



Biogeographic variations of picophytoplankton in three contrasting seas: the Bay of Bengal, South China Sea and Western Pacific Ocean

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ABSTRACT: Marine picophytoplankton are abundant in many oligotrophic oceans, but the known geographical patterns of picophytoplankton are primarily based on small-scale cruises or timeseries observations. Here, we conducted a wider survey (5 cruises) in the Bay of Bengal (BOB), South China Sea (SCS) and Western Pacific Ocean (WPO) to better understand the biogeographic variations of picophytoplankton. Prochlorococcus (Pro) were the most abundant picophytoplankton (averaging $[1.9-3.6] \times 10^4$ cells ml⁻¹) across the 3 seas, while average abundances of Synechococcus (Syn) and picoeukaryotes (PEuks) were generally 1-2 orders of magnitude lower than Pro. Average abundances of total picophytoplankton were similar between the BOB and SCS $(4.7 \times 10^4 \text{ cells ml}^{-1})$, but were close to 2-fold less abundant in the WPO $(2.5 \times 10^4 \text{ cells ml}^{-1})$. Pro and Syn accounted for a substantial fraction of total picophytoplankton biomass (70-83%) in the 3 contrasting seas, indicating the ecological importance of Pro and Syn as primary producers. Pro were generally abundant in oligotrophic open waters; however, the exceptional presence of Pro near the SCS coast was potentially associated with the Kuroshio intrusion. Syn and PEuk abundances were higher near freshwater-dominated areas, which was likely due to dilution waters. Water temperature and cold eddies were also major drivers responsible for the biogeographic distributions of picophytoplankton. Although Pro, Syn and PEuks showed negative correlations with nutrient concentrations, their maximal abundances in vertical distribution showed positive correlations with the nutricline depth, indicating that nutrient availability plays a 2-faceted role in regulating the biogeographic variation in picophytoplankton.

KEY WORDS: Picophytoplankton \cdot Abundance \cdot Biogeographic variation \cdot Bay of Bengal \cdot South China Sea \cdot Western Pacific Ocean

1. INTRODUCTION

Marine picophytoplankton refer to a genetically diverse group of photosynthetic organisms, which include picocyanobacteria of the genera *Prochlorococcus* (Pro) and *Synechococcus* (Syn), as well as a diverse assemblage of picoeukaryotes (PEuks). Pro and Syn are both ecologically dominant and widespread members in warm oligotrophic waters (Liu et al. 2007, Seymour et al. 2010), although Pro are gen-

erally more abundant than Syn and PEuks, often by 10-fold or more in the open ocean (Jiao & Yang 2002, Flombaum et al. 2013). Furthermore, Syn and PEuks have a wide geographical distribution that covers both oligo- and mesotrophic oceanic and coastal areas from pole to pole (Olson et al. 1990, Flombaum et al. 2013, Grébert et al. 2018). Observations in the oligotrophic Pacific Ocean and Atlantic Ocean have revealed that picophytoplankton can account for approximately 60–80 % of the total primary produc-

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tivity (Campbell et al. 1997, Grob et al. 2011). In eutrophic coastal waters, picophytoplankton also play a significant role with their high abundance and carbon biomass, contributing nearly 42-55% of the total primary productivity (Gaulke et al. 2010), and are therefore thought to contribute roughly 1/3 to global net primary production (Quere et al. 2005). Given their ubiquitous distribution and high abundance, as well as their significant contribution to global net primary production, picophytoplankton have been considered to have substantial impacts on oceanic ecosystems and global biogeochemical cycles (Flombaum et al. 2013). Presenting the biogeographic variations of picophytoplankton in different ecosystems is thus central to our understanding of where and how these particular phytoplankton contribute to the biogeochemical cycles.

A large data set has been accumulated on picophytoplankton populations in the world oceans (e.g. Campbell et al. 1997, Zhang et al. 2013, Chen et al. 2015, Wei et al. 2019b), but a caveat to their investigation is that the abundances and distributions of different picophytoplankton populations were measured from just 1 oceanic or coastal ecosystem, i.e. previous estimates could not address large-scale spatial variations in picophytoplankton populations. Accordingly, our limited understanding of largescale biogeographic variations in picophytoplankton constrains our ability to incorporate them into marine biogeochemical models (Flombaum et al. 2013). As a part of the warm pool covering the tropical eastern Indian Ocean (EIO) and West Pacific Ocean (WPO) (Sengupta & Ravichandran 2001), the Bay of Bengal (BOB) has received far less attention in research on picophytoplankton than the South China Sea (SCS) and WPO. The SCS, as a crossroads of the EIO and WPO, is profoundly influenced by neighboring physical processes, such as circulations and water masses.

However, the 3 seas still have contrasting differences in structural and functional components (Qu et al. 2005, Cheung et al. 2008, Wei et al. 2019b). Importantly, the variability in picophytoplankton abundance and distribution is intimately related to these physical processes, as well as environmental factors (e.g. temperature, salinity, irradiance and nutrient availability) (Landry et al. 2003, Lee et al. 2014, Grébert et al. 2018). Nevertheless, to date, there is a lack of comprehensive knowledge on the abundances and distributions of picophytoplankton throughout the BOB, SCS and WPO. In that context, the BOB, SCS and WPO appear to be ideal fields for ecological surveys on biogeographic variations in picophytoplankton.

Here, we report on abundances and distributions of 3 picophytoplankton groups using flow cytometric (FCM) analyses in 3 contrasting seas: BOB, SCS and WPO. The purpose of this paper is to develop a more comprehensive picture of large-scale variability of picophytoplankton in these 3 complicated ecosystems. The abundance datasets were compiled to analyze the associated environmental variables, and the possible mechanisms shaping picophytoplankton variations in the 3 contrasting seas are also discussed.

2. MATERIALS AND METHODS

2.1. Field surveys

Sampling occurred on 5 cruises of opportunity aboard the RVs 'Shiyan I,' 'Dongfanghong 2' and 'Kexue 3' in the BOB, SCS and WPO (Fig. 1). We joined 3 SCS cruises including the northeastern SCS (referred to as NSCS; 7 July to 10 August 2017), the central SCS (CSCS; 12 May to 13 June 2017) and the western SCS (WSCS; 3 September to 13 October

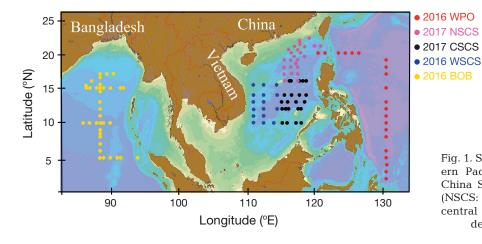


Fig. 1. Stations sampled in 2016 (WPO: Western Pacific Ocean, WSCS: western South China Sea, BOB: Bay of Bengal) and 2017 (NSCS: northern South China Sea, CSCS: central South China Sea). Different colors denote different sampling cruises

2016). These 3 cruises sampled 80 stations covering the entire SCS and its continental shelf area. The WPO cruise (3–20°N, 125–130°E) was conducted aboard the RV 'Kexue 3' from 27 September to 25 October 2016, and 21 stations were investigated. Another cruise of opportunity aboard the RV 'Dongfanghong 2' (8 October to 20 December 2016) sampled 36 stations in the BOB (5–20°N, 85–95°E), within the eastern Indian Ocean. A variety of oceanographic conditions and habitats were sampled between 3° and 23°N latitude.

2.2. Sampling and analysis

Seawater samples and measurements of temperature and salinity were performed using a SeaBird conductivity/temperature/depth (CTD) meter (SBE 19 Plus) equipped with 10 or 12 l Niskin bottles. FCM samples (2 ml) for picophytoplankton were generally collected from 7 depths at most stations, including surface (5 m), 25, 50, 75, 100, 150 and 200 m. At some stations in the SCS, however, picophytoplankton samples were collected from 10 depths (5, 10, 20, 25, 50, 65, 75, 100, 150, 200 m). Seawater samples for picophytoplankton were fixed and preserved on board following the standard methods detailed by Jiao & Yang (2002) and Jiao et al. (2005). A total of 2658 samples were collected in the BOB, SCS and WPO. Ancillary nutrient samples (300 ml) were filtered through 0.45 µm cellulose acetate membrane filters, and then quickly frozen at -20°C and analyzed as soon as possible. Nutrients, mainly including dissolved inorganic nitrogen (DIN; the sum of the concentrations of nitrate, nitrite and ammonia), dissolved inorganic phosphorus (DIP) and dissolved inorganic silicate (DSi), were determined using a Technicon AA3 Auto-Analyzer (Bran+Luebbe) according to the method reported by Brzezinski & Nelson (1986) and Karl & Tien (1992). In addition, we imposed a minimum nutrient concentration of 0.01 µmol l⁻¹ to avoid issues with detection limits.

2.3. Flow cytometry analysis

A total volume of only 198 μ l (flow rate at 66 μ l min⁻¹ running for 3 min) was analyzed using an FCM (BD Accuri C6) equipped with 488 and 640 nm laser beams. Approximately 10 000 – 40 000 events were detected on the FCM. Pro were generally the smallest picophytoplankton cells (~0.6 μ m) with low chlorophyll a (chl a) concentrations (Campbell et al.

1997, Liu et al. 2007), showing relatively low red fluorescence signals (FL3, >670 nm). Large PEuks (1-2 μm) showed the highest red fluorescence because they were mostly comprised of larger and richer chl a cells (Lee et al. 2014). Pro were discriminated from PEuks based on their lower FL3 and side scatter (SSC) signals, whereas Syn were identified by the orange fluorescence signals (FL2, 585 ± 42 nm) because of their characteristic contents of phycoerythrin (Bertilsson et al. 2003, Jiao et al. 2005). Within the present study, Syn, Pro and PEuks were differentiated by their fluorescence signals and scatter properties in the scatterplots of FL2 vs. FL3 and SSC vs. FL3. Yellow-green fluorescent beads (2 µm; Polysciences) were added as an internal standard (Calvo-Díaz & Morán 2006).

A recently published study revealed that the BD Accuri FCM is not an ideal flow cytometer to detect the picophytoplankton populations, and suggested that Pro in the well-lit layers of the water column may be underestimated with the BD Accuri FCM because of the low contents of photosynthetic pigments in cells and only partial resolution from the background noise (Ribeiro et al. 2016). To avoid loss of dispersion and resolution of target cells in the scatterplot due to the overlapping signals with noise, the trigger threshold (600) was set well below the lowest scatter signals from surface Pro cells. Scatter-to-fluorescence crosstalk and spurious scatter events were minimized by setting the polarization of the excitation laser perpendicular to the axis of flow. After these settings, 2 ml of DI (deionized) water were preferentially run at a steady flow rate of 66 µl min⁻¹ to collect and gate the scatter and fluorescence signals of noise. If higher background counts were recorded, the FCM was thoroughly cleaned with 5% bleach, and rinsed until the background counts fell below 8 events μl^{-1} . The fluorescence and scatter signals were captured with user-built detector assemblies with an extended range by combining the signals from 2 photomultipliers that operate at different gains. The relative gain settings were calibrated by a regression analysis of the events that fall within the linear window of both detectors (van den Engh et al. 2017). This approach allowed simultaneous detection of all picophytoplankton populations ranging from dim surfacedwelling Pro to bright PEuks.

The carbon biomass of Pro, Syn and PEuks in the 3 contrasting seas were calculated based on abundance-to-carbon conversion factors from the literature (Table 1). The conversion factor of PEuks previously used in the WPO was 1500–2100 fg C cell⁻¹ (Campbell et al. 1997). In the present study, however,

Table 1. Carbon conversion factors (fg C cell⁻¹) for 3 picophytoplankton populations in 3 contrasting seas. Syn: *Syne-chococcus*; Pro: *Prochlorococcus*; PEuks: picoeukaryotes

Study area	Pro	Syn	PEuks	s Reference
				Campbell et al. (1997)
South China Sea	43	112	293	Liu et al. (2007)
Bay of Bengal	32	129	160	Wei et al. (2019a)

an average conversion factor of 964 fg C cell⁻¹ for PEuks was derived from our estimated PEuks cell sizes according to the empirical relationship (Menden-Deuer & Lessard 2000): $\log C = 0.94 \times \log V - 0.60$. Thus, this conversion factor may provide a lower estimation of carbon biomass for PEuks than previous biomass values in the WPO.

2.4. Statistical analysis

All mean data are given \pm SD. To assess the relative significance of environmental variables in influencing picophytoplankton abundance across the 3 contrasting seas, canonical correspondence analysis (CCA) and Pearson's correlation coefficients were used to explore the relationship between the abun-

dances of 3 picophytoplankton groups and environmental factors. The CCA was performed using Canoco 4.5 software, and Pearson's correlation analysis was achieved using SPSS 25.0 (IBM). Furthermore, graphs were generated by Origin Pro 8.5 and Ocean Data View 4.0. In all tests, statistical significance was accepted at p < 0.05. Picophytoplankton abundance was log₁₀-transformed to improve the normality for statistical analysis. Unless otherwise stated, picophytoplankton abundance and biomass used for presenting the spatial distribution are expressed as the depth-weighted averages (calculated by dividing the trapezoidal integration of measured values for each variable by the maximum sampling depth) because Pro and PEuks abundances were relatively low in the surface layer (see Section 3). These depth-weighted averages, which involve a series of abundance variations with depth, were better suited for our distributional patterns than discrete sample data, and were calculated as per Crosbie & Furnas (2001):

$$A = \left[\sum_{n=1}^{n+1} \frac{(A_i + A_{i+1})}{2} \times (D_{i+1} - D_i)\right] / (D_{MSL} - D_S)$$
 (1)

where A_i is the picophytoplankton abundance (cells $\mathrm{ml^{-1}}$) or biomass ($\mu \mathrm{g \ C \ l^{-1}}$) in sampling layer i; n is the number of sampling layers, and D_i is the depth of sampling layer i (m); D_{MSL} and D_{S} are the depth of the maximum sampling layer (m) and the surface sampling depth (5 m), respectively.

3. RESULTS

3.1. Hydrography and nutrients

A surface warm tongue (>30°C) with salinity >34 was observed from 10 to 20° N in the WPO (Fig. 2), thereby indicating an apparent influence of the Kuroshio. The high surface temperature and salinity were analogously found in the NSCS, attributable to the fact that the Kuroshio intrusion brought higher salinity and warmer seawater, profoundly changing its water layer structure. By contrast, the SCS and BOB were characterized by low temperature and salinity (Table 2). In the SCS, surface temperature

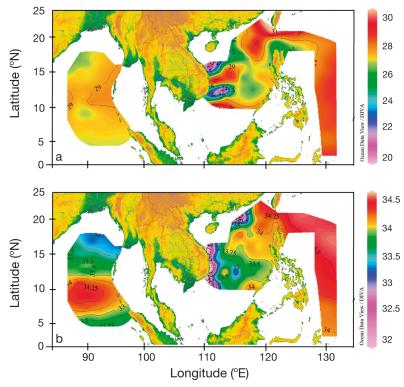


Fig. 2. Spatial variation in surface (a) temperature (°C) and (b) salinity in 3 contrasting seas

Table 2. Mean \pm SD of temperature (°C), salinity and nutrients (μ mol l⁻¹) across 3 contrasting seas. DIN: dissolved inorganic nitrogen; DIP: dissolved inorganic phosphorus; DSi: dissolved inorganic silicate

Study area	Гетреratur	e Salinity	DIN	DIP	DSi
West Pacific Ocean	25.9 ± 4.5	34.6 ± 0.4	1.85 ± 3.32	0.17 ± 0.05	1.91 ± 0.27
South China Sea	23.1 ± 5.2	34.0 ± 2.3	4.61 ± 1.42	0.38 ± 0.07	4.23 ± 1.15
Bay of Bengal	23.0 ± 5.3	34.3 ± 0.6	6.95 ± 2.65	0.45 ± 0.09	5.57 ± 1.89

and salinity were usually lower in the coastal regions due to freshwater influence, whereas sea surface salinity at some coastal stations was >34, indicating the effect of coastal upwelling (Fig. 2). Under the influence of coastal runoff, surface temperature in the northern part of the BOB was below 29°C, with a comparatively low surface salinity of approximately 32–33. However, the surface colder seawater (~27°C) with higher salinity (>34) was particularly observed around 10° N in the BOB. Analysis of the satellite altimetry revealed that a negative sea level anomaly was present at this site at the time of the 2016 BOB cruise (AVISO, http://las.aviso.altimetry.fr/). Collectively, the sea level anomaly and the hydrographic conditions at this site were consistent with the presence of a cyclonic eddy (i.e. cold eddy).

Contrasting vertical patterns of nutrient concentrations (DIN, DIP and DSi) from the surface to a maximum of 200 m were observed in the pooled dataset across the 3 seas (Fig. 3). The average values of nutrient concentrations were highest in the BOB followed by the SCS and WPO (Table 2). Analysis of the nutrient profiles revealed that high nutrient concentrations markedly shoaled in the BOB compared to the other 2 seas, and the nutricline depth was shallow (<50 m). In the SCS, nutrient concentrations were relatively low in the upper 50–75 m, but increased

rapidly from 75 to 200 m, indicating that the depth of the nutricline typically ranged between 50 and 75 m. Due to the pronounced stratification in the WPO, nutrients were generally depleted within the upper $100 \, \text{m}$, with a deep nutricline at ~ $100 \, \text{m}$. As expected, there were contrasting differences in physical and chemical backgrounds among the 3 seas.

3.2. Picophytoplankton abundance

Pro were the most abundant picophytoplankton across the 3 contrasting seas (Table 3). The maximum Pro abundance $(7.7 \times 10^5 \text{ cells ml}^{-1})$ was observed in the SCS, and average Pro abundance was also highest in the SCS $(3.6 \times 10^4 \text{ cells ml}^{-1})$, followed by the BOB $(3.3 \times 10^4 \text{ cells ml}^{-1})$ and WPO $(1.9 \times 10^4 \text{ cells ml}^{-1})$. Average abundances of Syn and PEuks were generally 1–2 orders of magnitude lower than Pro,

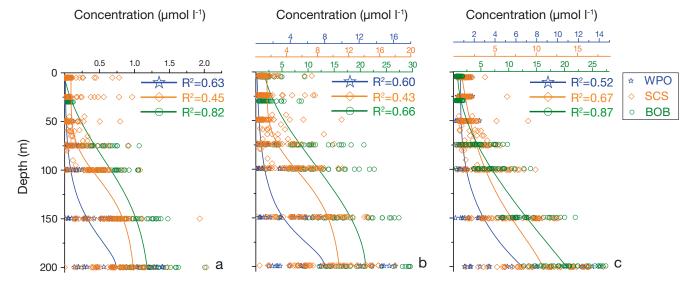


Fig. 3. Vertical profiles for concentrations (μ mol l⁻¹) of (a) dissolved inorganic phosphorus (DIP), (b) dissolved inorganic nitrogen (DIN) and (c) dissolved inorganic silicate (DSi). Symbols and colors represent 3 contrasting seas. Different colored solid lines indicate the curve-fitting trends for all nutrients data (n = 2080) versus depth. R^2 denotes the fitting variance of the nonlinear regression model. Note the x-axis scale in panels b and c; different colored x-axes correspond to 3 contrasting seas (WPO: Western Pacific Ocean, SCS: South China Sea, BOB: Bay of Bengal)

	Cell type	West Pacific Ocean	South China Sea	Bay of Bengal
Pro	Range	$0.4 \times 10^3 - 1.1 \times 10^5$	$0.4 \times 10^3 - 7.7 \times 10^5$	$0.7 \times 10^3 - 2.0 \times 10^5$
	Mean	$(1.9 \pm 0.7) \times 10^4$	$(3.6 \pm 0.4) \times 10^4$	$(3.3 \pm 0.5) \times 10^4$
Syn	Range	$0.1 \times 10^3 - 2.9 \times 10^4$	$0.1 \times 10^3 - 1.5 \times 10^5$	$0.2 \times 10^2 - 1.9 \times 10^5$
-	Mean	$(4.2 \pm 0.9) \times 10^3$	$(8.1 \pm 1.1) \times 10^3$	$(1.1 \pm 0.3) \times 10^4$
PEuks	Range	$ND - 1.2 \times 10^4$	$ND - 9.2 \times 10^4$	$ND - 4.5 \times 10^4$
	Mean	$(9.3 \pm 1.7) \times 10^2$	$(2.5 \pm 0.2) \times 10^3$	$(3.1 \pm 0.1) \times 10^3$
Total	Range	$0.7 \times 10^3 - 1.2 \times 10^5$	$0.5 \times 10^3 - 8.2 \times 10^5$	$1.1 \times 10^3 - 4.2 \times 10^5$
	Mean	$(2.5 \pm 0.4) \times 10^4$	$(4.7 \pm 0.5) \times 10^4$	$(4.7 \pm 0.8) \times 10^4$

Table 3. Range and mean \pm SD of abundance (cells ml⁻¹) for 3 groups of picophytoplankton (n = 2658) across 3 contrasting seas. ND: not detected; Syn: $Synechococcus_i$; Pro: $Prochlorococcus_i$; PEuks: picoeukaryotes

while average Syn abundance was found as high as 1.1×10^4 cells ml⁻¹ in the BOB. Moreover, the maximum Syn abundance (1.9×10^5 cells ml⁻¹) was also recorded in the BOB. Overall, the average abundances of the 3 picophytoplankton groups were somewhat similar between the BOB and SCS (Pearson's correlation, r = 0.98, p < 0.05), but were 2- to 3-fold less abundant in the WPO.

3.3. Spatial distribution

The depth-weighted average Pro abundance (referred to as 'Pro abundance') was usually high in oligotrophic open waters (Fig. 4). The maximal Pro abundance was found in the CSCS. Apart from the

CSCS, Pro were exceptionally abundant along the NSCS coast. In the BOB, Pro abundance was relatively high in the northern part and around 10°N, where the distributional pattern appeared to closely follow the spatial variation in surface currents or water masses. Pro abundance in the WPO, however, was much less abundant than in the SCS and BOB. The depth-weighted average Syn and PEuks abundances (referred to as 'Syn and PEuks abundances') were geographically uneven and varied significantly across the 3 contrasting seas. Syn and PEuks abundances were relatively high in the BOB and SCS, but were low in the WPO. In contrast to Pro, PEuks abundance was broadly higher near the SCS coast, whereas Syn abundance was higher in both coastal and open waters.

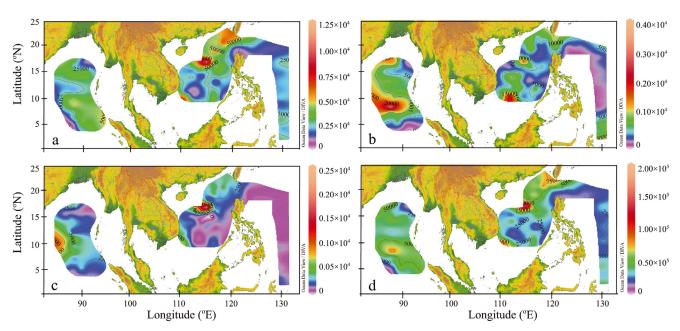


Fig. 4. Spatial distributions for depth-weighted average abundances (cells ml⁻¹) of (a) *Prochlorococcus* (Pro), (b) *Synechococcus* (Syn), (c) picoeukaryotes (PEuks) and (d) total picophytoplankton in 3 contrasting seas. Note the different logarithmic scales on the color bars, reflecting the variability in magnitude of respective picophytoplankton abundances

All data points of picophytoplankton abundance against depth were analyzed to plot the fitting curves in Fig. 5. There were some similarities in the vertical patterns of the 3 picophytoplankton groups among the BOB, SCS and WPO. Syn were most abundant in surface layer, and only very few cells were counted below 100 m. In the BOB, Syn abundance occurred evenly from the surface to 50 m, and then decreased drastically with depth. In the SCS and WPO, Syn abundance was vertically homogeneous from the surface to depths of about 50-75 and 100 m, respectively. The highest Pro abundance in the BOB was found in the subsurface around 50 m, while in the SCS and WPO, Pro had strong deep maxima at 50-75 and 100 m. PEuks and Pro had similar vertical patterns across the 3 seas, and the peaks of PEuks abundance in the BOB, SCS and WPO were evident at 50, 50-75 and 75 m, respectively. The depths of maximal abundances of Pro, Syn and PEuks were significantly correlated with the nutricline depths in the 3 contrasting seas (Fig. 3; Pearson's correlation, r = 0.87, p < 0.01), indicating that the growth of 3 picophytoplankton populations in vertical distribution were possibly controlled by the nutricline.

3.4. Carbon biomass

The spatial distributions of depth-weighted average carbon biomass (referred to as 'C biomass') for the 3 picophytoplankton populations were similar to

those of abundances (Figs. 4 & 6; Pearson's correlation, r=0.71, p<0.01). This finding indicated that the C biomass of picophytoplankton appeared to be mainly determined by the abundance rather than the conversion factor (Table 1). The C biomass of total picophytoplankton showed the highest value in the SCS (14.8 μ g C l⁻¹) and the lowest in the BOB (0.9 μ g C l⁻¹).

Pro and Syn accounted for a substantial fraction of total picophytoplankton biomass (approximately 70–83%) across the 3 seas (Table 4). In contrast, PEuks were not a large component of the total picophytoplankton biomass. The relative biomass of PEuks was low in the BOB (17%), yet markedly increased by 7 and 13% in the SCS and WPO, respectively. The spatial variation of average C biomass for PEuks in the 3 contrasting seas was opposite to that of average abundance (Table 3). This opposite variation may be attributed to the different cellular sizes in PEuks (Table 1). The average C biomass of total picophytoplankton was highest in the SCS (3.0 μ g C l⁻¹) followed by the WPO (2.9 μ g C l⁻¹) and BOB (2.6 μ g C l⁻¹).

3.5. Associated environmental variables governing picophytoplankton biogeography

CCA and Pearson's correlation analysis were used to identify which key environmental variables explained the biogeographic variations of picophytoplankton in the 3 contrasting seas (Fig. 7, Table 5). CCA is a multivariate statistical analysis of weighted

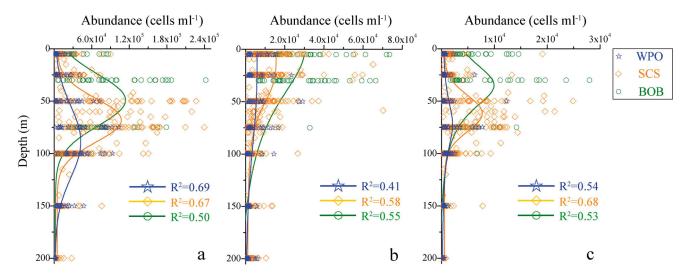


Fig. 5. Vertical profiles for abundances (cells ml^{-1}) of (a) *Prochlorococcus* (Pro), (b) *Synechococcus* (Syn) and (c) picoeukaryotes (PEuks). Symbols and colors represent 3 contrasting seas (WPO: Western Pacific Ocean, SCS: South China Sea, BOB: Bay of Bengal). Different colored solid lines indicate the curve-fitting trends for all abundance data (n = 2658) versus depth. R^2 denotes the fitting variance of the nonlinear regression model. Note the different logarithmic scales on the x-axes, reflecting variability in the magnitude of the respective picophytoplankton abundances

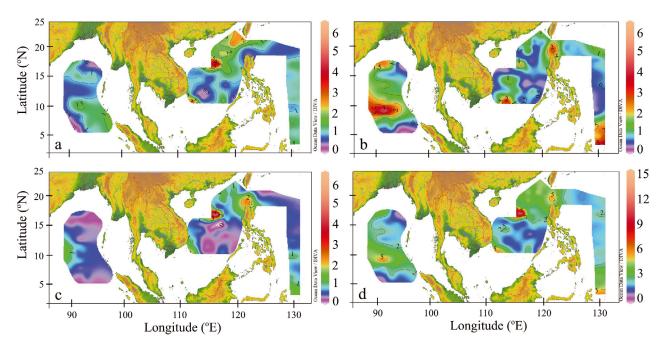


Fig. 6. Spatial distributions of depth-weighted average carbon biomass (μ g C l⁻¹) of (a) *Prochlorococcus* (Pro), (b) *Synechococcus* (Syn), (c) picoeukaryotes (PEuks) and (d) total picophytoplankton in 3 contrasting seas. Note the different scale on the color bar in panel (d)

averaging ordination, particularly relating the biological assemblages of species to known environmental factors (ter Braak & Verdonschot 1995). Moreover, CCA is an efficient indirect gradient analysis when species have regression response curves with respect to environmental gradients (Hill 1991). Questions in the relationship between species and environmental variables can be answered more directly by the CCA ordination. Within our study, the species—environment correlations of CCA axes 1 and 2 were 0.632 and 0.873, respectively. The correlation coefficients among environmental axes were 0, while the species axes were approximately vertical to each other. The p-value for the significance test of all

Table 4. Mean \pm SD carbon biomass (µg C l^{-1}) of *Prochlorococcus* (Pro), *Synechococcus* (Syn) and picoeukaryotes (PEuks) along with respective relative biomass (%; fraction of total picophytoplankton biomass) in 3 contrasting seas

Cell ty	pe	West Pacific Ocean	South China Sea	Bay of Bengal
Pro	Mean Percentage	1.13 ± 0.22 38	1.48 ± 0.79 49	1.05±0.34 40
Syn	Mean Percentage	0.93 ± 0.39 32	0.81 ± 0.35 27	1.14±0.56 43
PEuks	Mean Percentage	0.89 ± 0.37 30	0.72 ± 0.46 24	0.45±0.29 17
Total	3	2.95	3.01	2.64

canonical axes was 0.002. Taken together, the relationships between Pro, Syn and PEuks and environmental variables (temperature, salinity, DIN, DIP and DSi) can be explained by the CCA ordination diagram (Fig. 7).

In the CCA ordination diagram, 3 picophytoplankton groups were positively correlated with water temperature (Pearson's correlation, r > 0.323, p < 0.01; Table 5), indicating that temperature was an important factor in regulating the biogeographic variations of picophytoplankton. A negative correlation was observed between salinity and Syn (r = -0.272, p < 0.01) and PEuks (r = -0.076, p < 0.05), indicating their coastal distributions especially in the SCS (Fig. 4). However, there was no significant correlation between salinity and Pro abundance (r = 0.031, p > 0.05), thus indicating that the variation in salinity seemed to have little control over the abundance of total picophytoplankton due to the absolute dominance of Pro abundance (Table 3, Fig. 4). The 3 picophytoplankton groups showed a negative correlation with nutrients (r < -0.261, p < 0.01), indicating that the abundances of Pro, Syn and PEuks declined with the increase in nutrient concentrations and were thus found to have a maximum in oligotrophic waters (Fig. 4). This result also indicated that picophytoplankton growth was not constrained by low nutrient levels. Consequently, water temperature and nutrient availability were important factors explaining the biogeographic variations of picophytoplankton, whereas salinity explained relatively little.

4. DISCUSSION

4.1. Abundance, spatial distribution and associated environmental variables

Our dataset differs from previous efforts in having a wider biogeographic and environmental representation. Although Ribeiro et al. (2016) suggested that the low concentrations of photosynthetic pigments in Pro cells in surface waters may lead to an underestimation of abundance by the BD Accuri FCM, the average Pro abundances observed in the 3 contrasting seas are of the same magnitude as previous studies in other marine ecosystems (e.g. Flombaum et al. 2013, van den Engh et al. 2017, Wei et al. 2019a, Zhao et al. 2019). In addition, Pro cells in deeper waters containing relatively higher concentrations of pigments have allowed better detection on the BD Accuri FCM (Ribeiro et al. 2016); thus, this underestimation does not affect the spatial distribution patterns of abundance and biomass for Pro demonstrated herein (Bergo et al. 2017). Given the high sensitivity of our BD Accuri FCM after a series of instrument settings, however, we speculate that the potential disappearance of Pro at those depths is rather a reflection of the already low abundances found in shallower depths compared with other studies than a problem with the detection limit. To date,

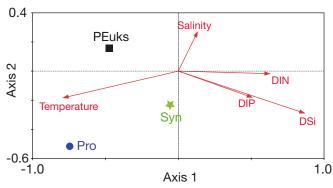


Fig. 7. Canonical correspondence analysis (CCA) plot for the relationship between picophytoplankton abundance and environmental variables across 3 contrasting seas (n = 882). The colored symbols correspond to 3 picophytoplankton groups (i.e. Pro: *Prochlorococcus*; Syn: *Synechococcus*; PEuks: picoeukaryotes). The vectors show the direction and strength of the environmental variables relative to the overall distribution. Picophytoplankton abundances were log transformed prior to analysis. DIN: dissolved inorganic nitrogen; DIP: dissolved inorganic phosphorus; DSi: dissolved inorganic silicate

the BD Accuri FCM remains a major instrument for marine picophytoplankton research. For instance, Bergo et al. (2017) examined the spatial dynamics of abundance and biomass partitioning of Pro, Syn and PEuks across environmental gradients using a BD Accuri FCM; both Brandini et al. (2019) and Zhao et al. (2019) detected the Pro, Syn and PEuks abundances on the BD Accuri FCM to resolve the functional relationships between picophytoplankton abundances and oceanic properties.

The average abundances of total picophytoplankton observed for BOB, SCS and WPO within our study are comparable with other estimates (Table 3). For example, our estimated abundance of total picophytoplankton for the WPO $(2.5 \times 10^4 \text{ cells ml}^{-1})$ was in good agreement with previously reported abundance of $(2.0-3.8) \times 10^4$ cells ml⁻¹ (Landry et al. 2003, Cheung et al. 2008); Madhupratap et al. (2003) and Cai et al. (2007) estimated the total picophytoplankton abundances of $(0.2-18.5) \times 10^4$ and 5.1×10^4 cells ml⁻¹, respectively, for the BOB and SCS, which were also close to our estimates of 4.7×10^4 cells ml⁻¹. However, the estimated average abundances of Pro, Syn and PEuks in the 3 contrasting seas showed some differences with the average abundances reported by Agusti et al. (2019) (Table 6). The average Syn abundance in the WPO $(4.2 \times 10^3 \text{ cells ml}^{-1})$

Table 5. Pearson's rank correlation coefficients between environmental factors and abundances of 3 picophytoplankton groups (n = 882). Picophytoplankton abundances were log transformed prior to analysis. Syn: Synechococcus; Pro: Prochlorococcus; PEuks: picoeukaryotes; DIN: dissolved inorganic nitrogen; DIP: dissolved inorganic phosphorus; DSi: dissolved inorganic silicate. *p < 0.05 (2-tailed); **p < 0.01 (2-tailed)

	Temperature	Salinity	DIN	DIP	DSi
Syn Pro PEuks	0.640** 0.323** s 0.535**	0.031	-0.316**	-0.339** -0.261** -0.374**	-0.467**

Table 6. Mean ± SD abundances of 3 picophytoplankton populations (Syn: *Synechococcus*; Pro: *Prochlorococcus*; PEuks: picoeukaryotes) in each ocean basin and globally (Agusti et al. 2019)

(Pro 10 ⁴ cells ml ⁻¹)	Syn $(10^3 \text{ cells ml}^{-1})$	PEuks (10 ³ cells ml ⁻¹)
Atlantic Ocean	17.2 ± 0.8	4.9 ± 0.4	1.5 ± 0.1
Indian Ocean	9.6 ± 0.6	8.1 ± 1.5	1.3 ± 0.1
Pacific Ocean	8.3 ± 0.3	11.4 ± 2.4	2.1 ± 0.3
Global scale	12.6 ± 0.4	7.7 ± 0.8	1.7 ± 0.1

was 2- to 3-fold less than that $(11.2 \times 10^3 \text{ cells ml}^{-1})$ observed by Agusti et al. (2019) in the Pacific Ocean, whereas the average Syn abundance in the BOB $(10.1 \times 10^3 \text{ cells ml}^{-1})$ was slightly higher than that in the Indian Ocean $(8.1 \times 10^3 \text{ cells ml}^{-1})$. The average Pro and PEuks abundances in the BOB and WPO also differed from those in the Indian Ocean and Pacific Ocean, respectively.

Syn and PEuks are able to tolerate low temperature and are most abundant in coastal regions of the SCS with intermediate nutrient concentrations (Zubkov et al. 2000, Liu et al. 2007, Zhang et al. 2013). In the present study, Syn and PEuks abundances were inversely associated with salinity (Fig. 7). It is likely that salinity is a function of the freshwater input, which directly determines the high abundances of Syn and PEuks in coastal regions (Olson et al. 1990, Lee et al. 2014). As such, Syn and PEuks abundances were higher near the freshwater-dominated areas of the SCS (Fig. 4), which is likely due to the mesotrophic dilution waters (Zhang et al. 2013, Zhao et al. 2019).

Pro were the most abundant picophytoplankton group in the 3 contrasting seas (Table 3). Because of the predominance of Pro, the maximum carbon biomass of total picophytoplankton also occurred in the CSCS (Fig. 6), thus indicating the ecological importance of Pro as a primary producer. Pro were present in high abundances at low nutrient concentrations (Fig. 4), consistent with adaptation to growth under low nutrient availability (Olson et al. 1990, Zubkov et al. 2000, van den Engh et al. 2017, Wei et al. 2019b). It is usually believed that elevated nutrient concentrations and heavy metals induced by coastal currents or upwellings could be toxic to Pro growth (Mann et al. 2002, Gaulke et al. 2010, Lee et al. 2014). As a result of these toxic effects, Pro abundance was usually high in the oligotrophic open waters (Fig. 4). Water temperature was also one of the key factors limiting the coastal distribution of Pro (Fig. 7); however, this response to water temperature is consistent with previous reports (Moore et al. 1995, Zinser et al. 2007, Agusti et al. 2019). For example, Pro growth is generally limited below water temperatures of 15-18°C and inhibited at water temperatures above 25°C (Moore et al. 1995, Buck et al. 1996). Interestingly, Pro were exceptionally abundant along the NSCS coast other than in the CSCS (Fig. 4). The hydrographic conditions near the NSCS coast, characterized by high temperature and salinity, were apparently affected by the Kuroshio intrusion (Fig. 2). We therefore confirmed that the exceptional presence of Pro near the coast of the SCS was potentially associated with the Kuroshio intrusion. Analogous studies have

shown that the effect of the Kuroshio intrusion potentially resulted in the exceptional presence of Pro near the coast (Chen et al. 2011, Lee et al. 2014). Collectively, Pro are most abundant in the oligotrophic open waters, but they are by no means restricted to this habitat. In addition, light, water turbidity and disturbance, competition among groups, ecotype succession and grazing pressure are all important controlling factors (Landry et al. 2003, Grébert et al. 2018, Wei et al. 2019a). The partial increase in nutrient availability particularly in the oligotrophic open oceans can also stimulate the Pro growth (discussed below), indicating that some undiscovered factors may also influence the biogeographic variations of picophytoplankton at the global scale. However, our research is not fine-scaled enough to resolve the close associations between physiological features and biogeographic distributions of picophytoplankton; more data are needed.

4.2. Physical processes in regulating the biogeographic variation of picophytoplankton

Three picophytoplankton populations showed negative correlations with nutrient concentrations (Fig. 7, Table 5), thus high nutrient concentrations seem to restrict their coastal distributions. The same conclusion was reached by Gaeta et al. (1999) and Brandini et al. (2019), who also find a negative correlation between the dynamics of picophytoplankton abundance and nutrient concentrations. However, comparison with other observations indicates that nutrient availability may be a major factor responsible for the coastal distributions of picophytoplankton, especially for the Syn and PEuks (Jiao et al. 2005, Liu et al. 2007, Zhang et al. 2013, Zhao et al. 2019). On the other hand, the maximal abundances of 3 picophytoplankton groups in vertical distribution showed strong positive correlations with the nutricline depth (Figs. 3 & 5). Therefore, nutrient availability has a 2-faceted role in regulating the spatial patterns of picophytoplankton in different marine ecosystems. Light irradiance is a key limiting factor for the temporal-spatial distributions of picophytoplankton (Flombaum et al. 2013, Agusti et al. 2019). Recent studies suggested that Pro are more sensitive to sunlight, particularly to UV (Llabrés & Agustí 2006), but this does not occur for Syn. Accordingly, Pro are better adapted to capture the blue wavelengths that predominate deeper in the water column (Moore et al. 1995, Dusenberry et al. 2000, Grébert et al. 2018). Due to the apparent stratification in the tropical and subtropical oligotrophic gyres,

nutrients were almost depleted within the euphotic zone, whereas nutrient supply was sufficient in the deeper layer (Fig. 3). Picophytoplankton abundance was not higher under conditions with high nutrients below the euphotic layer, which may be reasonably driven by light limitation. Therefore, light availability may help explain this 2-faceted role of nutrients in regulating the biogeographic variation in picophytoplankton. Several investigations have revealed that 2 ecotypes of Pro can be adapted to grow at different light intensities (e.g. Zinser et al. 2007), but we did not observe the co-occurrence of different Pro on our BD Accuri FCM analysis.

In the absence of physical input of new nutrients, particularly for oligotrophic waters, nutrient availability is significant for the picophytoplankton growth. Previous research on physical processes has indicated that Pro, Syn and PEuks alternatively dominate in different water masses or circulations, because their hydrographic properties such as temperature, salinity and nutrients can be important controlling factors for picophytoplankton distributions (Agustí 2004, Flombaum et al. 2013). For example, the exceptional presence of Pro near the SCS coast may be potentially caused by the effect of the Kuroshio intrusion (Jiao & Yang 2002, Chen et al. 2011, Lee et al. 2014); high Syn and PEuks abundances are observed in areas dominated by coastal currents or upwelling regions (Chen et al. 2011). In particular, picophytoplankton within an eddy have different biogeographic characteristics than outside the eddy (Chen et al. 2015). Processmodel studies indicated that both vertical and lateral nutrient supplies by mesoscale eddies are considerably effective for picophytoplankton growth near the margins of the oligotrophic gyre (Oschlies 2002). Eddy-related nutrient injections within the euphotic zone, especially cold eddies, can significantly enhance biological production in the oligotrophic waters (Mc-Gillicuddy & Robinson 1997, Vaillancourt et al. 2003). Šantić et al. (2011) also highlighted that the abundance of Pro can be influenced by nutrient availability, but as a consequence of the movement of cold eddies. The sea level anomaly and the hydrographic conditions at 10° N in the BOB were consistent with the presence of a cold eddy (Figs. 2 & 3). Thus, Pro abundance was relatively high around the 10°N in the BOB (Fig. 4) because of the relatively high nutrient concentrations supplied by the cold eddy. Overall, cold eddies play an important role in influencing biogeographic distributions of picophytoplankton because of eddy-induced upward nutrient pumping. Inversely, the biogeographic variations in picophytoplankton may be an indicator for water masses or circulations.

4.3. Size-derived conversion factors and carbon biomass

There is significant uncertainty over carbon biomass of picophytoplankton, stemming from the uncertainties in both cell size and abundance-to-carbon conversion factors. Within the present study, we used a conversion factor of 964 fg C cell⁻¹ for PEuks in the WPO (Table 1). However, this conversion factor may provide a lower estimation of carbon biomass for PEuks than previous biomass values in the WPO. Liu et al. (2007) also indicated that the estimated PEuks cell size (1.31 \pm 0.30 μm) in the SCS is somewhat smaller than other estimates, thus resulting in a low conversion factor (293 fg C cell⁻¹; Table 1). Overall, the main reason that our estimates of carbon biomass of PEuks may be on the low end of all the estimates is that we have adopted newly derived conversion factors; however, we believe it is a more accurate carbon density factor than most previously used values.

Although the carbon biomass was much higher in the SCS and WPO than in the BOB, it is essential to emphasize that these calculated carbon biomasses are not applied to compare the biogeographic variation, which is principally due to great plasticity in the conversion factor under different taxonomic and ambient conditions. Meanwhile, it is not meaningful to directly compare our biomass estimates with previous reports, since different carbon conversion factors were used in different studies.

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