Infection dynamics of *Lernaeocera lusci* on sand goby *Pomatoschistus minutus* in the Oosterschelde (The Netherlands)

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**ABSTRACT:** Male and female sand gobies *Pomatoschistus minutus* (Pallas) from the Oosterschelde (The Netherlands) were examined for the crustacean mesoparasite *Lernaeocera lusci* (Bassett-Smith) from April to July 1992. Prevalence increased steeply from 4% in April to 80% (females) and 57% (males) in June. The peak abundance of recently attached infective stages (pennella larvae) on sand gobies was found in May. By June, about 70% of parasites were in the post-metamorphosis stage and possessed egg strings. Significant differences in abundance between males and females were found in June but not in May. This may be due to sex-specific differences in behaviour during spawning time. The majority of post-spawning fish probably died in July. Fish of the new year class which were captured in July harboured no parasites.

**KEY WORDS:** *Pomatoschistus minutus* · *Lernaeocera lusci* · Parasitism · Oosterschelde

**INTRODUCTION**

*Lernaeocera lusci* (Bassett-Smith) is a crustacean parasite which uses sole *Solea solea* as intermediate host. Though bib *Trisopterus luscus* is the typical definitive host, adult females can also be found attached to many other demersal fish species (Boxshall 1974, Kabata 1979, Tirard 1991). *L. minuta*, which was previously distinguished from *L. lusci* on grounds of its small size (Kabata 1979), can be equalled to *L. lusci* because of the host-dependency of parasite size (Van Damme et al. 1993). Sand goby *Pomatoschistus minutus* (Pallas), which was previously considered to be the typical host species of *L. minuta*, thus appears on the host list of *L. lusci*.

High prevalences of *Lernaeocera minuta* (henceforth *L. lusci*) have been recorded on sand gobies along the German coast (86% by Mann 1964, 22% by Petersen 1992) and along the Belgian coast (39% by Hamerlynck et al. 1989). In the Belgian as well as in the southwestern Dutch coastal area, 2 transmission waves could be distinguished (Hamerlynck et al. 1989): a first wave occurred in late autumn when juvenile sand gobies were infected, and a second transmission wave occurred in the following spring when mature, spawning sand gobies were infected.

Mann (1964) and Petersen (1992) found that *Lernaeocera lusci* was responsible for a significant decrease in the condition factor of sand gobies. Moreover, both fat content of the liver and hemoglobin content were lower in infected gobies as compared to non-infected individuals (Mann 1964). However, neither study provided a detailed account of the dynamics of the host and parasite populations, though this may be essential for a better understanding of the interaction between host and parasite individuals. For example, fish size and fish sex were not considered, both of which may have acted as confounding variables in the investigations.

In this study, we provide further details on the infection dynamics of *Lernaeocera lusci* during the spring transmission wave in a tidal bay in the southwestern North Sea in 1992. In a future paper the effect of *L. lusci* on hematological parameters will be presented.

**MATERIAL AND METHODS**

Sand goby samples were collected in the western part of the Oosterschelde, a tidal bay in the southwest Netherlands. Surface water temperatures measured at
Table 1. *Pomatoschistus minutus*. Sampling dates, water temperature, number of male, female and juvenile sand gobies caught, sex ratios (male:female) and mean total lengths (TL ± SD) in the Oosterschelde in 1992. The year class to which the fish belonged is indicated in parentheses. *F* : ANOVA to test for differences in mean length between males and females (ns = not significant).

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<tr>
<td></td>
<td></td>
<td>n</td>
<td>TL</td>
<td>n</td>
<td>TL</td>
<td>n</td>
</tr>
<tr>
<td>6 Apr</td>
<td>9</td>
<td>17</td>
<td>65.4 ± 6.4</td>
<td>16</td>
<td>63.2 ± 5.5</td>
<td>0</td>
</tr>
<tr>
<td>28 Apr</td>
<td>11</td>
<td>8</td>
<td>65.5 ± 14.8</td>
<td>10</td>
<td>62.7 ± 2.2</td>
<td>0</td>
</tr>
<tr>
<td>15 May</td>
<td>12</td>
<td>38</td>
<td>66.3 ± 8.0</td>
<td>81</td>
<td>63.7 ± 6.1</td>
<td>0</td>
</tr>
<tr>
<td>26 May</td>
<td>15</td>
<td>1</td>
<td>64.0</td>
<td>61</td>
<td>60.4 ± 7.0</td>
<td>0</td>
</tr>
<tr>
<td>11 Jun</td>
<td>16</td>
<td>32</td>
<td>65.8 ± 7.4</td>
<td>46</td>
<td>63.6 ± 5.3</td>
<td>0</td>
</tr>
<tr>
<td>4 Jul</td>
<td>18</td>
<td>3</td>
<td>62.2</td>
<td>1</td>
<td>64.1</td>
<td>31</td>
</tr>
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On 6 April, 28 April, 26 May and 4 July the samples were taken with a commercial shrimp trawler using a 5 m beam trawl. The samples of 15 May and 11 June were taken with a research vessel using a 3 m beam trawl. All fish collected (except for some juvenile fish of the 1992 year class caught on 4 July) belonged to the 1991 year class. Due to the use of different sampling methods (e.g. differences in mesh size) the fish numbers do not reflect true densities.

Prior to statistical analysis GSI was arcsine transformed.

\[
\text{GSI} = \frac{\text{gonad weight}}{\text{total body weight} - \text{weight stomach content}}
\]

The female parasites which were attached to the gill arches or in the gill cavity were counted and removed from the host. They were assigned to one of 8 developmental stages, based on the staging system of Van Damme & Hamerlynck (1992) (Table 2). Prevalence (percentage of fish infected), abundance (mean number of parasites per fish), intensity (number of parasites on 1 host individual) and mean intensity (mean number of parasites per infected fish) were used according to the recommendations of Margolis et al. (1982). The relationship between the dependent variable parasite intensity (y) and independent variables fish length (x₁) and GSI (x₂) of females on 13 May and 11 June was analysed by a multiple regression analysis:

\[
y = a + b₁x₁ + b₂x₂
\]

where a is the intercept, and b₁ and b₂ are regression coefficients.

Table 2. Classification of adult female *Lernaeocera lusci* on *Pomatoschistus minutus* (after Van Damme & Hamerlynck 1992)

<table>
<thead>
<tr>
<th>Substage</th>
<th>Definition</th>
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<tbody>
<tr>
<td>Pennella (P1)</td>
<td>Straight body, no flexure</td>
</tr>
<tr>
<td>Pennella (P2)</td>
<td>One point of flexure</td>
</tr>
<tr>
<td>Immature (U)</td>
<td>Two or three points of flexure</td>
</tr>
<tr>
<td>Mature pregravid (W)</td>
<td>Genital region not swollen</td>
</tr>
<tr>
<td>Mature gravid (X)</td>
<td>Genital region fully swollen</td>
</tr>
<tr>
<td>X1</td>
<td>No external egg strings</td>
</tr>
<tr>
<td>X2</td>
<td>Immature eggs</td>
</tr>
<tr>
<td>Y</td>
<td>Mature pigmented eggs</td>
</tr>
<tr>
<td>Dead parasite (Z)</td>
<td>Remains of holdfast embedded in host tissue</td>
</tr>
</tbody>
</table>

**RESULTS**

All fish collected before July belonged to the 1991 year class (1+). In July this year class had all but disappeared and was replaced by fish belonging to the 1992 (0+) year class. The numbers of female and male fish caught and sex ratios are given in Table 1. Between 4 April and 26 May the proportion of females in the catches increased progressively.

In all months, females were smaller than males, but these differences were not significant (ANOVA, p > 0.05) (Table 1). Juveniles of year class 1992 which were collected on 4 July were significantly smaller than the 1+ females (ANOVA; F = 166, p < 0.001) and the 1+ males (ANOVA; F = 229, p < 0.001) of June 1992 (Table 1).

Prevalence and abundance of *Lernaeocera lusci* on female and male sand gobies are shown in Fig. 1. Prevalence steeply increased between 28 April and 15 May and then gradually increased to maximal levels of 77% (females) and 53% (males) in June. For both
sexes there was a similar pattern for abundance: between the beginning of April and the end of June abundance increased from 0.06 to 1.77 (females) and from 0 to 1.42 (males). There were no significant differences in abundance between males and females on 28 April (Kruskal-Wallis; $H = 0.23; p > 0.05$) and on 15 May ($H = 0.27; p > 0.05$). However, female sand gobies harboured significantly more parasites than males on 11 June ($H = 4.03; p < 0.05$). Fish with more than 4 parasites were found in May and June (Fig. 1).

Fish length and parasite intensity were positively correlated on 15 May and on 26 May for both males and females. On 28 April and 11 June the correlations between these 2 variables were not significant ($p > 0.05$) (Table 3).

The mean GSI for female gobies was significantly lower on 11 June ($0.05 \pm 0.05$) than on 9 April ($0.08 \pm 0.03$) and 15 May ($0.09 \pm 0.05$) (Tukey test, $p < 0.05$) (Fig. 2). The correlations between fish length and GSI were not significant on 15 May ($n = 44, R = -0.24, p > 0.05$) and on 11 June ($n = 37, R = 0.93, p > 0.05$).

Overall, the multiple regression equations between parasite intensity, GSI and fish length were significant on 15 May ($p < 0.01$) but were not significant on 11 June ($p > 0.05$). Neither fish length nor GSI contributed significantly in the prediction of parasite intensity on 11 June (Table 4).
The population structure of *Lernaeocera lusci* was characterised by a clear temporal pattern (Fig. 3). Juvenile stages (P1 and P2) were mainly found on 28 April and on 15 May, whereas adult female parasites with egg strings were found mainly after 28 May. This indicates that transmission occurred mainly prior to the latter dates. There was no clear temporal pattern in the occurrence of X1, X2 and Y substages. Overall, the majority (84%) of adult female parasites belonged to the X1 substage.

**DISCUSSION**

After a short pelagic stage of approximately 1 mo, sand gobies start to live in close proximity to the bottom (Fonds 1973), and become increasingly susceptible to infection by *Lernaeocera lusci*. In late autumn in the Belgian and Dutch coastal area, a small proportion (± 10%) of this demersal sand goby population is infected with *L. lusci* (Hamerlynck et al. 1989). These latter authors recorded no further infection in winter and a second, dramatic, rise in infection level during spawning time (May/June). The males of *Pomatoschistus minutus* build nests under bivalve shells which they cover completely with sand. They guard the eggs of 1 or more females until hatching (Fonds 1973, Hesthagen 1977). During egg guarding, male *P. microps* exhibit low feeding activities and hence reduced mobility (Maghnagen 1990). Fonds (1973), Magnhagen & Kvarnemo (1989) and Hamerlynck & Cattrijsse (in press) suggested this may also be the case in *P. minutus*. Furthermore, Hamerlynck & Cattrijsse (in press) found that males consume predominantly *Pomatoschistus* eggs, polychaetes, bivalves and *Gastrosaccus*, which is a benthic mysid species. The hypothesis that males are often hiding in their nests (Hesthagen 1977, Maghnagen 1990) may also explain why sometimes small numbers of males are caught in early summer (Swedmark 1958). In the present study this was the case on 26 May when 98% (n = 62) of the gobies were females (Table 1). On the other hand, females roam about during the spawning period (Hesthagen 1977, Hamerlynck & Cattrijsse in press) with consequent higher probability of becoming infected by *Lernaeocera lusci* than the males. These sex-specific behavioural differences probably explain the different probabilities of males and females accumulating *L. lusci*.

Transmission of pennella larvae to sand gobies in spring is restricted to a very short period. Indeed, P1
and P2 stages were predominantly found in 1 sample (May), whereas the June samples contained mainly mature parasites. The most plausible explanation for this apparently narrow transmission window was recently given: it was found that all infective stages of Lernaeocera lusci leave sole in a very short period, probably triggered by rising temperatures (Van Damme & Ollevier unpbl.). Based on the present study we can provide some estimates for the duration of the life cycle stages of L. lusci. The majority of the parasites which infected sand gobies after 28 April were mature by 11 June, suggesting a duration of 1 to 2 mo from initial infection to maturation.

In the present study no evidence for parasite-induced host mortality nor for a relationship between parasite intensity and mating success of sand gobies was found. Hosts with more than 5 parasites were found at all sampling dates in May and June (Fig. 1), suggesting that heavily infected hosts suffer no high mortality during this sampling period. Hamerlynck et al. (1989) suggested that Lernaeocera lusci may further exacerbate the lowered condition of post-spawning sand gobies and hence induce post-spawning host mortality, but this could not be tested due to the small sample size in July. Furthermore, no significant correlation was found between the gonadosomatic index and the parasite intensity of females on 15 May and 11 June (Table 4). However, field sampling may be inadequate to find such a relationship. Probably, the female sand gobies are infected with L. lusci at a time when they have already released 1 or more batches of eggs. The effect of these parasites on the reproductive success of the sand gobies, if present, may therefore be obscured by many confounding variables.

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**LITERATURE CITED**


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