

NOTE

Streptococcus iniae type II infections in rainbow trout *Oncorhynchus mykiss*

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ABSTRACT: Clinical and pathological findings (anorexia, hemorrhage, lethargy, loss of orientation and exophthalmia) indicated that *Streptococcus iniae* type II is responsible for a fatal disease in rainbow trout. Histopathological findings revealed that *S. iniae* type II produces a systemic disease, including a diffuse necrotizing myositis. The distribution of viable bacteria in infected tissues substantiated the pathological findings, confirming that *S. iniae* type II is responsible for a generalized septic disease of rainbow trout.

KEY WORDS: Rainbow trout · *Streptococcus iniae* · Serotype · Pathology

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INTRODUCTION

Streptococcus iniae was first described as a pathogen of fish in 1994, when it was cultured from diseased tilapines *Oreochromis* spp. and rainbow trout *Oncorhynchus mykiss* in Israel (Eldar et al. 1994, 1995). A year later the same agent was isolated in the USA from infected tilapines and striped bass *Morone saxatilis* (Perera et al. 1994), suggesting the pathogen and its associated disease were widely distributed. Although the North American and Israeli isolates cluster in 2 distinct epidemiological clones (Eldar et al. 1997b), the pathological outcomes are similar (Eldar et al. 1995, Perera et al. 1994, Eldar & Ghittino 1999), with losses that often exceed 50% of the population. Increased awareness and improved identification schemes have subsequently demonstrated that *S. iniae* is present in different parts of the world and among various fresh- and saltwater fish species (Zlotkin et al. 1998, Ferguson et al. 2000, Bromage & Owens 2002, Colorni et al. 2002). Genetic profiles that characterize virulent and environmental strains have also been revealed (Weinstein et al. 1997, Fuller et al. 2001).

S. iniae has also been isolated from clinical specimens of human blood, urine and skin (Weinstein et al. 1997, Lau et al. 2003), in which it was associated with infections that can have a fatal course. *S. iniae* is, therefore, an emerging pathogen of increased clinical significance.

In *Streptococcus iniae*-infected rainbow trout, the disease is characterized by a subacute to acute course, with specific lesions of which the 'hallmarks' are panophthalmitis and meningitis; pathological changes in other organs are minor (Eldar & Ghittino 1999). Recently, in Israel, novel virulent strains of *S. iniae* were isolated from diseased rainbow trout that showed major pathological changes, suggesting a severe generalized disease. The new strains were characterized as Serotype II strains; they differed from 'classical' type I strains by serological, phenotypic and genetic criteria (Bachrach et al. 2001). Serotype II strains are able to enter phagocytes and multiply within them, causing them subsequently to undergo death through apoptotic processes (Zlotkin et al. 2003). *S. iniae* type II strains have also been detected in the USA, indicating the wide distribution of *S. iniae* variants (Barnes et al.

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2003). In the present work we assessed the pathological outcomes of infections of rainbow trout by *S. iniae* serotype II. Our results indicate that Serotype II strains produce an acute septic disease accompanied by multisystem organ involvement.

MATERIALS AND METHODS

Sampling procedure for pathology and bacteriological analysis. Specimens were collected from 2 trout farms located in northern Israel (Upper Galilee) which are supplied with water at a constant temperature of 16°C. At the time of collection both farms were experiencing heavy mortalities, and bacteriological examination (30 fish from each farm, 50 to 350 g each) revealed pure colonies of β -hemolytic Gram-positive cocci. Bacteriological identification (Eldar et al. 1995), authenticated by PCR analysis (Zlotkin et al. 1998), confirmed the presence of arginine dehydrolase-negative isolates of *Streptococcus iniae*. PCR analysis with the p14 primer did not produce the 750 bp band, indicating that all current isolates were of type II (Bachrach et al. 2001). Tissues (brain, heart, spleen, kidney, liver, intestine, gills and muscle) from 10 diseased rainbow trout were fixed in 10% neutral buffered formalin and stained with hematoxylin and eosin (HE). Gross lesions were recorded.

Distribution of bacteria in tissues. Bacterial colony-forming units (CFUs) were counted in the tissues of 20 rainbow trout (100 to 200 g each), infected with *Streptococcus iniae* type II and showing clinical signs of streptococcosis (darkening of the skin combined with multifocal ecchymoses, lethargy, loss of orientation and ocular pathologies). Organs (brain, heart, spleen, kidney, liver, intestine, gills and muscle) were weighed and blended over a 60-mesh grid. Tissue homogenate samples (1 g) were suspended in 10 ml phosphate-buffered saline (PBS) (15 mM Na₂HPO₄, 145 mM NaCl, pH 7.20) supplemented with 0.2% Triton X-100 (Sigma) and serially diluted. The bacterial CFU population was determined by plate counting.

The bacterial loads in tissues of rainbow trout infected by *Streptococcus iniae* type I were assessed with a 2-step cohabitation method. First, 10 naïve rainbow trout (50 to 80 g each) were infected by intraperitoneal inoculation of (7.5×10^5 CFU fish⁻¹) virulent *S. iniae* Dan-15 (type I strain), as described previously (Eldar et al. 1997a). Then, on Day 10 post-infection, 5 clinically diseased fish were removed and added to a group of 50 naïve rainbow trout (100 to 150 g each). The latter fish were infected by cohabitation, a process that closely resembled natural disease acquisition. Progress of the disease was monitored daily, and fish were considered to be infected by *S. iniae* when they

simultaneously exhibited 3 of the following 4 clinical signs: lethargy, black discoloration, loss of orientation and ocular pathologies. On Day 14 of cohabitation, 30 of the 50 fish exhibited clinical signs of the disease, and at this point 20 diseased fish were randomly chosen and sacrificed for bacteriological analysis. Bacterial counts were carried out as previously described. The re-isolation of *S. iniae* Dan-15 was confirmed by means of phenotypic and genetic tools (Bachrach et al. 2001).

Statistical analysis. The results are presented as the means \pm SDs of means of the data (CFU g⁻¹ tissue) obtained for each organ in triplicate. The data were transformed to decimal logarithmic values. To overcome the uncertainty whether these values were distributed normally, a non-parametric procedure was applied, i.e. analysis of variance for the logarithmic transformations of *Streptococcus iniae* CFU counts. Duncan's multiple range test was applied to the differences between the CFU counts obtained in different organs. The statistical analysis was performed with SAS software (Alice 1985).

RESULTS AND DISCUSSION

The major gross pathological signs that characterized *Streptococcus iniae* type II infection included lethargy, discoloration, gill pallor, bilateral corneal opacity and exophthalmia (present in 80 to 100% of the fish). Hemorrhage in the anterior chamber of the eye was observed in 30% of the diseased fish; inflammation of the anus in 10%; and external and internal hemorrhage in 40%. Skin (ecchymotic) hemorrhage was present mainly on the lateral sides and around the anal area; hemorrhage was more pronounced in the internal organs (particularly the spleen and the fat tissue around the intestine), and consisted of multifocal to coalescent petechiae. The most consistent visceral abnormality was splenomegaly, observed in 70% of the fish. Other internal lesions (hepatic lesions, congested kidneys, dilated intestine) were present, if at all, on less than 10% of the fish. These macroscopic findings point to an acute disease that differs from that observed in *S. iniae* type I infection. Histopathological findings, particularly those of the somatic muscles, confirmed this interpretation. While no injuries were observed in the skeletal muscles of fish infected by *S. iniae* type I, those of fish infected by *S. iniae* type II showed multifocal to diffuse areas of degeneration and necrosis, accompanied by heterophilic histiocytic infiltration. Necrotic fibers that had lost their striations appeared in small clumps; the cytoplasm of the individual cells was vacuolated and hypereosinophilic (Fig. 1). *S. iniae* type II bacterial counts in skeletal muscles (4.5×10^7 CFU g⁻¹) were unexpectedly high, being second only to those in

the central nervous system (CNS) (3×10^8 CFU g^{-1} ; $p < 0.05$).

As described previously for *Streptococcus iniae* type I infections (Eldar & Ghittino 1999), the present clinical findings of lethargy and loss of orientation correlated with intracranial edema (100% of the fish) resulting from CNS meningeal infection and damage. In addition, peripheral autonomic ganglia, especially those next to the olfactory tract, showed multifocal areas of subacute ganglionitis with histiocytic infiltration and foci of neuronal necrosis (hypereosinophilia, vacuolation of the cytoplasm and karyolysis). Although CNS lesions resemble those of type I strains, *S. iniae* type II infections were characterized by a significant augmentation in the count of viable bacteria in brain tissue (7×10^5 CFU g^{-1} in cases of infection by *S. iniae* type I versus 3×10^8 CFU g^{-1} in infections by *S. iniae* type II, $p < 0.005$; Table 1). Nonetheless, in both cases the highest bacterial load was found in the CNS, confirming *S. iniae* neurotropism.

Bacterial counts in infected tissues other than the CNS (Table 1) confirmed that *Streptococcus iniae* type II is responsible for an overwhelming septic disease characterized by large numbers of live bacteria, in accordance with similar models described in warm-blooded animals (Saetre et al. 2000, Yuste et al. 2002). The histopathological findings of the viscera were consistent with acute bacterial infection: there was mild to moderate centrilobular hepatitis with histiocytic infiltration in the liver; hyaline droplets were observed in the cytoplasm of tubular cells in the kidneys, and there

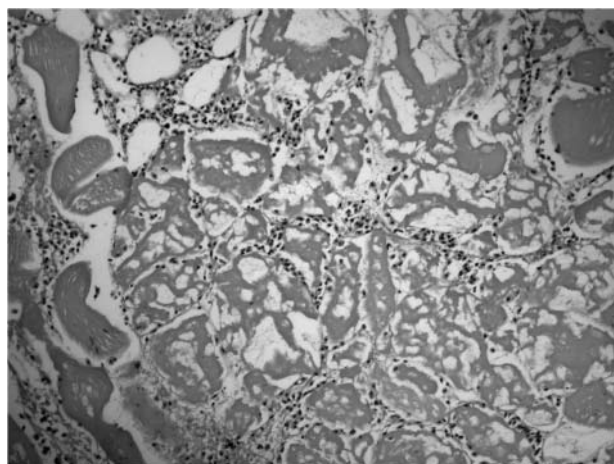


Fig. 1. *Oncorhynchus mykiss* infected by *Streptococcus iniae* type II. Striated muscle, temporal region: severe acute diffuse myositis and necrosis. HE $\times 400$

Table 1. *Streptococcus iniae* type I and type II CFU counts (decimal logarithmic transformations) in homogenates of infected tissues and the differences between types. Different letters indicate significant differences in CFU counts of each type between organs. For *S. iniae* type I: A>B,C $p < 0.01$; B>C $p < 0.01$. For *S. iniae* type II: a>b $p < 0.05$, a>c,d $p < 0.01$, b>d $p < 0.05$

Organ	<i>S. iniae</i> type I CFU g^{-1} tissue	<i>S. iniae</i> type II CFU g^{-1} tissue	F	Significance between <i>S. iniae</i> types
Brain	5.7 ± 0.6^A	8.5 ± 0.4^a	45.6	$p < 0.005$
Spleen	3.7 ± 0.6^B	7.4 ± 0.4^{bc}	81.0	$p < 0.001$
Kidney	5.4 ± 0.4^A	7.3 ± 0.2^{bc}	51.2	$p < 0.002$
Liver	3.5 ± 0.4^B	7.2 ± 0.3^{bc}	161.5	$p < 0.001$
Muscle	1.0 ± 0.2^C	7.5 ± 0.5^b	468.4	$p < 0.001$
Heart	3.4 ± 0.4^B	6.7 ± 0.6^d	68.9	$p < 0.002$

was mild lymphocyte depletion of interstitial tissue; and there was severe congestion in the spleen.

The augmented capability of *Streptococcus iniae* type II strains to survive in fish phagocytes and to induce their apoptotic death was recently suggested to be part of an advantageous mechanism that plays a critical role in the establishment of a generalized septic disease (Zlotkin et al. 2003). The data generated in the present study correlate with and extend previous results, and indicate that the augmented invasiveness of *S. iniae* type II also extends to parenchymal tissues, and so leads to a generalized septic disease. Although *S. iniae* type II infection results in a septic condition with multi-systemic organ involvement, the main clinical sign, CNS malfunction, resembles a similar condition in warm-blooded animals and humans, in which up to 70% of patients with sepsis develop CNS symptoms known as septic encephalopathy (Harris 1987, Pfister et al. 1993, Bone 1994, Cottagnoud et al. 2002). The question of whether a pathogen (*S. iniae* type II) that proliferates in internal tissues more successfully than another (*S. iniae* type I) is of higher virulence, or vice versa, is complex and beyond the scope of this report.

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