NOTE

Herpesvirus infection with severe lymphoid necrosis affecting a beaked whale stranded in the Canary Islands

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ABSTRACT: This report describes the pathologic findings in a single, adult female Cuvier’s beaked whale Ziphius cavirostris stranded in the Canary Islands. The study indicated that this whale died with a severe, systemic, herpesviral infection and clearly exhibited lesions different from those of the fat and gas embolic syndrome described in beaked whale mass strandings associated with sonar exposure. This is the first report of a cetacean alphaherpesvirus infection of the lymphoid system in a beaked whale.

KEY WORDS: Beaked whale · Herpes virus · Lymphoid system

INTRODUCTION

To date, 87 beaked whale stranding events were recorded along the coast of the Canary Islands, with a total of 119 individual animals of 6 species stranded: Cuvier’s beaked whale Ziphius cavirostris, Gervais beaked whale Mesoplodon europaeus, Blainville’s beaked whale Mesoplodon densirostris, True’s beaked whale Mesoplodon mirus, Sowerby’s beaked whale Mesoplodon bidens and the northern bottlenose whale Hyperodon ampullatus. Of these 119 animals, at least 86 were Cuvier’s beaked whales (Martín & Tejedor 2009). This species is sighted regularly at sea, especially off the east coast of Fuerteventura and Lanzarote (V. Martín pers. comm.) and in the waters around El Hierro island (N. Aguilar de Soto pers. comm.).

In the Canary Islands waters, 28 cetacean species have been identified of which 24 species have been found stranded. Etiopathologically, 62.32% of the cetaceans stranded in the Canary Islands subjected to pathological studies have been diagnosed as natural (i.e. non-anthropogenic) pathological entities that included infectious diseases, neonatal pathology, intraspecific interactions and typical mass strandings; and 33.33% of the cases have been diagnosed as anthropogenic entities including fishing interaction (by-catch), atypical mass-stranding linked to naval exercises, ship collisions, and other anthropogenic-related pathology (Arbelo 2007).

There are few reports of herpesviral infections in marine mammals. Most reports are morphological, where the diagnosis is based on gross pathology (Baker 1992), histopathology and immunohistochemistry (Kennedy et al. 1992) or transmission electron microscopy (TEM) (Martineau et al. 1988, Van Bressem et al. 1994). So far, no cetacean herpesviruses have been isolated with cell culture; thus, molecular diagnostic tools are essential to establish their taxonomic classification. Amplification of conserved DNA regions within the herpesvirus polymerase gene (VanDevanter et al. 1996) and terminase gene (Hargis et al. 1999) has been possible using universal PCRs. Further sequencing of the resultant amplicons has provided new reports of alphaherpesvirus in the bottlenose dolphin Tursiops...
truncatus (Blanchard et al. 2001, Manire et al. 2006, Smolarek-Benson et al. 2006), and gammaherpesvirus in bottlenose dolphin, Risso’s dolphin *Grampus griseus*, dwarf sperm whale *Kogia sima* and Blainville’s beaked whale *Mesoplodon densirostris* (Saliki et al. 2006, Smolarek-Benson et al. 2006). Using a universal nested PCR, Esperón et al. (2008) reported the first herpes simplex-like infection found in a stranded dolphin.

**MATERIALS AND METHODS**

On April 18, 2005, 1 female adult (555 cm long) Cuvier’s beaked whale was found dead on the southern coast of Fuerteventura Island (Faro la Entallada). Necropsy was performed approximately 48 to 72 h post-mortem (code 3) (Fernandez 2003, Fernandez et al. 2005).

Necropsy was carried out on the beach following the protocol used previously in other cases of stranded beaked whales (Fernandez et al. 2005). Lung, heart, muscle, thoracic and abdominal lymph nodes, liver, spleen, kidney, stomach, brain and intestines were sampled for routine light microscopic examination. To search for fat emboli, formalin-fixed lung and lymph node samples were post-fixed with osmium and embedded in paraffin following the procedure indicated in Fernandez et al. (2005). Lung, liver, spleen and muscle were sampled for bacteriologic aerobic culture.

For electron-microscopy (EM), formalin-fixed samples from spleen and lymph nodes were processed for routine transmission EM. A universal nested PCR (VanDevanter et al. 1996) that amplifies a conserved region within the polymerase gene of the *Herpesviridae* family was used in spleen, liver, muscle and lung. To detect possible carry over contamination, 2 blank controls (water, PCR-grade) were used for each reaction, one for DNA extraction and another for PCR reaction. A positive control of a herpes simplex-like sequence found in a stranded bottlenose dolphin (Esperón et al. 2008) was also used. The amplicon was directly sequenced by triplicate. Sequenced products were compared with sequences available in Genbank using the Blast search. A neighbor-joining phylogram was made using Mega 4.0 software (see Fig. 5).

Sequenced products were compared with sequences available in Genbank using the Blast search.

**RESULTS AND DISCUSSION**

The stranded whale exhibited a relatively good body condition. The main external findings consisted of post-mortem shark bites in the melon and in the dorsal and ventral abdominal regions. The stomach contained little ingesta indicating the animal had been anorectic for some time prior to death. The thorax was filled with a dark red fluid without blood clots. The lungs, trachea, and primary bronchi were congested and contained frothy fluid. The pulmonary lymph nodes were enlarged, and the spleen was smaller than normal for this species. Small, white to gray necrotic foci were observed in sections of the spleen. The liver was autolytic and congested. The kidneys were congested, and numerous adult parasites (*Crassicauda* spp.) were found within the ureters. *Cetobacterium ceti* and *Clostridium sordelli* spp. (both non-pathogenic bacteria) grew from routine aerobic bacterial cultures of brain, lung and spleen.

Microscopically, small, superficial dermal vessels were congested. Lung parenchyma was diffusely congested with areas of alveolar edema and atelectasia. No cardiac lesions were observed. The lymph nodes and spleen had severe, diffuse, coagulative necrosis and fibrinonecrotic vasculitis with prominent thrombi (Figs. 1 & 2). A large number of monocytic cells in spleen and lymph nodes had intranuclear inclusion bodies consistent with those of herpesviral or adenoviral infection (Fig. 3). The kidneys showed a chronic, parasitic, granulomatous nephritis, and in the liver, hepatocytes were atrophic and contained grey/brown, intracytoplasmatic lipofuscin pigment. Small fibrin thrombi were observed in the hepatic sinusoids. Submucosal and serosal vessels of the stomach were congested. Of the 4 samples processed, no fat emboli were seen. Although tissues were moderately autolytic, no lesions of gas embolism were observed. The histopathologic diagnosis was a systemic, widespread, necrotizing lymphadenitis and splenitis with acute necrotizing vasculitis and thrombosis.

Ultrastructurally, many, non-encapsulated, 100 nm herpes virions were observed in nuclei of monocytes.

Fig. 1. *Ziphius cavirostris*. Pulmonary lymph node. Vessels have vasculitis associated with congestion and thrombosis. Hematoxylin & eosin. Scale bar = 200 µm
from spleen and one abdominal lymph node (Fig. 4) and PCR testing detected Herpesvirus-specific bands in lung and spleen. Results of PCR showed a 198 bp band. Sequencing and further comparison with GenBank records showed a novel sequence highly related to cetacean alpha herpesvirus. The homology to their closest sequences in the Genbank (AF196646 and AY949832) was 84.2%. The sequence of the isolate obtained in this study was classified within the cetacean alphaherpesviruses group, and submitted to GenBank (Accession Number GU066291) (Fig. 5). It was concluded that this beaked whale had a severe, systemic herpesviral infection manifested most prominently in the lymphoid system.

Very few data are available about herpesviral systemic infections in cetaceans; one case of encephalitis in a harbour porpoise has been reported (Kennedy et al. 1992). Unfortunately, no brain was tested in the present study. In addition, 2 cases of generalized systemic herpesviral infections in bottlenose dolphins (Blanchard et al. 2001) have been described, and 2 novel sequences from herpesviral polymerase genes were detected. Main pathological findings were observed in the lymphoid system, heart, skin and adrenal glands. These sequences showed a close homology with those described in other bottlenose dolphins with skin diseases (Smolarek-Benson et al. 2006) and therefore they may be considered as the same viral species. In the present study, the PCR results showed a novel sequence classified within the cetacean alphaherpesviruses group.

Few cases involving stranded beaked whales have been published with detailed pathologic findings. This case report documents an infectious disease affecting these poorly known whale species, and records the first case of an alphaherpesvirus affecting the lymphoid system in beaked whales. The virus showed a marked tropism towards the lymphoid system, infecting mainly monocytes which would be the key cell in the pathogenesis of the necrosis observed in the lymph nodes and spleen, as well as the acute necrotizing vasculitis and thrombosis, despite the fact that no herpesviral inclusion bodies or virions were observed in vascular endothelial cells, either histologically or ultrastructurally.

This is the first report of a cetacean alphaherpesvirus infection causing a severe lymphoid necrosis in a stranded beaked whale (Ziphius cavirostris).
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LITERATURE CITED


Fig. 5. Neighbor-joining phylogram of 25 selected sequences from cetacean alpha- and gammaherpesvirus (HV) and other phylogenetically related alphaherpesviruses. Either the virus name or the host species is given alongside each GenBank Accession Number. The isolate from the present study (294P, GenBank Accession Number GU066291) showed greatest homology (84.2%) with GenBank sequences AF196646 and AY949832.