Changes in splenic melano-macrophage centres of dab *Limanda limanda* during and after infection with ulcer disease

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ABSTRACT: Splenic melano-macrophage centres (MMC) and associated parenchymic structures were compared in 3 groups of dab *Limanda limanda* L. from the North Sea: healthy fish, fish with ulcers, and fish with healed ulcers still recognizable as scars on the body surface. Dab with open ulcers showed enlarged splenic MMC somewhat reduced in frequency and with increased haemosiderin content. During this stage of infection the density of pigmented granules within MMC was reduced in favour of homogeneously scattered cells or a centrally located aggregation of cells, all of which lacked phagocytic inclusions. Leucocytes were accumulated to an elevated degree around splenic capillaries. Because of the reduced occurrence of single pigment-bearing cells in the splenic parenchyma and the very regular, rounded shape of the MMC, it is thought that during this stage of the disease the turnover rate in formation and destruction of MMC was diminished. On the other hand, the increase in seemingly non-phagocytic leucocytes in the MMC indicates a possible augmentation of the humoral immune response. After healing of the ulcers all splenic changes accompanying infectious conditions regressed. However, a return to the normal picture seen in healthy fish did not occur as long as scars were still visible in the skin.

INTRODUCTION

Melano-macrophage centres (MMC) in haematopoietic organs of bony fish occur in head kidney, spleen, and, more rarely, in the liver. They are aggregations, mainly of macrophages, in which phagocytosed substances are catabolized, remobilized, or deposited. According to the varying proportions of melanin and lipofuscin contained, their coloration varies between light yellow and nearly black in histological sections. Increased occurrence of melanin in poikilothermic animals is attributed to the catabolism of fatty acids at low temperatures (Edelstein 1971). Lipofuscin is considered to be a catabolic product resulting from the destruction of cellular components, especially biological membranes (Agius & Agbede 1984). Another pigment that is abundant in MMC, named haemosiderin, derives from the catabolism of erythrocytes.

In addition to phagocytosing foreign particles or effete cells from an organism, MMC are involved in immunological functions. For this reason they have been interpreted as precursors of the germinal centres in the lymph nodes of higher vertebrates (Ellis 1980). Size, frequency, and pigmentation of MMC have been observed to vary with starvation, disease, pollution, and fish age (Agius 1979, Agius & Roberts 1981, Weeks & Warriner 1984, Brown & George 1985, Wolke et al. 1985, Kranz & Gercken 1987). Owing to these reactions, MMC have been discussed as indicators of the health status of fish in biological effect monitoring (Blazer et al. 1987). However, a clear distinction of MMC alterations caused by the different single influences mentioned has not yet been established. If MMC are to be useful as indicators of exposure to noxious agents, changes from the norm induced in the MMC by the agents should disappear once contact with the noxious agents is removed.

MMC vary between the different haematopoietic organs. Splenic MMC are more abundant and larger than those in the liver; compare to kidney, they are more variable in pigmentation and show a closer degree of aggregation (Agius 1979, Kranz & Peters 1984).

The present investigation examined alterations in splenic MMC of dab *Limanda limanda* L. from the North Sea: the MMC of healthy individuals were com-
pared with those of fish suffering from open ulcers and with those of fish that had recovered from the disease but in which scars were still visible on the body surface. Ulcer disease is a widespread infectious condition that has been recorded in 15 fish species from the North Sea (Dethlefsen 1984).

In cod Gadus morhua, which are frequently affected by ulcer disease, viruses have been considered as the primary infectious agent; invariably, however, infections with bacteria of the genus Vibrio follow (Jensen & Larsen 1982). The disease is characterized, especially in the chronic stage, by large, bloody skin ulcers, which usually extend into the underlying musculature. The borders of the ulcers contain necrotic tissue and macrophage aggregations (Roberts 1978).

### MATERIALS AND METHODS

In May 1986, 81 dab were taken from the Dogger Bank area in the North Sea. Thirty of them had open ulcers on the body surface, 21 had recovered from ulcerous inflammation, as seen from scars on the skin, and 30 reference fish showed no external or internal sign of disease. To avoid any influence of sexual diversity or gonadal maturation, only female dab between 19 and 26 cm in length were used (healthy fish: 21.8 ± 1.44 cm; ulcerous fish: 21.6 ± 1.85 cm; scarred fish: 22.2 ± 1.79 cm). The dab were killed and dissected, and the spleens were fixed with Bouin’s fluid. Organs were processed using routine histological methods to yield sections 3 µm in thickness. The following stains were used: haematoxylin and eosin as a general stain; Azan for identification of fibrous tissue elements; and Prussian blue to detect haemosiderin in MMC.

Quantitative changes in the number and size of the splenic MMC were determined by counting and by planimetry. The results were expressed in terms of the frequency of MMC in a defined splenic area, the average size of MMC per section of organ, and the percentage of organ area occupied by MMC. Frequency and size of splenic MMC in the three study groups were compared using the U-statistic of Wilcoxon, Mann and Whitney.

### RESULTS

The spleen of teleosts has generally been considered as storage organ for blood cells, and beyond that it has functions in haematopoiesis, destruction of blood cells, and immunological processes. The basic structure of the spleen is a network of reticular cells (Figs. 1 and 2), in which erythrocytes, leucocytes, pigment cells, and a widely ramified system of blood vessels are embedded.

This reticular stroma forms thicker wall-structures around terminal capillaries, called ellipsoidal sheaths (Fig. 3), the stroma sometimes encapsulates MMC, in which case the MMC appear as uniformly rounded structures (Fig. 4).

The MMC and single macrophages in the parenchyma are recognizable in histological sections because of their cytoplasmic pigments: black-brown melanin, yellow lipofuscin, and haemosiderin.

The splenic MMC of dab contained low amounts of melanin compared with other species (e.g. plaice) and their haemosiderin content and certain other of their features along with those of their associated parenchymal structures differed according to the study group: healthy, ulcerous, or scarred dab (see Tables 1 and 2). The most obvious feature in the spleen of dab with open ulcers on the body surface was an increase in size of the MMC. On average, the MMC were 50% larger than in normal fish (p < 0.001). This corresponded approximately to an overall increase in MMC area per organ section, because the frequency of MMC, although slightly reduced in number, was not significantly altered (p > 0.3).

Haemosiderin always occurred only in the macrophages assembled in the MMC (Figs. 5 and 6); single cells scattered in the splenic parenchyma between ellipsoids and MMC never showed positive reactions with Prussian blue. Haemosiderin in MMC occurred most frequently in the spleens of ulcerous fish (Table 2), although pigmented inclusions or phagosomes displaying very fine granulation were mostly loosely arranged in their MMC. No distinct differences in melanin content of splenic MMC were obvious in the 3 study groups.

In dab with open ulcers, the MMC often contained cells lacking cytoplasmic inclusions in addition to the pigment-bearing macrophages. The unpigmented cells contained a large, often round, nucleus with a marginal distribution of chromatin. These cells were either loosely scattered in the MMC or were concentrated in the centre of the MMC such that the phagosome- and/or pigment-containing macrophages were restricted to

<table>
<thead>
<tr>
<th>MMC</th>
<th>Healthy</th>
<th>Ulcerous</th>
<th>Scarred</th>
</tr>
</thead>
<tbody>
<tr>
<td>Size (µm²)</td>
<td>984.3±67.2</td>
<td>1422.7±103.0</td>
<td>1131.5±83.2</td>
</tr>
<tr>
<td>Area per organ area (%)</td>
<td>4.4±0.5</td>
<td>5.7±0.5</td>
<td>4.9±0.4</td>
</tr>
<tr>
<td>Frequency (n mm⁻³)</td>
<td>44.3±3.9</td>
<td>40.1±2.4</td>
<td>45.2±3.6</td>
</tr>
</tbody>
</table>

Table 1 Limanda limanda. Size, MMC area as percentage of organ area, and frequency of MMC in spleens of healthy, ulcerous, and scarred dab from the North Sea. Values shown are means ± standard error.
Table 2. *Limanda limanda*. Qualitative evaluation of histological characteristics associated with splenic MMC in healthy, ulcerous, and scarred dab from the North Sea. Percentage values refer to numbers of organs examined. Unless otherwise stated, evaluations refer to H&E staining of tissue sections.

<table>
<thead>
<tr>
<th>Histological characteristic</th>
<th>Healthy</th>
<th>Ulcerous</th>
<th>Scarred</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haemosiderin content of MMC (Prussian blue staining)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>33 %</td>
<td>13 %</td>
<td>19 %</td>
</tr>
<tr>
<td>Moderate</td>
<td>37 %</td>
<td>60 %</td>
<td>76 %</td>
</tr>
<tr>
<td>Strong</td>
<td>29 %</td>
<td>27 %</td>
<td>5 %</td>
</tr>
<tr>
<td>Arrangement of pigments within MMC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Compact</td>
<td>50 %</td>
<td>27 %</td>
<td>38 %</td>
</tr>
<tr>
<td>Scattered</td>
<td>13 %</td>
<td>23 %</td>
<td>30 %</td>
</tr>
<tr>
<td>Marginal</td>
<td>37 %</td>
<td>50 %</td>
<td>24 %</td>
</tr>
<tr>
<td>Distinct encapsulation of MMC (Azan staining)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Connective tissue extending through MMC (Azan staining)</td>
<td>56 %</td>
<td>79 %</td>
<td>67 %</td>
</tr>
<tr>
<td>Irregularly notched surface of MMC</td>
<td>11 %</td>
<td>32 %</td>
<td>19 %</td>
</tr>
<tr>
<td>Distinct RES^a* structures within splenic parenchyma (Azan staining)</td>
<td>89 %</td>
<td>64 %</td>
<td>71 %</td>
</tr>
<tr>
<td>Distinct ellipsoid sheaths</td>
<td>93 %</td>
<td>86 %</td>
<td>91 %</td>
</tr>
<tr>
<td>Aggregations of leucocytes surrounding ellipsoids</td>
<td>55 %</td>
<td>68 %</td>
<td>57 %</td>
</tr>
<tr>
<td>Single macrophages within splenic parenchyma^b</td>
<td>47 %</td>
<td>17 %</td>
<td>29 %</td>
</tr>
<tr>
<td>Single macrophages within blood vessels^b</td>
<td>30 %</td>
<td>17 %</td>
<td>29 %</td>
</tr>
</tbody>
</table>

^a RES: reticulo-endothelial system
^b Single macrophages were only evaluated if showing distinct coloration with yellow to brown pigments

The border of the MMC (Fig. 7). In Azan-stained splenic sections from ulcerous fish, reticular cells often occurred, stroma-like, within the MMC and also surrounded them by forming a thin capsule (Table 2). The irregular notched shape of the MMC, seen in normal fish, was seldom seen within diseased fish. Single macrophages, recognizable because of their yellow to brownish inclusions, occurred with reduced frequency within the parenchyma of ulcerous fish (cf. Figs. 5 and 6). The same held true for melanin-containing cells within the splenic blood vessels of these fish (Fig. 8).

In dab with open wounds, the prominent reticular structures observed within and surrounding MMC contrasted with the less well developed reticular stroma within the splenic parenchyma and the ellipsoidal sheaths associated with the blood vessels. In these fish, leucocyte accumulations were abundant around ellipsoids.

All alterations of cellular splenic components noted during the open ulcer phase were clearly less pronounced after healing of the ulcers (Table 2). However, as long as scars were visible on the body surface most of the altered parameters had not yet returned to the values observed in the fish considered healthy. Statistically, the MMC size differed significantly between ulcerous and scarred fish ($p < 0.025$) but not between healthy and scarred fish ($p > 0.06$). The frequency of MMC was never significantly different between any 2 of the 3 studied groups.

**DISCUSSION**

In dab with ulcer disease the proportion of the spleen occupied by MMC was increased.

The mechanism of macrophage settlement in the spleen of fish has been revealed by means of injected carbon particles (Ferguson 1976, Lamers & Parmentier 1985). The carbon first appeared in the ellipsoidal sheaths and then, ingested by single macrophages presumably deriving from the reticular cells surrounding the blood vessels, migrated through the parenchyma and finally settled within existing or newly formed MMC.

The increase of splenic MMC tissue during ulcerous infection of dab was due to an increase in area of the MMC rather than an increase in their frequency. This means that newly settled splenic macrophages aggregated with already existing MMC. The almost negligible reduction in frequency of MMC during disease may have been due to a fusion of some MMC.

Earlier investigations have shown that MMC both in
Figs. 1 to 4. *L. limanda*. Fig. 1. Parenchymal structure of spleen, with blood vessels (large arrow), reticular stroma (white arrow), ellipsoidal sheaths (small arrow), erythrocytes, and nuclei of leucocytes. Azan, 500 x. Fig. 2. Large splenic MMC of an ulcerous dab surrounded by reticular fibres. Large arrow: cells of the reticular stroma; small arrow: phagosomes. Azan, 500 x. Fig. 3. Small splenic MMC of a healthy dab. MMC surface is irregular, and phagosomes are densely packed. Arrows: ellipsoidal sheaths. H&E, 300 x. Fig. 4. Large splenic MMC of an ulcerous dab. MMC surface is regular, and phagosomes are smaller and more loosely packed. H&E, 300 x.
Figs. 5 to 8. *Limanda limanda*. Fig. 5. Small splenic MMC of a healthy dab showing negative Prussian blue staining (phagocytic inclusions show yellow to brown coloration). Arrows: migrating macrophages in splenic parenchyma. Prussian blue, 300 x. Fig. 6. Large encapsulated splenic MMC of an ulcerous dab with positive proof of haemosiderin (phagocytic inclusions show dark blue coloration). Prussian blue, 300 x. Fig. 7. Splenic MMC from an ulcerous dab. Centre of MMC is free of macrophages containing phagocytic inclusions. Nuclei of cells in this area are very large, with marginally distributed chromatin. Arrow: capsule of the MMC; bv: blood vessel. H&E, 500 x.

Fig. 8. Macrophages with large phagocytic inclusions within a large splenic blood vessel. An MMC is situated next to the vessel endothelium (arrow). H&E, 500 x.
The investigation has shown that during the chronic ulcer disease has been overcome. Pigmented macrophages or concentrated in the centre structures in the spleen of dab for reorganization after cells were either interspersed among the aggregated occurrence of cells lacking pigment deposits. These
tion. Perhaps this time corresponds to the period, increase in size of splenic MMC, is the increased present investigation, which is connected with the antibodies were still demonstrable 12 mo after injec-
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The second important observation recorded in the present investigation with dab clearly show that all changes in splenic parameters that have occurred during ulcerous infection are attenuated after healing of the wounds. However, the levels found in healthy fish are not totally regained as long as scars are visible. This interval between healing of ulcers and disappearance of visible scars extends over about 1 yr after healing of the wounds. However, the levels found

Haemosiderin is derived from catabolism of haemoglobin from effete erythrocytes and is an intermediate metabolic product that occurs during recycling of components for erythropoiesis. There are 2 possible mechanisms by which the augmented haemosiderin-iron content may have come about: (1) the increased catabolism of damaged erythrocytes caused, for example, by lytic toxins from infectious agents; and (2) the increased retention of iron within MMC as a protective mechanism. It is known that in mammals a reduction of the iron content in blood serum results in a bacteriostatic effect (Bullen 1981).

The second important observation recorded in the present investigation, which is connected with the increase in size of splenic MMC, is the increased occurrence of cells lacking pigment deposits. These cells were either interspersed among the aggregated pigmented macrophages or concentrated in the centre of the MMC. Because an unequivocal identification of this cell type is impossible using light microscopy it can only be speculated that they represented immuno-competent cells with functions other than phagocytosis. Lamers (1986) reported that parallel to the macrophage-associated transport of injected antigen (which followed the same route as injected carbon particles) pyroninophilic cells appeared in the splenic parenchyma and in the MMC. These cells were plasma cells or precursor cells of haemo-, lympho- or granulopoiesis. If these pyroninophilic cells are identical to the cells predominantly occurring in the MMC of dab with ulcers this would indicate that an important additional function of MMC during this last, chronic, stage of infection is the production of immuno-competent cells in a manner analogous to that of the germinal centres of lymph nodes in mammals.

During enhancement of phagocytic activity, the spleen of fish is characterized by an augmented migration of macrophages within the parenchyma between ellipsoids and MMC (Ferguson 1976). Destruction of MMC is accompanied by the occurrence of pigmented macrophages within blood vessels which possibly derive from fragmentation of the MMC (Kranz & Gercken 1987). In both cases MMC have lost their regular rounded surface structure. In the splenic parenchyma and blood vessels of ulcerous dab, the frequency of macrophages was reduced. In parallel, the surface of the MMC exhibited a regular shape. These observations can be interpreted as the result of at least a non-elevated turnover of phagocytic cells in the formation and destruction of MMC. Possibly such augmented activity might have happened mainly during an earlier stage of disease.

The results of the present investigation with dab clearly show that all changes in splenic parameters that have occurred during ulcerous infection are attenuated after healing of the wounds. However, the levels found in healthy fish are not totally regained as long as scars are visible. This interval between healing of ulcers and disappearance of visible scars extends over about 1 yr (B. Watermann pers. comm.). Up to now, the literature has not included any explicit statement on the extent to which alterations in MMC decline after interruption of the noxae. The only investigations dealing with the fate of MMC after their stimulation are from Herraez & Zapata (1986) who describe a fragmentation of MMC following an immense enlargement induced by intoxication of the fish with phenylhydrazine, and from Lamers & De Haas (1985) who discovered that injected antibodies were still demonstrable 12 mo after injection. Perhaps this time corresponds to the period, needed by the MMC and associated parenchymal structures in the spleen of dab for reorganization after ulcer disease has been overcome.

The investigation has shown that during the chronic
phase of ulcer disease the size of splenic MMC is increased even though the main process of enlargement may occur earlier. The MMC changes in ulcerous fish tend to disappear with the removal of the noxae, although this process may be a long and drawn out one.

LITERATURE CITED


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