_2$ to the atmosphere was therefore proportional to the difference between the actual concentration in the water column [$^{15}\text{N}_w(t)$] and the concentration at air saturation ($^{15}\text{N}_o$), as shown by Broecker & Peng (1974):

$$\text{Loss} = a[^{15}\text{N}_o - ^{15}\text{N}_w(t)] \quad (2)$$

where a is the air-water transfer coefficient for N_2 . The air-water transfer coefficient for N_2 was calculated as the transfer coefficient for Ar multiplied with the ratio between the diffusion coefficients for N_2 and Ar

Table 1. Denitrification activity measured with different concentrations of $^{15}\text{NO}_3^-$ in the water overlying the sediment. The standard error of the mean is presented in parentheses ($n = 4$)

Date (1992)	Concentration of added $^{15}\text{NO}_3^-$ (μM)	Denitrification of <i>in situ</i> $^{14}\text{NO}_3^-$ ($\mu\text{mol m}^{-2} \text{h}^{-1}$)	Denitrification of added $^{15}\text{NO}_3^-$ ($\mu\text{mol m}^{-2} \text{h}^{-1}$)
16 Jun	49	19.7 (1.7)	70 (4)
	85	23 (2)	120 (20)
11 Aug	47	15.6 (1.2)	26 (3)
	106	12.1 (1.1)	39 (2)
17 Sep	41	66 (8)	13.2 (1.4)
	88	71.9 (1.7)	37 (3)

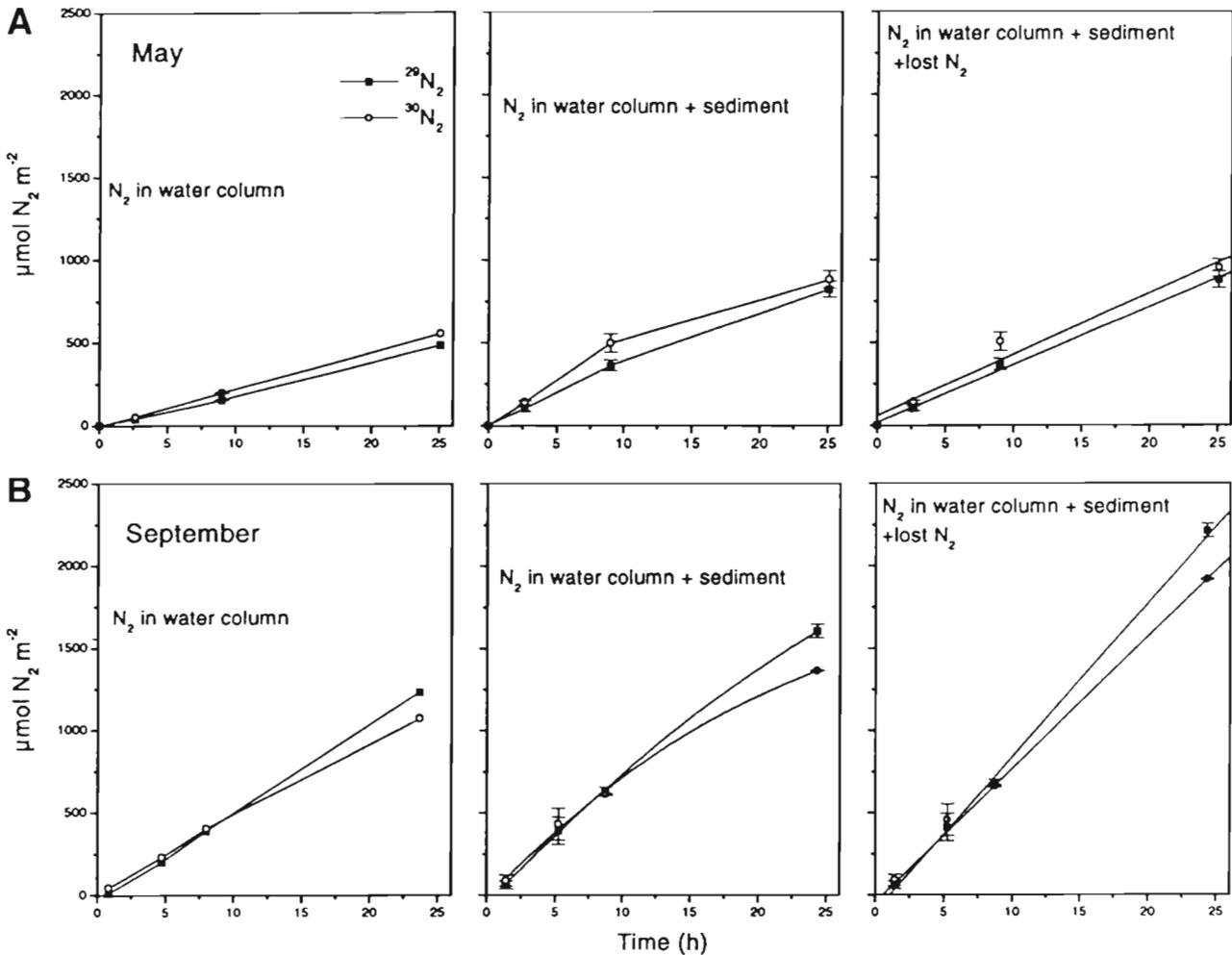


Fig. 3. Accumulation of $^{15}\text{N}_2$ in the water column of the field enclosure, in the sediment plus the water column, and in the sediment, the water column plus the amount of $^{15}\text{N}_2$ lost to the atmosphere in (A) May and (B) September. Error bars represent standard error of the mean (n = 3)

(Broecker & Peng 1974). The transfer coefficient for Ar was calculated from Ar depletion from the enclosure, assuming a first order decrease in concentration with time (Fig. 4).

RESULTS

The isotope pairing technique adapted to core-tube incubations

The evolution of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ in the $^{15}\text{NO}_3^-$ -amended core tubes could be described as a linear function of time ($0.87 < r^2 < 0.99$ for all incubations; see Fig. 2 for examples), suggesting constant gas production during the incubation period. The rate of denitrification of $^{15}\text{NO}_3^-$ increased when $^{15}\text{NO}_3^-$ was elevated, but denitrification of *in situ* NO_3^- ($^{14}\text{NO}_3^-$) was inde-

pendent of the $^{15}\text{NO}_3^-$ concentration in the water column (Table 1). This suggests first order dependency of denitrification with respect to NO_3^- , and homogenous mixing of $^{14}\text{NO}_3^-$ and $^{15}\text{NO}_3^-$ in the denitrification zone (cf. Nielsen 1992).

Denitrification rates at the 3 different sites in the lake were similar (east end: 2.7 ± 0.33 [mean \pm SE], center: 2.1 ± 0.28 , west end: 2.3 ± 0.24 $\text{mmol N m}^{-2} \text{ d}^{-1}$, hypothesized mean difference = 0, $p \gg 0.05$). This indicates a homogeneous sediment environment and justifies that only a single site was selected for the annual denitrification measurements.

Denitrification activity, measured as nitrogen gas production in the laboratory during 1992 to 1993, showed a marked seasonal variation (Fig. 5). Maximum rates (~ 2.4 $\text{mmol N m}^{-2} \text{ d}^{-1}$) were observed during winter and late fall when NO_3^- in the water column peaked. Lowest activity (< 0.3 $\text{mmol N m}^{-2} \text{ d}^{-1}$) was

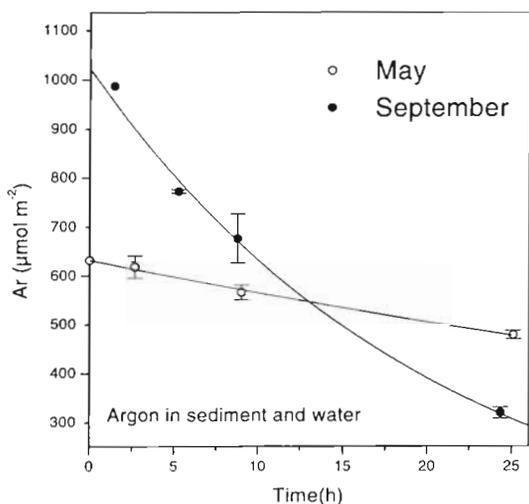


Fig. 4. Decrease of excess argon in the sediment and the water column of the enclosure. Error bars represent standard error of the mean ($n = 3$). In May the transfer coefficient was $-0.011 \pm 0.002 \text{ h}^{-1}$, assuming exponential decay. In September the transfer coefficient was $-0.048 \pm 0.003 \text{ h}^{-1}$.

observed during summer months when NO_3^- was $< 1 \mu\text{M}$. Annual denitrification was approximately 30% of the activity estimated by the nitrogen mass balance approach (511 ± 18 vs $1880 \text{ mmol N m}^{-2} \text{ yr}^{-1}$). According to the laboratory measurements of nitrogen gas production, less than 13% of the nitrogen load to the lake ($4067 \text{ mmol N m}^{-2} \text{ yr}^{-1}$ for the 1992 to 1993 period, J. P. Jensen pers. comm.) was lost by denitrification. The nitrogen mass balance approach, however, proposed that denitrification removed approximately 46% of the nitrogen input.

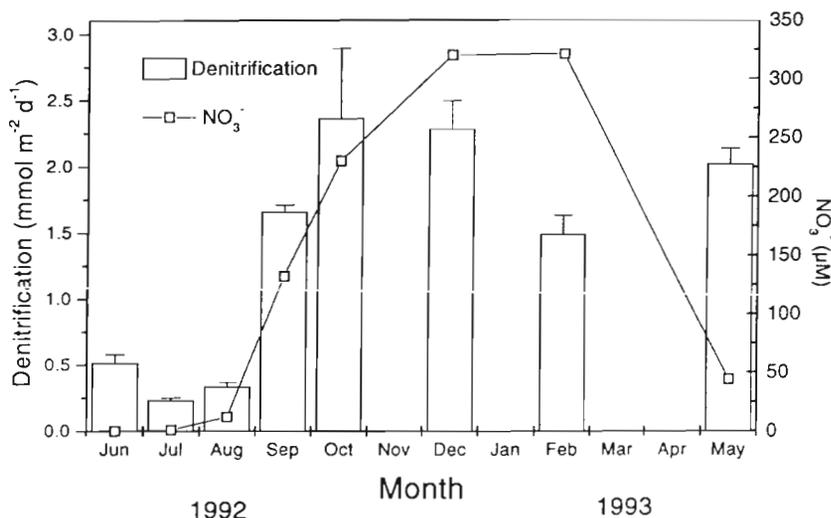


Fig. 5. Annual denitrification activity estimated in laboratory-incubated sediment and lake water NO_3^- concentration. Columns represent the mean denitrification activity ($n = 4$); error bars represent standard error of the mean.

The isotope pairing technique adapted to field incubations

Linear $^{29}\text{N}_2$ and $^{30}\text{N}_2$ evolution with time was observed during both the May and the September incubations ($r^2 > 0.97$), when the amount of $^{15}\text{N}_2$ lost to the atmosphere was included in the pool of produced gas (Fig. 3). This shows that rates of $^{29}\text{N}_2$ and $^{30}\text{N}_2$ production were constant during the 24 h of incubation. Less than 7% of the $^{15}\text{N}_2$ gas produced in the enclosure during the May 1996 experiment was lost during the incubation, but in September approximately 30% of the produced $^{15}\text{N}_2$ gas was lost from the enclosure (Fig. 3). This difference was probably caused by different wind velocities, regulating the air-water exchange of gases. The average wind velocities, measured at the nearest meteorological station, Ødum II, Denmark (56.18° N , 10.8° E), were $2.8 \pm 0.3 \text{ m s}^{-1}$ during the May measurements and $5.2 \pm 0.7 \text{ m s}^{-1}$ during the September measurements. Although Ødum is situated approximately 20 km northeast from Lake Søbygaard and local wind conditions may differ, the 4-fold higher air-water gas exchange rates in September compared to May (Fig. 4) are consistent with a 2-fold higher wind velocity, since the rate of exchange is roughly proportional to the square of the wind velocity (Kanwisher 1963).

In May 1996 denitrification rates measured in the enclosure installed in the field exceeded laboratory-generated rates by a factor of ~ 6 , whereas in September 1996, when the wind velocity was highest, the activity in the enclosure was ~ 26 times higher than the activity measured in the laboratory-incubated sediment cores (Fig. 6). Nitrate concentrations in the field enclosures and in the laboratory-incubated cores were $< 1 \mu\text{M}$ both in May and in September. The rates obtained in the laboratory in 1996 were similar to the rates in 1992 to 1993, when NO_3^- concentrations in the water column were $< 1 \mu\text{M}$.

DISCUSSION

In the present study we found a significant difference between denitrification rates measured as nitrogen gas production in laboratory-incubated sediment and rates estimated from the nitrogen mass balance of the lake. The following hypotheses explaining this difference were evaluated: (1) The denitrification assay failed to measure factual nitrogen gas production. (2) The nitrogen mass balance

obtained incorrect estimates of denitrification. (3) The sampled sediment was not representative of the lake. (4) The laboratory incubation system failed to simulate factors that control denitrification in the field.

Hypothesis 1: In a parallel experiment with sediment from this lake we have shown that the isotope pairing technique and the N_2 flux method provided identical and reliable estimates of the denitrification activity (Risgaard-Petersen et al. 1998).

The tests performed in the present study also confirmed that potential artifacts associated with the denitrification assay can be excluded. If non-homogeneous mixing of $^{15}NO_3^-$ and $^{14}NO_3^-$ in the sediment was a significant problem for the denitrification assay, then the estimated *in situ* denitrification activity would increase when the $^{15}NO_3^-$ concentration was elevated, because more of the activity would be detected as measurable $^{29}N_2$ production (Nielsen 1992). Our data showed, however, that the estimate of the *in situ* denitrification activity was independent of the amount of added $^{15}NO_3^-$ (Table 1).

The indication that denitrification showed first order dependency of NO_3^- (Table 1) showed that the process was NO_3^- limited. This confirmed that tracer additions did not interfere with the estimate of the *in situ* denitrification activity.

Constant production of $^{29}N_2$ and $^{30}N_2$, as indicated by linear $^{29}N_2$ and $^{30}N_2$ evolution with time (Figs. 2 & 3), showed that the incubation time was sufficiently short to avoid significant depletion of the tracer and sufficiently long compared to the initial non-steady-state period when the $^{15}NO_3^-$ gradient is established in the sediment.

All tests performed in this study and the in study of Risgaard-Petersen et al. (1998) therefore indicate that the isotope pairing technique correctly measured actual denitrification in the laboratory-incubated sediment from Lake Søbygaard.

Hypothesis 2: The central parameters used in the mass balance model, e.g. water balance, nitrogen load, nitrogen export, and nitrogen burial, are all well described for this lake (Jensen et al. 1992), which justifies the denitrification estimate determined by this method. Since nitrogen burial in the lake is insignificant (Jensen et al. 1992), the indication by the mass balance approach of a 46% removal of the nitrogen input to the lake via denitrification is furthermore consistent with general trends for lakes and estuaries. Nixon et al. (1996) developed an empirical model to predict the percentage of total nitrogen input that is exported from lakes and estuaries from the hydraulic residence time Q (unit for Q is month):

$$\% \text{ total nitrogen exported} = -27.0 \times \log(Q) + 64.8 \quad (3)$$

The average hydraulic residence time for the lake in 1992 to 1993 was 0.83 mo (J. P. Jensen pers. comm.),

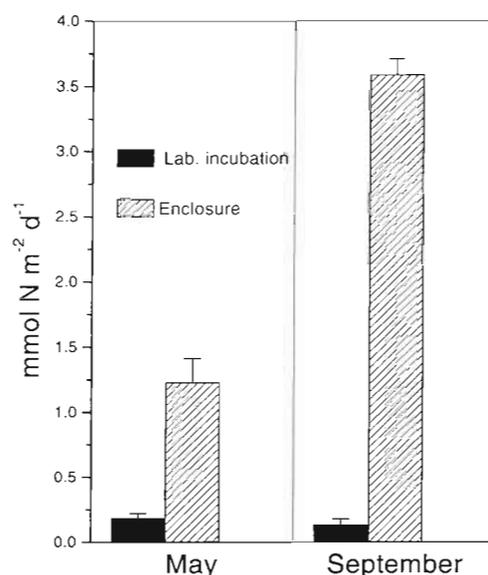


Fig. 6. Denitrification measured in the enclosure in the field and in the laboratory with the core-tube incubation technique. Error bars represent standard error of the mean ($n = 6$ for the core-tube incubations; for the enclosures standard error is calculated from the regression data)

and according to Eq. (3) 33% of the nitrogen input to the lake would be removed (% total nitrogen exported = 66.9). We therefore believe that the mass balance approach obtained reliable estimates of the denitrification activity in the lake.

Hypothesis 3: Results from our investigation of the spatial heterogeneity of denitrification activity in the lake sediment were not consistent with the hypothesis that non-representative sediment sampling was responsible for the divergence between laboratory measurements and the mass balance estimate. Similar denitrification rates were measured at the 3 sites in the lake, suggesting that the lake had a homogenous sediment environment.

Hypothesis 4: The observation that the denitrification activity measured in the field in the enclosures was 6 to 26 times higher than the activity measured in the laboratory-incubated cores, using the same methodology (Fig. 6), supports the hypothesis that the laboratory incubation system failed to simulate factors that control the *in situ* activity. We attribute these factors to mechanisms other than molecular diffusion controlling benthic solute exchange. Previous data of Søndergaard (1990) indicate that benthic solute exchange in the lake is controlled by such mechanisms. Søndergaard found both enhanced gross phosphorus release from the lake sediment compared to what could be estimated from porewater profiles, and rapid changes in the phosphate porewater profiles after storms.

A mechanism responsible for benthic solute exchange in the lake might be porewater transport, induced by surface waves or the current. Movements of waves at the surface of the overlying water induce pressure changes which are transmitted into the sediment (Sleath 1984), and these repeated pressure changes generate flows of porewater in permeable sediments (Rutgers van der loeff 1981, Shum 1992). Interaction between water currents and the sediment topography may also generate pressure gradients and concurrent porewater flow (Huettel et al. 1996). This porewater flow acts in addition to molecular diffusion and can therefore enhance sediment-water exchange of solutes (Malan & McLachlan 1991, Forster et al. 1996, Zeibis et al. 1996). Presence of wave- or current-induced porewater flow is likely to enhance benthic denitrification, because the process is dependent on the rate of NO_3^- transport towards the anoxic denitrification zone (Seitzinger 1988, Christensen et al. 1989). If wave-induced porewater flow was significant in the sediment of the lake, highest denitrification activity would be expected in sediments incubated in the enclosures, since such porewater flow might act in the flexible field enclosures used in the present study, but evidently not in sediments isolated in rigid core tubes. The largest discrepancy between estimates obtained in the laboratory and in the field would furthermore be expected in periods with relatively high wind velocity, because surface waves are developed due to the friction between water and wind in the initial stage, and consequently larger pressure changes are transmitted into the sediment at high wind velocities than at low wind velocities. Both of these trends were observed in the present study (Fig. 6).

In the present study we have shown that the nitrogen gas production approach may obtain lower estimates of the denitrification activity than the mass balance approach. Our results are in accordance with previous studies showing that laboratory measurements may underestimate *in situ* denitrification activity in shallow lakes (Andersen 1977, van Luijn et al. 1996). Our data strongly indicate that this underestimation can be due to under-representation of *in situ* transport processes in the rigid core tubes used for laboratory incubations, and we attribute these transport processes to wave-induced porewater transport. However, although this hypothesis appears attractive, we want to emphasize that the exact regulation of benthic denitrification in the lake is far from elucidated in this study. Indeed, more detailed studies on the effect of physical processes (waves, current, etc.) on the denitrification process are needed.

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