Effects of Saharan dust on the microbial community during a large *in situ* mesocosm experiment in the NW Mediterranean Sea

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ABSTRACT: The response of the microbial community to Saharan dust deposition was investigated in 6 large mesocosms (52 m³) deployed at an oligotrophic coastal site in the NW Mediterranean Sea in June 2008 (DUNE project). The mesocosms represented well the environmental conditions observed at the study site during the 8 d experimental period, and the triplicate mesocosms exhibited high reproducibility for each treatment. Dust deposition resulted in an increase in chlorophyll a concentration (0.22 ± 0.03 µg l⁻¹), as compared to that in the control treatments (0.12 ± 0.01 µg l⁻¹), but no treatment effect was observed for bacterial heterotrophic abundance at 5 m depth. Results from the fingerprinting technique CE-SSCP indicate a temporal evolution of the structure of the total (16S rRNA gene) and active (16S rRNA transcripts) bacterial community, and Saharan dust deposition had a noticeable structuring effect on the active bacterial community. Combining results from 16S rRNA gene clone libraries and CE-SSCP indicates that the relative contribution of *Alteromonas macleodii* to the active bacterial community was enhanced 2-fold following dust addition. The 2 operational taxonomic units (OTUs) *Thiothrix* and *Alteromonas*, belonging to *Gammaproteobacteria*, and the *Bacteroidetes* OTU NS5-1 were specific to the clone libraries from the dust-amended mesocosms or more abundant in these than in the control ones. CARD-FISH analyses, however, indicate that these OTUs had overall low abundances (1 to 5% of total DAPI-counts). Despite the pronounced temporal trend observed during the experimental period, dust deposition had a small, but noticeable structuring effect on the heterotrophic bacterial community that was detectable only at the OTU level at 99% similarity of the 16S rRNA gene.

KEY WORDS: Saharan dust deposition · *In situ* mesocosms · CE-SSCP · CARD-FISH · Clone libraries · *Alteromonas macleodii* · NW Mediterranean Sea

INTRODUCTION

Dry and wet atmospheric deposition represents a significant source of inorganic and organic nutrients and organic carbon to the ocean (Jickells et al. 2005, Jurado et al. 2008). In high-nutrient low-chlorophyll regions, dust deposition enhances iron availability and thus stimulates phytoplankton primary production and potentially carbon dioxide drawdown (Boyd et al. 2007, Cassar et al. 2007). In low-nutrient low-chlorophyll (LNLC) regions, the simultaneous input of phosphate and iron from dust could create a favourable environment for enhanced dinitrogen fixation (Mills et al. 2004). In the subtropical northeast Atlantic Ocean,
dust addition markedly stimulated phytoplankton primary production and induced a shift from *Cyanobacteria* to diatoms, while the response in microbial community respiration was minor, resulting in an increase in the ratio of primary production to respiration (Duarte et al. 2006). In contrast, estuarine heterotrophic bacterial and phytoplankton primary production were both stimulated by organic nitrogen introduced by rainfall (Seitzinger & Sanders 1999). These studies demonstrate that atmospheric inputs can impact autotrophic and heterotrophic microbial communities with important consequences for ocean biogeochemistry.

In the Mediterranean Sea, which is during the stratified period mostly a LNLC region, Saharan dust events represent a dominant source of atmospheric deposition. Previous studies have demonstrated that, during the summer stratification period, atmospheric dust is the main external source of phosphorus (Ridame & Guieu 2002) and of iron (Bonnet & Guieu 2006) in the surface mixed layer. Microcosm experiments performed in coastal and offshore Mediterranean waters have shown that dust stimulated primary production (Bonnet et al. 2005, Herut et al. 2005, Lekunberri et al. 2010) and is suspected to stimulate nitrogen fixation (Bonnet & Guieu 2006). Direct observations of natural dust deposition events in the eastern (Herut et al. 2005) and western (Pulido-Villena et al. 2008) Mediterranean Sea and in a Mediterranean reservoir (Reche et al. 2009) revealed increases in the abundance and metabolism of autotrophic and heterotrophic microbial populations.

While the observed response of the phytoplankton community was linked in some studies to changes in major functional groups (Herut et al. 2005, Duarte et al. 2006), little is known about the effect of dust addition on the structure of the heterotrophic bacterial community. Using microcosm experiments with water from a Mediterranean reservoir, Reche et al. (2009) observed no noticeable differences in the bacterial diversity and richness between dust-amended and control treatments. Conversely, dust addition induced changes in the bacterial community composition in microcosms performed with coastal Mediterranean waters (Lekunberri et al. 2010). The only study that has thus far investigated the impact of dust on the activity of specific bacterial groups reports that instantaneous metabolic rates of *Prochlorococcus* and a bacterial subpopulation dominated by SAR11 were reduced upon dust addition in the North Atlantic Ocean (Hill et al. 2010). This study suggests that the community response was linked to changes in the microbial community structure (Hill et al. 2010). The question of whether the response of specific bacterial groups to dust addition could play an important role in the cycling of elements introduced by dust, however, remains poorly understood.

The objective of the present study was to investigate the impact of dust deposition on the structure of the total and active bacterial community and to identify the bacterial groups that were most responsive to dust addition. We used, for the first time in this context, large trace-metal-clean *in situ* mesocosms that allowed us to test for dust deposition effects close to *in situ* conditions in LNLC waters.

**MATERIALS AND METHODS**

**Study site and sample collection.** The results presented here were obtained during the first field campaign of the DUNE project (A DUS experiment in a low Nutrient, low chlorophyll Ecosystem, www.obs-vlfr.fr/LOV/DUNE). A detailed description of the mesocosms, the collection and treatment of the Saharan dust, the dust deposition and the sampling is given by Guieu et al. (2010). Briefly, Saharan dust was collected in Tunisia, a dominant source region of atmospheric dust for the NW Mediterranean Sea. Saharan dust was submitted to mechanical and physicochemical transformations before deposition in the mesocosms. The finest fraction (<20 µm) was used for the experiment since dust particles larger than 20 µm are rapidly removed during atmospheric transport (Maring et al. 2003). In the present study, the fertilization mimicked a wet deposition event using evapocondensed dust. Six large mesocosms, each containing 52 m³ of raw seawater and extending to 15 m depth, were deployed at a coastal site in the NW Mediterranean Sea (Bay of Elbo, Corsica, France: 42°22’ N, 8°33’ E) in June 2008. Of the 6 mesocosms, 3 were amended with each 10 g m⁻² of Saharan dust simulating a relatively strong atmospheric dust event (hereafter referred to as D-mesocosms), and 3 mesocosms were established as controls (hereafter referred to as C-mesocosms). Samples from the 6 mesocosms were taken at 5 m depth before dust addition (t₀), and after 3 d (t₃), 6 d (t₆) and 8 d (t₈) after the amendment. Real-time measurements revealed that several biogeochemical parameters remained stable between t₃ and t₈; therefore, the experiment was terminated after 8 d. Samples were collected from a floating platform, using a Teflon pump. Throughout the experiment, seawater was also collected outside the mesocosms (hereafter referred to as OUT, n = 1) at a distance of about 50 m. Water temperature was 19.8 ± 0.5°C inside all mesocosms throughout the experiment, as was the water temperature outside the mesocosms.

**Enumeration of heterotrophic bacteria, autotrophic prokaryotic and eukaryotic cells.** The abundance of heterotrophic bacteria, autotrophic prokaryotic and eukaryotic cells was determined by flow cytometry.
according to previously described protocols (Lebaron et al. 1998, Marie et al. 2000). Subsamples (2 ml for heterotrophic bacteria and 4 ml for autotrophic cells) were fixed with 2% formaldehyde (final conc.), incubated for 30 min at 4°C, frozen in liquid nitrogen and stored at −80°C until analysis. Synechococcus spp. were discriminated by their strong orange fluorescence (>630 nm) and pico- and nanoeukaryotes were discriminated by their scatter signals of the red fluorescence at 585 ± 21 nm. For heterotrophic bacteria, samples were stained with SYBR Green I (Invitrogen Molecular Probes) at 0.025% (vol/vol) final concentration for 15 min at room temperature in the dark. Counts were performed with a FACSCalibur flow cytometer (Becton Dickinson) equipped with an air-cooled argon laser (488 nm, 15 mW). Stained bacterial cells, excited at 488 nm, were discriminated and enumerated according to their right-angle light scatter (SSC, related to cell size) and green fluorescence measured at 530 ± 30 nm.

Concentration of chlorophyll a. To determine the concentration of chlorophyll a, 2 l of raw seawater were filtered on GFF filters (25 mm diameter). Filters were extracted in acetone, and the concentration of chlorophyll was determined by fluorometry on a Trilogy Laboratory Fluorometer (Turner Designs).

Nucleic acid extraction. To determine the bacterial community structure by the fingerprinting method CE-SSCP, raw seawater (0.5–1 l) was first prefiltered through a 3 µm pore size filter (47 mm, Nucleopore), and the <3µm filtrate sequentially filtered through a 0.8 µm PC filter (47 mm, Nucleopore), and a 0.2 µm PC filter, respectively, using a peristaltic pump. The 0.8 µm and 0.2 µm PC filtrates were stored frozen at −80°C until analysis. For both size fractions, the structure of the total (16S rRNA genes) and active (16S rRNA transcripts) bacterial communities was determined, based on the DNA and RNA extracts, respectively.

For DNA and RNA extraction the PC filters were submerged in 425 µl lysis buffer (40 mM EDTA, 50 mM Tris, 0.75 M sucrose) and then subjected to 3 freeze-thaw cycles using liquid nitrogen and a water bath at 65°C. Freshly prepared lysozyme solution (1 mg ml⁻¹) was added to the PC filters and incubated at 37°C for 45 min. Proteinase K (0.2 mg ml⁻¹) and sodium dodecyl sulphate (SDS 1%) were then added and the PC filters were incubated at 55°C for 1 h. Purification of DNA and RNA was performed with the Qiagen AllPrep DNA/RNA extraction kit. 1.5 ml RLT+ buffer (Qiagen) containing 14.3 M β-mercaptoethanol was added, mixed well and incubated for 10 min at room temperature. The DNA and RNA were purified separately from the lysate on the individual columns according to the manufacturer's instructions. The molecular size and the purity of the DNA were analysed by agarose gel electrophoresis (1%).

Reverse transcription of RNA. RNA samples (2 µg) were incubated together with 0.5 ml primer w34 (1 mM final concentration) at 94°C for 5 min before cooling rapidly in a water/ice bath. Reverse transcription was achieved by adding 5 ml of deoxynucleoside triphosphates (dNTPs; 0.8 mM final concentration), 1 ml of Moloney murine leukemia virus (M-MLV) reverse transcriptase (20 U) and 5 µl of 5X M-MLV buffer (Promega) and allowing the reaction to proceed at 42°C for 1 h. The cDNA samples were stored at −20°C.

Capillary electrophoresis – single-strand conformation polymorphism analysis (CE-SSCP). CE-SSCP is a fingerprinting method that allows an image to be obtained of the structure of the total (16S rRNA genes) and active (16S rRNA transcripts) bacterial community. CE-SSCP enables determination of the presence and relative abundance of up to 40 of the most abundant operational taxonomic units (OTUs) of varying taxonomic ranks (Lami et al. 2009, West et al. 2008). Short fragments (~200 bp) of the V3 region of the 16S rRNA gene were amplified from DNA or from a cDNA copy of the RNA using the bacterial specific primers w49 (5′-ACG GTC CAG ACT CCT ACG GG-3′) (Delbes et al. 2000) and w34 (5′-TTA CCG CGG TGC TGG CAC-3′) (Lee et al. 1996), the latter 5′ labelled with phosphoramidite (TET, Eurogentec). Amplifications were performed according to a previously described protocol (West et al. 2008). The electropherograms were analysed by GeneScan software (Applied Biosystems), using the second-order least square size calling method to normalize mobilities between different runs as described previously (Ghiglione et al. 2005). The similarity of the SSCP profiles was assessed as described previously (West et al. 2008) by using SAFUM software (Zemb et al. 2007), which normalizes the total area of the SSCP profiles and the mobilities between different runs using the internal standard. The Bray-Curtis distance was then calculated and similarity dendrograms were constructed using the complete linkage method by using Primer-E5 software package (Clarke & Warwick 2001).

Assignment of environmental CE-SSCP peaks. The 16S rRNA genes of several clones from each OTU determined from the clone libraries as described below were amplified directly from E. coli cells in PCR reaction mixes according to a previously described protocol (West et al. 2008). The clone electropherograms were superimposed on the environmental profiles for peak assignment. The areas of the peaks that could be assigned from the clones were obtained after peak detection by GeneScan Analysis.

Sequencing and phylogenetic analysis of 16S rRNA gene clone libraries. To investigate the composition of
the bacterial community, 16S rRNA clone libraries were constructed for the <0.8 µm fraction for one C-mesocosm and one D-mesocosm at 5 m depth at the end of the experiment. The results from the clone libraries enabled CE-SSCP peak assignment and probe design. The clone libraries were constructed according to the methods described previously (West et al. 2008). Plasmid DNA (192 clones for each library) was sequenced using the BigDye™ Terminator kit and the 3730xl Automatic Sequencer by Macrogen using primer 27F (5’-AGA GTT TGA TCM TGG CTC AG-3’). The software BioEdit version 7.0.9.0 (Tom Hall, Ibis Therapeutics) was used to visualize the electropherograms and the sequences were trimmed by PHRED (Ewing et al. 1998). All sequences were checked for chimera formation using CHECK_CHIMERA software (Maidak et al. 2001) and Bellerophon (Hugenholtz & Huber 2003). A total of 33 clones were eliminated (plastids and bad sequences). Phylogenetic groups or OTUs were defined at 99% similarity from the exported CLUSTALW alignment using Cluster software with the single linkage setting (Klepac-Ceraj et al. 2006). The closest relatives were found by submitting a representative sequence of each OTU to BLASTN. Each OTU was named by the closest BLAST hit or by clusters previously identified. One or more sequences from the OTU were fully sequenced as described above using the primer 1492R (5’-TAC GGY TAC CTT GTT ACG ACT T-3’) and the full-length sequences were assembled in BioEdit. The nucleotide sequences data from this study have been deposited in the GenBank database under the accession numbers HM117272 to HM117275 (plastid sequences), HM117276 to HM117458 (C-mesocosm bacterial sequences), HM117439 to HM117462 (cyano-bacterial sequences) and HM117263 to HM117626 (D-mesocosm bacterial sequences).

**Probe design and specificity.** The probes Thiothrix_577a (5’-GCA TCA CAT CCT AGC TAC-3’), Thiothrix_577b (5’-GCA TTA CAT CCT AGC TAC-3’) and NS5_175 (5’-TAT CAG GAT ATG GGG-3’) were designed and their specificity evaluated using the PROBE Design and PROBE Match tools of the ARB software package and the greengenes.arb database (Ludwig et al. 2004). As a cultured representative of the Thiothrix and NS5-1 OTU was not available, the hybridization conditions were optimized by carrying out hybridizations with the 18 samples using different formamide concentrations (from 45% to 60%). To obtain maximum specificity of the probe, the formamide concentration with highest fluorescence signal was selected (55%).

**Abundance of major and specific bacteria using CARD-FISH.** We applied CARD-FISH (Pernthaler et al. 2002) to determine the quantitative importance of major phylogenetic and more specific bacterial groups. CARD-FISH was performed using general and specific rRNA-targeted oligonucleotide probes labelled with horse-radish peroxidase (HRP; ThermoScientific). The probes Eub338-I, -II and -III (Amann et al. 1990, Daims et al. 1999), Alf968 (Amann et al. 1997), Gam42a (Manz et al. 1992) Bet42a and CF319a (Manz et al. 1996) were used to target most of the domain Bacteria and the major bacterial groups. A negative probe (the Eub antisense probe Non338) was used to determine non-specific binding (Glöckner et al. 1999). Unlabeled competitor probes were used for Betaproteobacteria and Gammaproteobacteria (Manz et al. 1992). The more specific probes used in this study were SAR11-152R, SAR11-441R, SAR11-542R, SAR11-732R (SAR11 cluster) (Morris et al. 2002, Sekar et al. 2003). The recently designed probe NorSea72_88 (5’-GCC ACT CGT CAT CAG ATT-3’) and 2 helper probes NorSea72_69H (5’-CCA AGC AAT CCA TGT TAC-3’ and NorSea72_107H (5’-GCC TTA CGC ACC CGT GC-3’) (NorSea72 group belonging to Bacteroidetes) (Laghdass et al. 2010) were also applied in the present study. Two probes were specifically designed to target the group Thiothrix belonging to Gammaproteobacteria (Thiothrix_577a and Thiothrix_577b) and a part of the NS5 cluster (NS5-1) within Bacteroidetes (NS5_175) (see Table 2).

For each sample, 100 ml of raw seawater were filtered onto 0.2 µm PC membranes (47 mm) and incubated overnight at 4°C on pads soaked with 2% paraformaldehyde (PFA). Membranes were rinsed with 0.2 µm filtered Milli-Q water and dried before storing at −20°C until CARD-FISH analysis as described previously (Pernthaler et al. 2002, Sekar et al. 2003). The total number of cells (DAPI cells) and cells affiliated with a specific bacterial group (FITC labelled) were counted using semi-automated microscopy (Olympus Provis AX70) and an image analysis system (ImagePro). Data were collected from 10 counted fields with at least 300 cells per sample. Relative abundances of the bacterial groups are presented either as mean values and standard deviations of triplicate filter pieces (see Table 3).

**RESULTS**

**Response in the abundance and biomass of the microbial community**

The addition of Saharan dust resulted in a rapid increase in phytoplankton biomass at 5 m depth
Laghdass et al.: Saharan dust effects on the microbial community

Table 1. Temporal changes in the chl a concentration (µg l−1), the abundance of Synechococcus spp., pico- and nanoeukaryotes and heterotrophic bacteria at 5 m depth in the control mesocosms (C, n = 3), the dust-amended mesocosms (D, n = 3), and samples collected outside the mesocosms (OUT, n = 1). Samples were collected before dust-addition (t0), after 3 d (t3), 6 d (t6) and 8 d (t8). Mean values ± SD of the 3 C- and 3 D-mesocosms are given. At t0, the samples of the C- and D-mesocosms, and the OUT sample were pooled (n = 7).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>t0</th>
<th>t3</th>
<th>t6</th>
<th>t8</th>
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<tr>
<td></td>
<td>C</td>
<td>D</td>
<td>OUT</td>
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<td>Chl a</td>
<td>0.09 ± 0.02</td>
<td>0.13 ± 0.03</td>
<td>0.20 ± 0.01</td>
<td>0.12 ± 0.00</td>
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<td>Synechococcus spp (×10³ cells ml⁻¹)</td>
<td>14.6 ± 1.3</td>
<td>12.8 ± 1.9</td>
<td>16.8 ± 1.2</td>
<td>14.0 ± 1.0</td>
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<tr>
<td>Pico- and nanoeukaryotes (cells ml⁻¹)</td>
<td>471 ± 120</td>
<td>706 ± 192</td>
<td>805 ± 109</td>
<td>684 ± 152</td>
</tr>
<tr>
<td>Heterotrophic bacteria (×10⁵ cells ml⁻¹)</td>
<td>3.40 ± 0.3</td>
<td>4.75 ± 0.5</td>
<td>5.13 ± 0.3</td>
<td>4.50 ± 0.4</td>
</tr>
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</table>

(Table 1). The initial concentration of chlorophyll a (chl a) was 0.09 µg l⁻¹ and it was about 2-fold higher in the D-mesocosms than in the C-mesocosms and the OUT samples throughout the experiment. The abundance of Synechococcus spp. was higher in the D-mesocosms (17 × 10³ cells ml⁻¹) than in the C-mesocosms and OUT samples (13 to 14 × 10³ cells ml⁻¹), but only at t3 and t6. Pico- and nanoeukaryotes revealed a different pattern, with higher abundances at t0 and t6 in the D-mesocosms (1155 ± 98 cells ml⁻¹, mean ± SD of t0 and t6, n = 6) than in the C-mesocosms (898 ± 121 cells ml⁻¹, mean ± SD of t0 and t6, n = 6) and the OUT samples (766 ± 171 cells ml⁻¹, mean ± SD of t0 and t6, n = 6) (Table 1). Before dust addition, heterotrophic bacterial abundance was 3.40 ± 0.3 × 10⁵ cells ml⁻¹ (mean ± SD, n = 7) and bacterial abundance varied between 4.5 and 5.88 × 10⁵ cells ml⁻¹ during the 8 d of the experiment with no pronounced differences detectable at 5 m depth between the D- and C-mesocosms (Table 1).

**Temporal pattern of the bacterial community structure**

The structure of the total and active bacterial community was examined by 16S rRNA gene and the 16S rRNA transcript CE-SSCP profiles. The Bray-Curtis dendrograms based on the presence or absence of peaks (corresponding to OTUs) and on the relative peak height are presented in Fig. 1. Overall, the CE-SSCP profiles of the 3 replicate C-mesocosms and D-mesocosms revealed good reproducibility throughout the experiment. CE-SSCP profiles of the total bacterial community were clustered according to the time point of sampling and the size fraction, but independent of treatment (Fig. 1A). This pronounced temporal trend was observed not only in all mesocosms, but also in the OUT samples. The dendrogram based on the rRNA transcripts, representative for the active bacterial community, revealed a similar temporal pattern (Fig. 1B). However, 8 d after dust addition, a treatment effect was detectable on the active community of the >0.8 µm size fraction (Fig. 1B).

**Composition of the bacterial community**

To identify the most abundant OTUs of the bacterial communities, 16S rRNA gene clone libraries were constructed at t8 for 1 C- and 1 D-mesocosm. Due to the higher number of OTUs present on the CE-SSCP profiles of the <0.8 µm size fraction (16 ± 1, mean ± SD, n = 6) than of the >0.8 µm size fraction (14 ± 0.5, mean ± SD, n = 5), the clone libraries were constructed for the former size fraction. A total of 351 clone sequences for the 2 clone libraries were analyzed. The cyanobacterial sequences accounted for 0.5 and 2% in the control and dust libraries, respectively, and they were excluded from further analyses. Alphaproteobacteria was the dominant group in both clone libraries (69 and 58% of total bacterial clone sequences in the C- and D-library, respectively) (Table 2). The contribution of Bacteroidetes was substantially smaller and similar in both clone libraries (about 10% of the total bacterial clone sequences). By contrast, the relative contribution of Gammaproteobacteria clones was roughly 2-fold higher in the D- than in the C-library (27 and 14% of total bacterial clone sequences, respectively).

When the sequences were clustered at 99% similarity of the 16S rRNA gene, a few noticeable differences between the 2 clone libraries were detectable. The cluster analysis resulted in the same number of OTUs...
Table 2. Phylogenetic affiliation of each OTU in clone libraries from 1 control (C2) and 1 dust-amended (D2) mesocosm 8 d after dust addition (t8). Each OTU was named by the closest BLAST hit from GenBank or by clusters previously identified. The number of clones indicates the total number of clones determined in both libraries. For the control (C t8) and dust (D t8) libraries, the relative contribution of clones of major bacterial groups is presented as the percent of total clones of the respective treatment (in bold). The relative contribution of the OTUs is presented as the percent of clones of the respective major bacterial group. The phylogenetic clusters in grey represent clones with complete sequences, and the number of complete sequences is indicated in parentheses. Unc: uncultured.

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<th>Identity (%)</th>
<th>Percent contribution</th>
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and singletons (26) for both libraries (Table 2). A higher number of OTUs was determined for Alphaproteobacteria (16 OTUs), whereas only 10 and 8 OTUs were determined for Gammaproteobacteria and Bacteroidetes, respectively (Fig. 2). Within Alphaproteobacteria, the 2 OTUs SAR11-IA and SAR11-IB (Kan et al. 2008) were the most dominant OTUs in both libraries (44 ± 3 and 21 ± 0.2% of Alphaproteobacteria clones, respectively, mean ± SD, n = 2). Gammaproteobacteria were dominated by the 3 OTUs Alteromonas macleodii, JTB148 and SAR86-1 in the C-library, each accounting for 20% of Gammaproteobacteria clones. By contrast, in the D-library, only the OTU A. macleodii had an important relative contribution (23% of Gammaproteobacteria clones). In the D-mesocosm, the number of total clones of A. macleodii (10) was 2-fold higher than in the C-mesocosm (5). It was further interesting to note that the OTU Thiothrix was detected only in the D-library, where it represented 11% of Gammaproteobacteria clones. Within the Bacteroidetes, the OTU NS4-1 was specific to the D-library (14% of Bacteroidetes clones) and the OTU NS5-1 had the highest relative contribution to Bacteroidetes clones in this library (28% of Bacteroidetes clones). In contrast, in the C-library the 4 OTUs NS5-2, Aegean179, NS7 and Winogradskyella contributed equally to Bacteroidetes clones (each accounting for roughly 10% of Bacteroidetes clone sequences). We observed no differences in the contribution of the 3 OTUs determined within the Actinobacteria between the 2 clone libraries.

A f-LIBSHUFF analysis (Schloss et al. 2004) allowed comparison of the 2 clone libraries. When the homologous coverage curve of one library (X) was compared with the heterologous curve XY, neither the comparison $(C_X - C_{XY})^2$ nor the reverse comparison $(C_Y - C_{XY})^2$ showed a significant difference between the 2 libraries, with values of $p = 0.0631$ and $p = 0.1928$ for $(C_X - C_{XY})^2$ and $(C_Y - C_{XY})^2$, respectively.

Identification of specific CE-SSCP peaks

To identify the CE-SSCP peaks that were different between the C- and the D-mesocosms at $t_8$ we performed CE-SSCP analysis of several clones from each OTU identified, and
aligned the migration of the clones to the environmental community profile (Fig. 2). One peak that revealed noticeable differences between the C- and the D-mesocosms was identified as *Alteromonas macleodii* at the end of the experiment (Fig. 2A,B). For the 16S rRNA transcript-based profile of the >0.8 µm size fraction, the relative contribution of this peak to total peak area was about 2-fold higher in the D-mesocosms than that in the C-mesocosms (14 ± 8 and 7 ± 1% of the total peak area, respectively, n = 3). For the 16S rRNA gene-based profile, the relative contribution of this peak to total peak area was, respectively, 2 ± 1 and 4 ± 2% for the C- and D-mesocosms for the 16S rRNA gene-based profile, and, respectively, 9 ± 1 and 6 ± 6% for the C- and D-mesocosms for the 16S rRNA transcript-based profile. Other peaks identified were SAR86, SAR11 and *Synechococcus*, but no differences were detectable for these OTUs.

**Contribution of bacterial groups to total abundance**

We used previously designed and 2 specifically-designed probes for the OTUs Thiothrix and NS5-1 (See ‘Materials and methods’) to quantify the relative abundance of the major and some more specific bacte-
Overall, no differences in the abundance of the major bacterial groups were detectable between treatments, and we observed some minor differences only at the OTU level. For the description of the relative abundance of most of the bacterial groups determined in the present study, we have therefore pooled the results from the different treatments and time points (n = 14). The proportion of cells that hybridized with the general EUB probes and the NON338 probe was on average 65 ± 13% (n = 14) and 3 ± 2% (n = 14) respectively. SAR11 had important contributions to bulk abundance in the C- and D-mesocosms and in the OUT samples throughout the experiment (35 ± 7% of DAPI-stained cells respectively, n = 14, results from all treatments and time points pooled), while the contribution of Roseobacter remained low (8 ± 3% of DAPI-stained cells, n = 14) (Table 3). The relative contributions of Gammaproteobacteria, SAR86 and Thiothrix were independent of treatment and did not vary over time (22 ± 4, 8 ± 2, 1 ± 1% of DAPI-stained cells, respectively, n = 14). Even though relative abundances were overall low (2 to 5% of DAPI-stained cells), Alteromonas revealed slightly higher abundances in the D-than in the C-mesocosms 8 d after dust addition (5 and 3% of DAPI-stained cells, respectively, n = 3 in both cases). Bacteroidetes displayed similar relative abundances in the D-mesocosms and the OUT samples throughout the experiment (31 ± 6% of DAPI-stained cells, n = 11, data from all D-mesocosms and OUT samples pooled), but the relative abundance of Bacteroidetes was substantially lower in the C-mesocosms at t8 (17 ± 4% of DAPI-stained cells, n = 3) (Table 3). While the OTU NorSea72 did not display any differences between treatments (5 ± 1% of DAPI-stained cells, n = 14), the OTU NS5-1 was slightly more abundant in the D-than in the C-mesocosms at t8 (2 ± 0.2% and 0.7 ± 0.1% of DAPI-stained cells, respectively, n = 3 in both cases). Combining the results of all bacterial groups considered here, no significant differences between the C- and the D-mesocosms were detectable after 8 d (Kruskal-Wallis, p > 0.8).
gated using trace-metal-clean in situ mesocosms. The 2 previous studies that addressed this question were carried out in microcosm experiments (up to 15 l) (Reche et al. 2009, Lekunberri et al. 2010). The lack of in situ observations is due to the episodic nature of atmospheric dust deposition that makes it difficult to follow related in situ changes at appropriate temporal and spatial scales. For the biogeochemical parameters determined during the DUNE project and the bacterial community structure, the control mesocosms represented the temporal changes that occurred in the Bay of Elbo well (Guieu et al. 2010), and the 3 replicate mesocosms exhibited high reproducibility in this study. Large in situ mesocosms represent therefore an original and appropriate tool to investigate an atmospheric dust event in a LNLC environment in close to in situ conditions.

During the DUNE project, dust deposition resulted in a rapid, but transient, increase in dissolved inorganic phosphate concentrations by 12 nM at the surface (Pulido-Villena et al. 2010). From our observations, dust deposition resulted in an approximately 2-fold increase in the concentration of chl a in the upper 5 m over the duration of the experiment (Table 1). In contrast, bacterial heterotrophic abundance did not reveal any marked increase at 5 m depth following dust addition (Table 1). The P-fertilizing effect of dust on the autotrophic community confirms previous microcosm experiments in the Mediterranean Sea (Bonnet et al. 2005, Herut et al. 2005, Lekunberri et al. 2010) and it has also been demonstrated for heterotrophic bacterial metabolism (Herut et al. 2005, Pulido-Villena et al. 2008, Reche et al. 2009, Lekunberri et al. 2010). Only in one study was this bulk response accompanied by noticeable changes in the bacterial community structure (Lekunberri et al. 2010). In this study, the addition of dust or inorganic phosphorus (P) to coastal Mediterranean waters resulted in a similar bacterial community structure in both treatments. This observation led to the conclusion that P introduced by dust addition was the factor driving observed changes in bacterial community structure (Lekunberri et al. 2010). Changes in bacterial community structure following P addition to Mediterranean waters are also reported from previous experimental studies (Lebaron et al. 2001, Schäfer et al. 2001, Pinhassi et al. 2006). In these previous studies, the final P concentrations were, however, substantially higher (0.3 to 0.6 µM) than in the DUNE mesocosms amended with dust (≤17 nM) (Pulido-Villena et al. 2010). If, in the present study, the input of the low P concentration due to dust addition had a structuring effect on the bacterial community composition, it was probably hidden in the natural temporal trend (Fig. 1).

Over the 8 d of the experiment, we observed a pronounced temporal evolution of the total (16S rRNA genes) and active (16S rRNA transcripts) bacterial community structure (Fig. 1). Changes in bacterial community structure are frequently observed on a seasonal time scale (Ghiglione et al. 2005, Fuhrman et al. 2006, Mary et al. 2006, Lami et al. 2009), but they can also occur on shorter temporal scales (Hewson et al. 2006, Lamy et al. 2009). Changes in bacterial community structure on a weekly time scale could be of importance during different phases of phytoplankton blooms, phases of transition between seasons or following environmental perturbations (Fandino et al. 2001, West et al. 2008). Roughly 2 wk prior to the DUNE experiment, a Saharan dust event (K. Desboeuf & F. Dulac pers. comm.) and heavy rainfalls (Wagener et al. 2010) occurred at the study site, which could explain the temporal pattern in bacterial community structure we observed during our study.

Despite this natural temporal trend, dust deposition had an impact, detectable at the OTU level, on a few particular features of the bacterial community composition. The OTUs Thiobrix and Alteromonas macleodii, belonging to Gammaproteobacteria, and the Bacteroidetes OTU NSS-1 were more abundant in or specific to the dust-amended mesocosms, compared to the control mesocosms at the end of the experiment (Table 2; Fig.2). The presence of these OTUs has so far not been linked to dust deposition. The OTU Thiobrix has been observed in clone libraries of the North Pacific coast (Brown et al. 2005) and it was present in many samples taken during the Global Ocean Sampling (Shaw et al. 2008). The cluster NS5 appears to be common in the coastal environment of temperate (DeLong et al. 1993, Alonso et al. 2007) and high latitude regions (Bano & Hollibaugh 2002). A noticeable pattern observed in our study was the relatively high contribution of the species A. macleodii to the active bacterial community of the >0.8 µm fraction in the D-mesocosms (Fig. 2). A. macleodii and related species are among the most widespread cultured Gammaproteobacteria. Previous observations that A. macleodii rapidly responds in nutrient enrichment experiments (Pukall et al. 1999, Schäfer et al. 2000) and that its presence in situ is often found in association with particulate matter (Acinas et al. 1999, López-López et al. 2005) have led to the idea that A. macleodii has an opportunistic way of life, which favours its dominance in response to enhanced nutrient supply. The analysis of the complete genome of a surface ecotype of A. macleodii indicates that it is well adapted to rapidly changing environmental conditions, by means of complex patterns of regulation and environmental sensing, features that could be linked to transient particle attachment (Ivars-Martinez et al. 2008).

Beside the clear impact on P cycling mentioned above, it is also expected that dust addition alters iron cycling. During the DUNE experiment, dissolved iron
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LITERATURE CITED


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