



FEATURE ARTICLE

Iron dynamics in a subtropical estuarine tidal marsh: effect of season and vegetation

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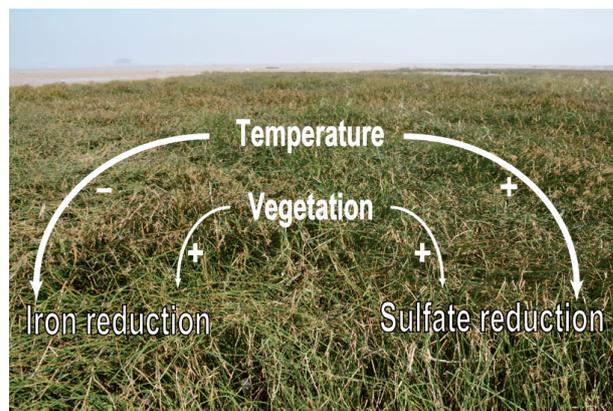
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ABSTRACT: To better understand how seasonal change and the presence of plants affect biogeochemical iron (Fe) cycling in tidal marsh sediments, we examined the seasonal dynamics of microbial Fe reduction (FeR) and related Fe speciation from 2012 to 2014 using *in situ* vegetated and unvegetated mesocosms created with macrobenthos-proof enclosures. The pools of Fe(III) oxides, Fe sulfides, pore-water Fe²⁺, and rates of microbial sulfate reduction (SR) peaked in summer (hot and wet), whereas rates of FeR and non-sulfidic Fe(II) levels peaked in winter (mild and dry). Sedge presence greatly increased the FeR rate, organic matter pools, and Fe(III) oxide levels, but decreased carbon:nitrogen ratios and sulfide abundances. FeR rates were mainly affected by the SR level, and temperature and plant primary productivity had comparable effects on SR rates. The dominant organic carbon mineralization pathway changed from SR in summer to FeR in winter in both vegetated and unvegetated mesocosms. FeR was more important in the vegetated mesocosms than in the unvegetated mesocosms, and seasonal change contributed more to the total variability of each anaerobic pathway than the presence of sedge. Our study reveals that temperature and plants both mediate variations in the organic carbon mineralization pathways of a subtropical estuarine tidal marsh.

KEY WORDS: Seasonality · Sedge · Microbial iron reduction · Microbial sulfate reduction · Organic carbon mineralization · Tidal marsh · Min River estuary



Both temperature and vegetation mediate variations in rates of microbial iron and sulfate reduction of a subtropical estuarine tidal *Cyperus malaccensis* marsh.

Photo: Min Luo

INTRODUCTION

Estuarine systems and sheltered coastal areas harbor extensive and highly productive (930–7600 g C m⁻² yr⁻¹) tidal marshes that receive substantial inputs of autochthonous and allochthonous organic carbon (C) (Schubauer & Hopkins 1984, Dame & Kenny 1986) and are foci of microbial activity, which result in high levels of organic C mineralization in the sediments (Mitsch & Gosselink 2000).

Microbial iron reduction (FeR) is a common pathway of organic C mineralization in many non-sulfidic

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intertidal systems (Kristensen et al. 2000, 2011, Kostka et al. 2002a, Kristensen & Alongi 2006, Hyun et al. 2007, Luo et al. 2016). The process is mediated by various Fe(III)-reducing microorganisms, such as *Geobacter*, *Bacillus*, *Shewanella*, and *Clostridium* (Lovley & Phillips 1986). Fe(III) can also be abiotically reduced by sulfides, which are primarily generated by microbial sulfate reduction (SR) and ultimately precipitated as Fe sulfides (e.g. FeS and FeS₂) (Berner 1970, Canfield 1989, Alongi 1995). FeR not only plays a key role in biogeochemical Fe, S, and C cycling, but also has an impact on greenhouse gas emissions and the adsorption and co-precipitation of trace metals that may be present in tidal marsh sediments (Huerta-Diaz & Morse 1992).

Previous studies on FeR have mainly investigated different habitats (Kristensen et al. 2000, 2011, Gribsholt & Kristensen 2002, Kostka et al. 2002a, Hyun et al. 2007), vegetation types (Kostka et al. 2002b), salinity or inundation gradients (Neubauer et al. 2005, Luo et al. 2016), or plant rhizospheres (Weiss et al. 2004, 2005). However, relatively few data are available regarding seasonal FeR dynamics in tidal marsh sediments, particularly for systems in the East Asian monsoon region, where hydrothermal conditions differ substantially from those in other coastal areas (Ding & Chan 2005).

In this study, a seasonal dataset of FeR and Fe speciation in the Shanyu tidal marsh of the Min River estuary, adjoining the East China Sea, was collated. Two sets of *in situ* mesocosms were established, with and without the indigenous sedge *Cyperus malaccensis*. Sediments for the mesocosms were collected from the same location to minimize inherent sediment heterogeneity (Holmer et al. 1999, Kristensen & Alongi 2006). The crab *Chiromantes dehaani* and mudskipper *Periophthalmus modestus* are ubiquitous to these tidal marshes, but are unevenly distributed. Because the bioturbation activities of these sediment-dwelling macrofauna also play important roles in sediment Fe and S geochemistry (Kristensen 2001, Gribsholt & Kristensen 2002, Kristensen & Alongi 2006, Ferreira et al. 2007), macrobenthos-proof enclosures were used to prevent the confounding effects of macrofauna. To accurately identify plant and seasonal effects, the mesocosms had almost identical hydrological conditions (i.e. tide and groundwater). Our strategy was to focus on creating conditions that eliminated spatial heterogeneity and macrobenthos disturbance, rather than completely reproducing natural conditions, and to facilitate quantitative comparisons of the relative effects of season and the presence of sedge plants on Fe dynamics.

MATERIALS AND METHODS

Site location and characteristics

The experiments were conducted in the eastern part of the Shanyu tidal marsh (ca. 31.20 km²) of the Min River estuary, on the coast of the East China Sea (Fig. 1); the characteristics of this site have been described in detail previously (Tong et al. 2010, Luo et al. 2014, 2016). The Min River estuary experiences a typical subtropical monsoon climate consisting of long, hot, and wet summers (June to September) and short, mild, and dry winters (January to February; Fig. 2). The average annual air temperature is approximately 21°C, with maximum temperatures in July (30 ± 1°C) and minimum temperatures in January (12 ± 2°C). Monsoon rains typically occur from March to June, although occasionally heavy precipitation events (typhoons) may occur as late as August. The average rainfall during the wet season (monthly average: 167 ± 66 mm) is much higher than during the dry season (monthly average: 48 ± 44 mm). The tidal marsh is flooded twice daily by semi-diurnal tides with a range of 2.5–6.0 m (Luo et al. 2014) and consequently is subjected to frequent periods of inundation (from 26% of the marsh in February to 31% in October). Inundation frequency is relatively stable over the course of a year ($p > 0.05$, coefficient of variation [CV] < 5%).

Experimental design

The mesocosms were established using specially designed polyethylene tanks (length: 60 cm; width: 40 cm; height: 36 cm) in which 2 columns of slots were drilled into the sides and bottoms to enable interactions with the surrounding sediments (Fig. 1c). Each tank was covered with 0.15 mm mesh filter fabric, and nylon-wire fencing was placed around the perimeter of each tank to prevent macrobenthos incursions. On 6 and 7 February 2012, 32 tanks were buried in the sediments with the nylon-wire fence ~20 cm above the surrounding ground level. The tanks were arranged in 2 lines 20–30 cm apart (Fig. 1d). Sediment was collected from a tidal creek bank ~20 m away and was then homogenized and passed through a 2 mm sieve to exclude any macrobenthos. Twenty of the tanks were uniformly planted with young *Cyperus malaccensis* shoots (density: 100 m⁻²; height: 5–10 cm) 1 wk following sediment transfer, and are hereafter termed 'vegetated mesocosms'; the other 12 tanks had no plants, and are

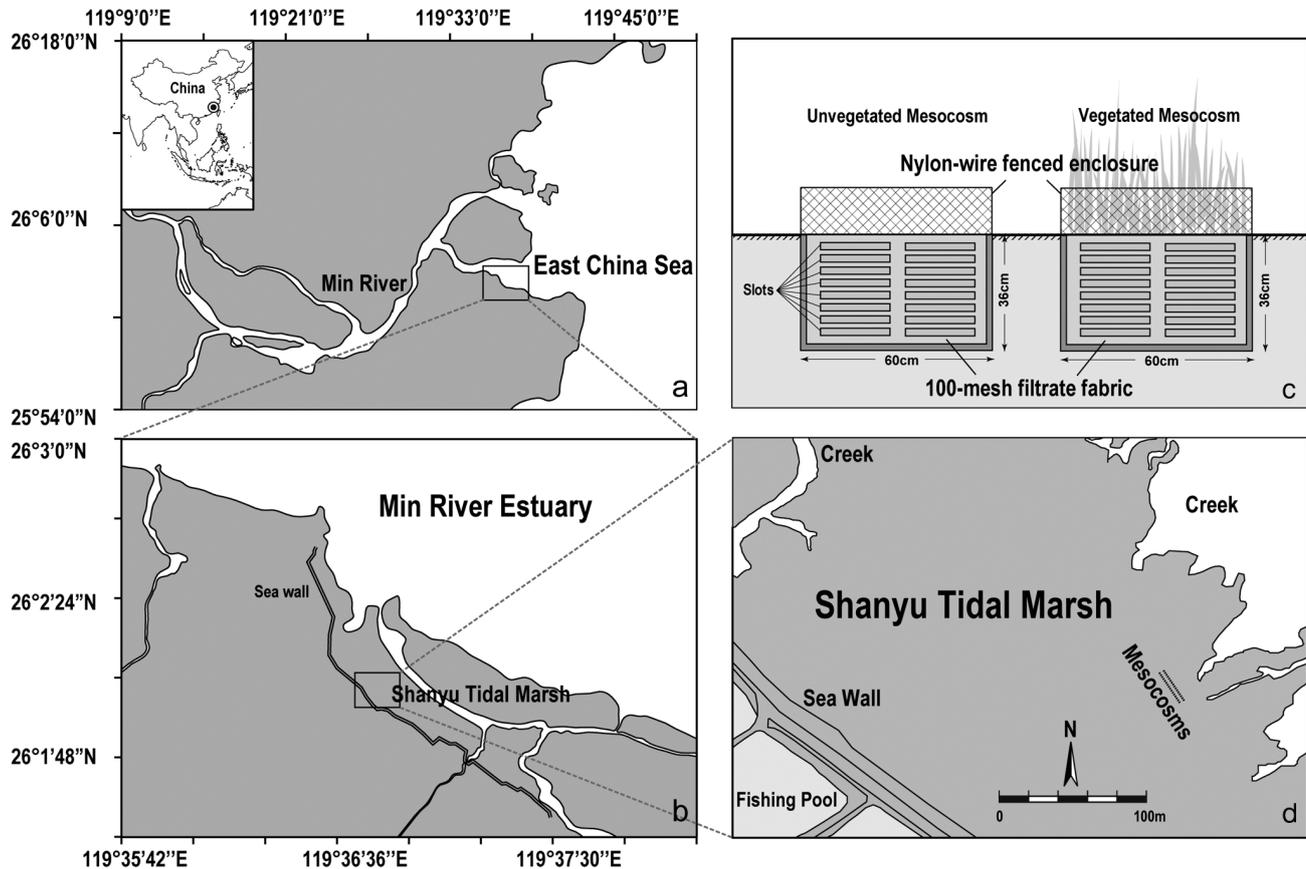


Fig. 1. (a,b,d) Location of the mesocosms. (c) Design of the vegetated and unvegetated mesocosms

hereafter termed ‘unvegetated mesocosms’. The mesocosms were then exposed to the *in situ* environment for 5 mo before sampling was initiated. The sampling periods were July and November 2012, February, April, June, and October 2013, and January 2014. Sediment excavated during sampling was returned to the mesocosms to avoid a collapse of the mesocosms.

Ecological measurements

Within three random plots (30 cm × 30 cm) among the vegetated mesocosms, all standing plant material (including live and dead roots) was harvested for aboveground biomass and all the sediments were washed in large 0.25 mm filter fabric bags to obtain belowground biomass during each sampling period. Biomasses were measured after the plant materials

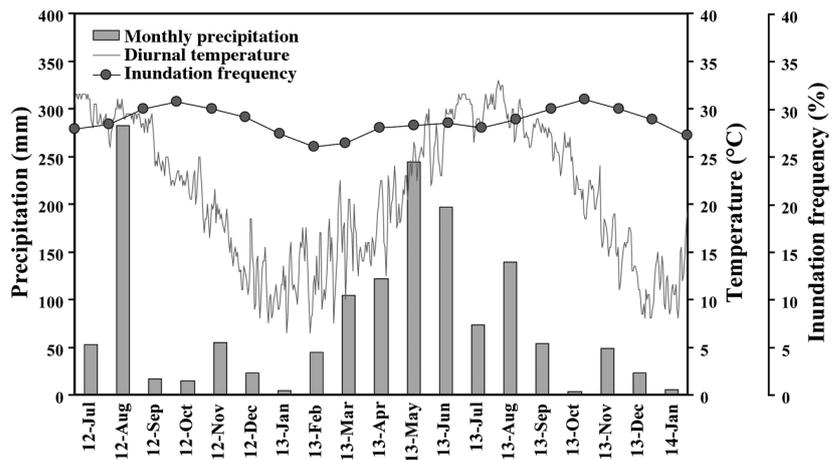


Fig. 2. Monthly precipitation, diurnal temperature, and monthly inundation frequency at the study site in the Min River estuary, East China Sea, during 2012–2014

had been oven-dried at 70°C for 48 h. Net primary productivity was calculated as the difference in total biomass (the sum of above- and belowground biomasses) between early in each sampling month and toward the end of that month (Wang et al. 2015).

Sample collection and preparation

In each sampling period, 5–7 sediment cores were taken from the 0–30 cm layer during low neap tide. This depth was chosen because the most dramatic impacts of seasonal change would be expected to occur in surface sediments (Böttcher et al. 2000, Kostka et al. 2002a). Cores were taken using polycarbonate corers (internal diameter: 5 cm; length: 50 cm) that were immediately capped at both ends following sampling and placed in vacuumized plastic bags. The samples were stored on ice and taken to the laboratory within 1 h of collection. In the laboratory, pretreatment of the sediment and porewater samples was conducted in a N₂-filled glove bag (AtmosBag, Sigma-Aldrich). Cores were extruded stepwise, and samples were obtained from the 0–2, 2–4, 4–6, 6–8, 8–10, 12–14, 20–22, and 28–30 cm layers. Sediment samples from each layer were homogenized and passed through a 2 mm sieve to remove roots. Porewater was extracted by centrifuging (4000 × *g* for 20 min) for pH, chloride (Cl⁻), and SO₄²⁻ analysis, and by *in situ* Rhizon samplers for dissolved organic carbon (DOC), Fe²⁺, and aqueous sulfide concentrations ($\Sigma\text{H}_2\text{S} = \text{H}_2\text{S}_{(\text{g})} + \text{HS}^- + \text{S}^{2-} + \text{S}_x^{2-}$; Bull & Taillefert 2001) analysis. The porewater samples were filtered through 0.45 μm syringe filters (Acrodisc, Pall) and drained into 10 ml glass vials; the filters used for Fe²⁺ and DOC analyses were prewashed with diluted hydrogen chloride (HCl). For $\Sigma\text{H}_2\text{S}$ determination, aliquots of the porewater samples were fixed in 20% zinc acetate, effervesced with N₂, and stored in bottles capped with Teflon-coated butyl rubber septa. The sediment and porewater samples were stored at 4°C until analysis (within 7 d).

Sediment and porewater geochemistry

Sediment total organic carbon (TOC) and total nitrogen (TN) levels were determined using a combustion analyzer (Vario EL III, Elementar), with an error margin lower than 0.3%. Carbonate was removed with 10% HCl prior to TOC determination. Solid-phase Fe species (including Fe(II), amorphous Fe(III), and crystalline Fe(III)) were extracted with 0.5 M deoxygenated HCl, 0.5 M deoxygenated HCl + hydroxylamine (0.1 g ml⁻¹), and citrate-bicarbonate-dithionite (0.35 M glacial acetic acid/0.2 M sodium citrate, buffered with 50 gl⁻¹ sodium dithionite) and analyzed by the sequential extraction method developed by Kostka & Luther (1994) and the 1,10-

phenanthroline method (APHA 2005). Solid-phase sulfur species (including acid-volatile sulfide [AVS] and chromium-reducible sulfide [CRS]) were determined by the sequential extraction method developed by Burton et al. (2008). Estimates of FeS and FeS₂ were based on the stoichiometric relationship between Fe and S in terms of differences in AVS and $\Sigma\text{H}_2\text{S}$ (Fe:S = 1:1 in AVS – $\Sigma\text{H}_2\text{S}$) and CRS (Fe:S = 1:2 in CRS) (Johnston et al. 2011). Non-sulfidic Fe(II) levels were calculated based on the difference in Fe(II) and Fe sulfides (non-sulfidic Fe(II) = Fe(II) – FeS – FeS₂). Further details can be found in previous studies (e.g. Luo et al. 2016).

Porewater pH was measured using a benchtop pH meter (Orion 3-Star Plus, Thermo Fisher Scientific), total alkalinity was determined using bromophenol blue method (detection limit = 1 μM; SD = 1%) as described by Sarazin et al. (1999), and DOC concentrations were analyzed using an Aurora 1030C TOC analyzer (OI Analytical). The 1,10-phenanthroline method (detection limit = 1 μM; SD = 1%; APHA 2005) was used to analyze porewater Fe²⁺ concentrations, and $\Sigma\text{H}_2\text{S}$ was determined by the methylene blue method (detection limit = 1 μM; SD = 1%; Cline 1969). Estimates of total carbon dioxide (TCO₂ = CO_{2(aq)} + HCO₃⁻ + CO₃²⁻) were then derived from the total alkalinity, porewater pH, and $\Sigma\text{H}_2\text{S}$ values (Stumm et al. 1996). Porewater SO₄²⁻ and Cl⁻ concentrations were determined by ion chromatography (Dionex ICS-2000). Changes in SO₄²⁻ concentrations (ΔSO_4^{2-}), which reflect the net SO₄²⁻ consumption, were calculated using Eq. (1) (Gribsholt & Kristensen 2002, Beck et al. 2008) assuming constant salinity over time:

$$\Delta\text{SO}_4^{2-} = (\text{SO}_4^{2-})_{\text{pw}} - (\text{Cl}^-)_{\text{pw}} \times \frac{(\text{SO}_4^{2-})_{\text{sw}}}{(\text{Cl}^-)_{\text{sw}}} \quad (1)$$

where (SO₄²⁻)_{pw} and (Cl⁻)_{pw} are the SO₄²⁻ and Cl⁻ concentrations in porewater, respectively, and (SO₄²⁻)_{sw} and (Cl⁻)_{sw} are the concentrations of SO₄²⁻ and Cl⁻ in seawater, respectively. The molar ratio of (SO₄²⁻)_{sw}/(Cl⁻)_{sw} is equal to 5.14 × 10⁻² in sea surface waters (Gribsholt & Kristensen 2002). A negative ΔSO_4^{2-} indicates the removal of SO₄²⁻, presumably by SR, whereas a positive ΔSO_4^{2-} indicates SR being offset by sulfide reoxidation (Thamdrup et al. 1994, Gribsholt & Kristensen 2002, Beck et al. 2008).

Rate measurements

Incubation experiments were conducted in a N₂-filled glove bag and are described in detail in a

previous study (Luo et al. 2016). Briefly, fresh sediment samples from each depth interval were thawed, pooled, mixed thoroughly, and then loaded into 50 ml glass jars with no head space. The jars were capped tightly with Teflon-coated butyl rubber septa and stored in the dark at field temperature for 72 h. At regular intervals (24 h), 5 to 7 jars were sacrificed for rate measurements. Anaerobic organic carbon mineralization rates (CMR) were determined based on the accumulation of porewater TCO_2 over time. The FeR rate (FeRR) was estimated from the generation of non-sulfidic Fe(II) and porewater Fe^{2+} over time, and the production of AVS and CRS over time was used to determine the SR rate (SRR). Our SRR estimates should be accepted with some caution, given that the method may discount sulfide reoxidation resulting in underestimates of the actual SRR (for more details and further discussion, see Luo et al. 2016). The contributions of FeR (FeR%) and SR (SR%) are presented in C units using the theoretical stoichiometry of C:Fe = 1:4 and S:C = 2:1 (Kristensen et al. 2000, 2011, Kostka et al. 2002a, Hyun et al. 2007, 2009).

Data analysis

The data were tested for homogeneity of variance and normality, then log transformed if necessary before further statistical analysis. Differences in ecological and geochemical properties among sediment depths, seasons, and mesocosms (vegetated versus unvegetated) were tested by 1-way ANOVAs. The effects of plant presence, season, and their interaction on FeR, SR, CMR, FeR%, and SR% were examined by 2-way ANOVAs. Individual means (within seasons or mesocosms) were compared by least significant difference post-hoc tests. DOC, ΔSO_4^{2-} , $\Sigma\text{H}_2\text{S}$, amorphous Fe(III), crystalline Fe(III), FeS, and FeS_2 levels in the vegetated and unvegetated mesocosms were compared using paired-sample *t*-tests. Data are presented as mean \pm SD, unless indicated otherwise. Bivariate correlations among ecological, geochemical, and environmental properties, Fe species, and anaerobic metabolism were tested using the Pearson correlation coefficient (*r*). The relative importance of temperature, precipitation, and primary productivity on SR was determined by stepwise multiple linear regression. The CV is the ratio of the SD to the mean. Significance was set at $\alpha = 0.05$ for all tests, which were performed in PASW Statistics 22.0.

RESULTS

Ecological properties

Cyperus malaccensis grew vigorously in the vegetated mesocosms 5 mo after planting, and the plant biomasses were comparable with the results of previous studies (Wang et al. 2015). Season significantly affected both biomass and primary productivity in the vegetated mesocosms (Tables 1 & 2). Above-ground biomass was significantly higher in the summer ($1099 \pm 117 \text{ g m}^{-2}$) than in the other seasons, whereas belowground biomass was significantly higher in the autumn ($2743 \pm 319 \text{ g m}^{-2}$) than in the other seasons. Primary productivity differed significantly among the seasons (Table 2), increasing from spring ($4.93 \pm 1.33 \text{ g m}^{-2} \text{ d}^{-1}$) to summer ($8.98 \pm 2.17 \text{ g m}^{-2} \text{ d}^{-1}$), and decreasing from autumn ($-0.41 \pm 1.00 \text{ g m}^{-2} \text{ d}^{-1}$) to winter ($-4.67 \pm 1.93 \text{ g m}^{-2} \text{ d}^{-1}$; Table 1).

Sediment geochemistry

Both TOC and TN levels ($\mu\text{mol g}^{-1}$ dry weight [dw]) were higher in the vegetated mesocosms (TOC: 2.19–3.03; TN: 0.19–0.27) than in the unvegetated mesocosms (TOC: 1.45–1.81; TN: 0.15–0.18; Tables 1 & 2). Higher C:N ratios were observed in the vegetated mesocosms (range: 11.2–12.2) than in the unvegetated mesocosms (range: 8.6–10.4; Table 2). Season had a significant effect on TOC and TN pools in the vegetated mesocosms, with higher values being recorded during winter and summer than during spring and autumn. In contrast, it had no effect on TOC and TN pools in the unvegetated mesocosms (Table 2). There were no significant differences in the C:N ratio among the sampling periods in both types of mesocosms (Table 2).

Porewater geochemistry

Porewater pH ranged from 6.4 to 7.1 and from 6.7 to 7.3 in the vegetated and unvegetated mesocosms, respectively (Fig. 3a), but the difference between mesocosms was not significant, nor were significant differences observed among seasons or sediment layers (Table 2).

Total alkalinity had a broader range and was significantly higher in the vegetated mesocosms (range: 6.9–21.6 mM) than in the unvegetated mesocosms (range: 4.7–15.0 mM; Fig. 3b). Total alkalinity was significantly higher during winter than during the

Table 1. Ecological and geochemical properties of surface sediments (0–30 cm) in vegetated and unvegetated mesocosms at the study site in the Min River estuary, southeastern China, during 2012–2014. Values are mean \pm SD. TOC: total organic carbon; TN: total nitrogen; dw: dry weight; na: not applicable

	Aboveground biomass (g m ⁻²)	Belowground biomass (g m ⁻²)	Primary productivity (g m ⁻² d ⁻¹)	TOC (μ mol g ⁻¹ dw)	TN (μ mol g ⁻¹ dw)	C:N ratio
July 2012						
Vegetated	1062 \pm 71	2311 \pm 136	8.64 \pm 1.22	2.75 \pm 0.09	0.23 \pm 0.01	12.1 \pm 0.9
Unvegetated	na	na	na	1.58 \pm 0.07	0.16 \pm 0.01	10.0 \pm 0.4
November 2012						
Vegetated	589 \pm 26	2640 \pm 211	-0.80 \pm 0.55	2.35 \pm 0.16	0.21 \pm 0.01	11.2 \pm 0.5
Unvegetated	na	na	na	1.50 \pm 0.10	0.16 \pm 0.01	9.5 \pm 0.8
February 2013						
Vegetated	385 \pm 42	2071 \pm 143	-5.32 \pm 1.54	3.03 \pm 0.09	0.27 \pm 0.01	11.3 \pm 0.4
Unvegetated	na	na	na	1.55 \pm 0.13	0.15 \pm 0.01	10.4 \pm 0.5
April 2013						
Vegetated	465 \pm 63	1845 \pm 105	4.93 \pm 0.77	2.19 \pm 0.09	0.19 \pm 0.01	11.5 \pm 0.1
Unvegetated	na	na	na	1.81 \pm 0.05	0.18 \pm 0.01	10.1 \pm 0.2
June 2013						
Vegetated	1135 \pm 71	2502 \pm 94	9.32 \pm 1.52	2.84 \pm 0.13	0.25 \pm 0.02	11.7 \pm 1.3
Unvegetated	na	na	na	1.45 \pm 0.21	0.17 \pm 0.02	8.6 \pm 0.5
October 2013						
Vegetated	756 \pm 70	2845 \pm 173	-0.02 \pm 0.61	2.42 \pm 0.39	0.20 \pm 0.03	12.2 \pm 0.3
Unvegetated	na	na	na	1.64 \pm 0.14	0.16 \pm 0.02	10.4 \pm 0.4
January 2014						
Vegetated	321 \pm 35	2354 \pm 172	-4.02 \pm 0.55	2.89 \pm 0.27	0.26 \pm 0.05	11.6 \pm 1.2
Unvegetated	na	na	na	1.55 \pm 0.13	0.15 \pm 0.01	10.4 \pm 0.1

Table 2. Summary of 1-way ANOVA and paired-sample *t*-tests of the effects of plant, season, and depth on ecological properties, sediment and porewater geochemistry, and iron (Fe) speciation in the Min River estuary, East China Sea, during 2012–2014. TOC: total organic carbon; TN: total nitrogen; DOC: dissolved organic carbon; V: data in the vegetated mesocosms; U: data in the unvegetated mesocosms; -: no data

	Plant		Season		Depth	
	<i>F</i> or <i>T</i>	<i>p</i>	<i>F</i> or <i>T</i>	<i>p</i>	<i>F</i> or <i>T</i>	<i>p</i>
Ecological and geochemical properties						
Aboveground biomass	-	-	$F_{3,17} = 54.230$	<0.001	-	-
Belowground biomass	-	-	$F_{3,17} = 8.478$	<0.01	-	-
Primary productivity	-	-	$F_{3,17} = 69.230$	<0.001	-	-
TOC ^a	$F_{1,40} = 106.998$	<0.001	V: $F_{3,17} = 5.635$ U: $F_{3,17} = 5.635$	<0.01 >0.05	-	-
TN ^a	$F_{1,40} = 41.505$	<0.001	V: $F_{3,17} = 1.388$ U: $F_{3,17} = 1.581$	<0.05 >0.05	-	-
C:N ratio	$F_{1,40} = 27.888$	<0.001	$F_{3,38} = 0.101$	>0.05	-	-
Porewater geochemistry						
pH	$F_{1,292} = 1.069$	>0.05	$F_{3,290} = 0.892$	>0.05	$F_{6,287} = 1.687$	>0.05
Total alkalinity ^a	$F_{1,292} = 73.501$	<0.001	$F_{3,290} = 43.707$	<0.001	$F_{6,287} = 14.652$	<0.001
DOC	$T_{146} = 15.685$	<0.001	$F_{3,290} = 43.753$	<0.001	$F_{6,287} = 1.307$	>0.05
SO ₄ ²⁻ ^a	$F_{1,292} = 15.786$	<0.001	$F_{3,290} = 17.880$	<0.001	$F_{6,287} = 1.013$	>0.05
Δ SO ₄ ²⁻ ^a	$T_{146} = 3.653$	<0.001	$F_{3,290} = 1.799$	>0.05	$F_{6,287} = 1.894$	<0.001
H ₂ S ^a	$T_{146} = 3.853$	<0.001	$F_{3,290} = 2.062$	>0.05	$F_{6,287} = 52.249$	<0.001
Fe speciation						
Porewater Fe ²⁺ ^a	$F_{3,290} = 23.451$	<0.001	$F_{3,290} = 23.451$	<0.001	V: $F_{6,140} = 19.683$ U: $F_{6,140} = 19.784$	<0.001 >0.05
Non-sulfidic Fe(II) ^a	$F_{1,292} = 1.151$	>0.05	$F_{3,290} = 9.391$	<0.001	$F_{6,287} = 176.915$	<0.001
Amorphous Fe(III)	$T_{146} = 35.290$	<0.001	$F_{3,290} = 7.716$	<0.001	$F_{6,287} = 9.697$	<0.001
Crystalline Fe(III)	$T_{146} = 11.364$	<0.001	$F_{3,290} = 0.994$	>0.05	$F_{6,287} = 22.404$	<0.001
FeS ^a	$F_{1,292} = 78.336$	<0.001	$F_{3,290} = 10.068$	<0.001	$F_{6,287} = 21.416$	<0.001
FeS ₂ ^a	$F_{1,292} = 52.823$	<0.001	$F_{3,290} = 92.157$	<0.001	$F_{6,287} = 7.114$	<0.001

^aData were tested after log₁₀ transformations

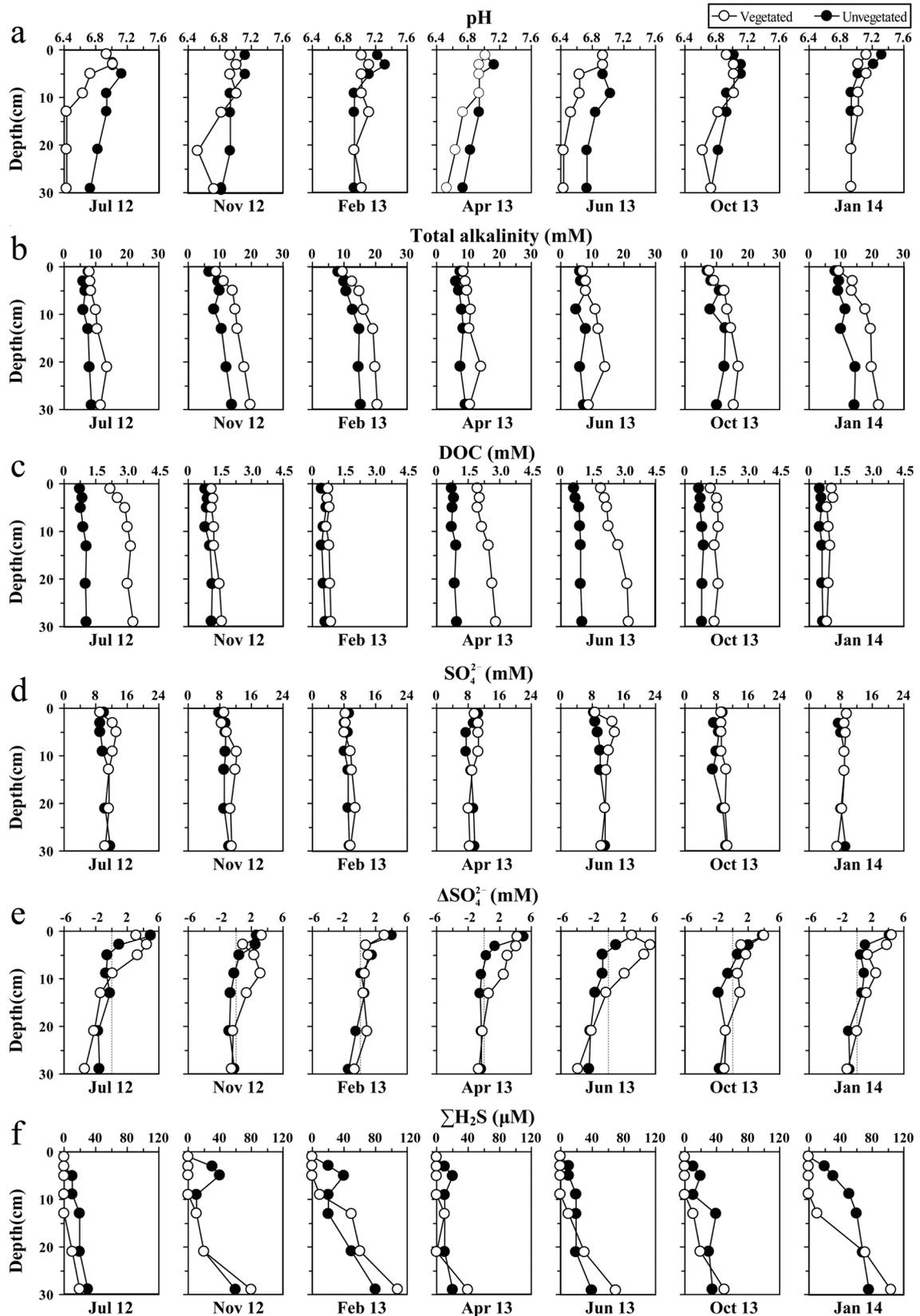


Fig. 3. Depth-specific porewater chemistry in vegetated and unvegetated mesocosms in the tidal marsh sediments of the Min River estuary, East China Sea, during 2012–2014. (a) pH, (b) total alkalinity, (c) dissolved organic carbon (DOC), (d) SO_4^{2-} , (e) ΔSO_4^{2-} , and (f) $\Sigma\text{H}_2\text{S}$

other seasons and generally increased with depth in both types of mesocosms (Table 2).

The vegetated mesocosms typically had a larger DOC pool (range: 0.5–3.3 mM) than the unvegetated mesocosms (range: 0.4–1.0 mM) in all sediment layers in every season (Fig. 3c). Seasonal changes in DOC levels occurred in the 2 types of mesocosms, both of which showed maximum levels in summer and minimum levels in winter (Table 2). Sediment depth did not appear to affect DOC (Table 2).

Porewater SO_4^{2-} concentrations were significantly higher in the vegetated mesocosms (range: 7.3–13.6 mM) than in the unvegetated mesocosms (range: 7.2–11.6 mM) (Fig. 3d) and were higher in the summer than in the other seasons. Sediment depth did not appear to affect porewater SO_4^{2-} (Table 2). A positive ΔSO_4^{2-} was observed in the 0–10 cm and 0–4 cm sediment layers of the vegetated and unvegetated mesocosms, respectively, whereas a negative ΔSO_4^{2-} was observed in the lower layers (Fig. 3e). Seasonal differences in ΔSO_4^{2-} were not observed in any mesocosm (Table 2).

Porewater $\Sigma\text{H}_2\text{S}$ was not detected in the upper 0–2 cm and 0–13 cm layers of the unvegetated and vegetated mesocosms, respectively, but generally increased with depth below these layers. Porewater $\Sigma\text{H}_2\text{S}$ was significantly higher in the unvegetated mesocosms (range: 0–81 μM) than in the vegetated mesocosms (range: 0–110 μM , Fig. 3f), but did not differ among seasons (Table 2).

Fe speciation

Porewater Fe^{2+} was lowest during the winter. In the vegetated mesocosms, porewater Fe^{2+} gradually increased with depth from 0.0 to 2.4 mM (Fig. 4a), but did not do so in the unvegetated mesocosms, where it remained at very low levels (range: 0 to 0.3 mM; Table 2).

The vegetated and unvegetated mesocosms did not differ in non-sulfidic Fe(II) levels (ranges: 12–216 $\mu\text{mol g}^{-1}$ dw and 6–173 $\mu\text{mol g}^{-1}$ dw, respectively; Fig. 4b), which significantly increased with depth in both mesocosms throughout the year (Table 2). The levels peaked during the winter in both mesocosms, and were significantly different from those during the other seasons (Table 2).

Crystalline Fe(III) was more abundant than amorphous Fe(III) throughout the year (Fig. 4c,d). Amorphous Fe(III) peaked during the summer, but season did not affect crystalline Fe(III) (Table 2). There were larger pools of both amorphous and crystalline Fe(III)

oxide in the vegetated mesocosms than in the unvegetated mesocosms (Table 2).

Regardless of mesocosm type, FeS_2 concentrations (0.0–18.0 $\mu\text{mol g}^{-1}$ dw) were lower than those of FeS (range: 0.4–29.8 $\mu\text{mol g}^{-1}$ dw; Fig. 4e,f). Both FeS and FeS_2 concentrations significantly changed with season, peaking during the summer and falling to their lowest levels during the winter (Table 2). The vegetated mesocosms had significantly smaller Fe sulfide pools than the unvegetated mesocosms (Table 2).

Anaerobic metabolism

CMR ranged from 68 to 90 $\text{mmol m}^{-2} \text{d}^{-1}$ in the vegetated mesocosms and from 59 to 67 $\text{mmol m}^{-2} \text{d}^{-1}$ in the unvegetated mesocosms; FeRR ranged from 95 to 170 $\text{mmol m}^{-2} \text{d}^{-1}$ in the vegetated mesocosms and from 70 to 113 $\text{mmol m}^{-2} \text{d}^{-1}$ in the unvegetated mesocosms; SRR ranged from 11 to 22 $\text{mmol m}^{-2} \text{d}^{-1}$ in the vegetated mesocosms and from 13 to 20 $\text{mmol m}^{-2} \text{d}^{-1}$ in the unvegetated mesocosms (Fig. 5). The vegetated mesocosms had significantly higher CMR, FeRR, and SRR than the unvegetated mesocosms (Table 3). In both types of mesocosm, SRR peaked during the summer and was lowest in the winter, whereas FeRR exhibited the opposite trend. Season had a significant effect on CMR patterns in the vegetated mesocosm, but had no similar effect in the unvegetated mesocosm.

SRR in both types of mesocosm were correlated with temperature ($r_{\text{vegetated}} = 0.79$, $n = 21$; $r_{\text{unvegetated}} = 0.77$, $n = 21$), and SRR in the vegetated mesocosms were also correlated with the primary productivity of *C. malaccensis* ($r_{\text{vegetated}} = 0.88$, $n = 21$). A stepwise multivariable linear regression analysis revealed that primary production accounted for comparable variation in SRR to temperature according to the standardized regression coefficients (Table 4). FeRR was negatively correlated with SRR in both types of mesocosm ($r_{\text{vegetated}} = -0.80$, $n = 21$; $r_{\text{unvegetated}} = -0.81$, $n = 21$). There were no significant relationships between the SRR and FeRR and precipitation and inundation frequency.

Contribution of anaerobic metabolism

FeR accounted for 26 to 63% and from 27 to 47% of total CMR in the vegetated and unvegetated mesocosms, respectively, whereas the contribution of SR ranged from 32 to 53% and from 44 to 60% in the vegetated and unvegetated mesocosms, respectively

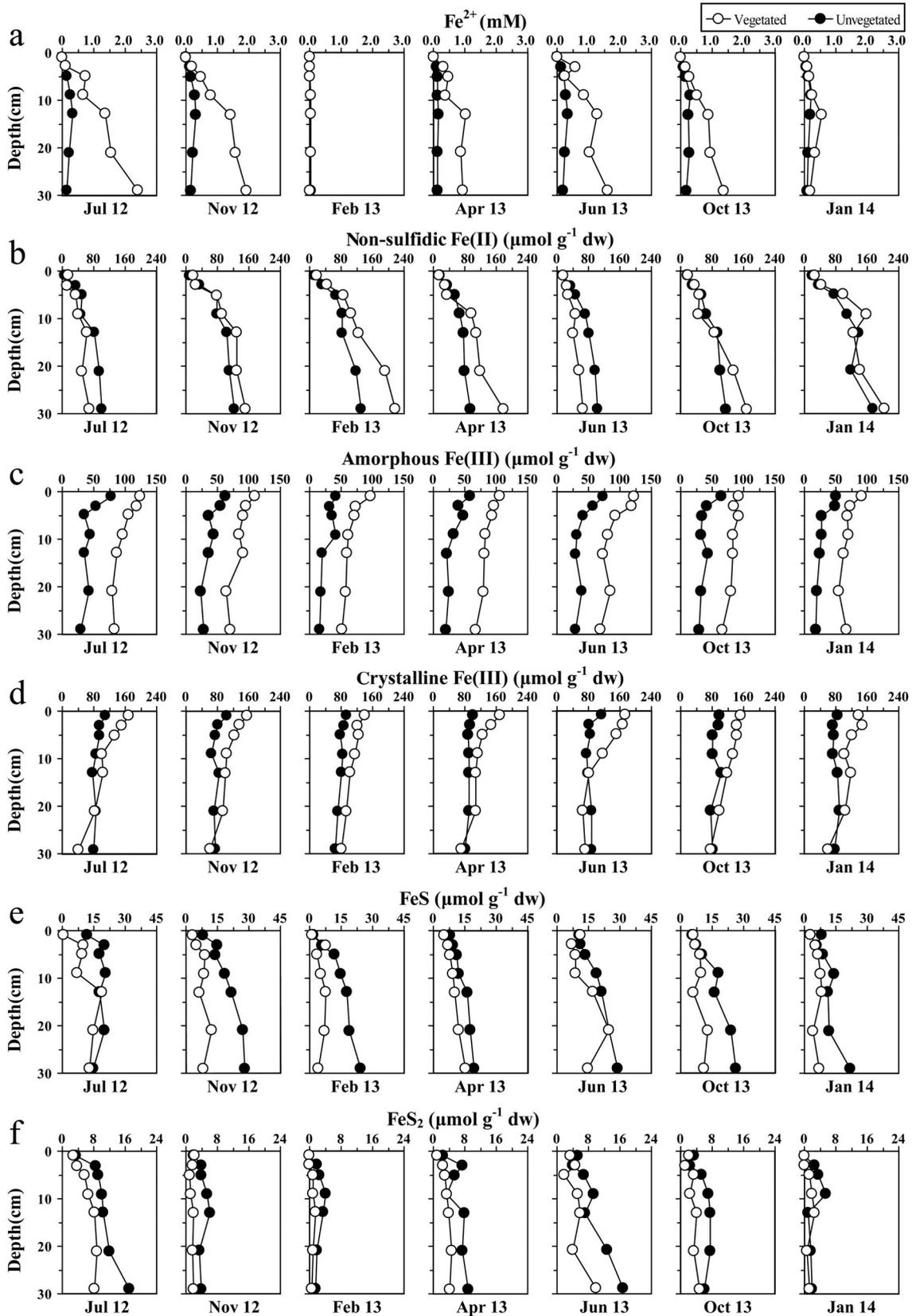


Fig. 4. Depth-specific iron (Fe) speciation in vegetated and unvegetated mesocosms in the tidal marsh sediments of the Min River estuary, East China Sea, during 2012–2014. (a) Porewater Fe^{2+} , (b) non-sulfidic Fe(II) , (c) amorphous Fe(III) , (d) crystalline Fe(III) , (e) FeS , and (f) FeS_2

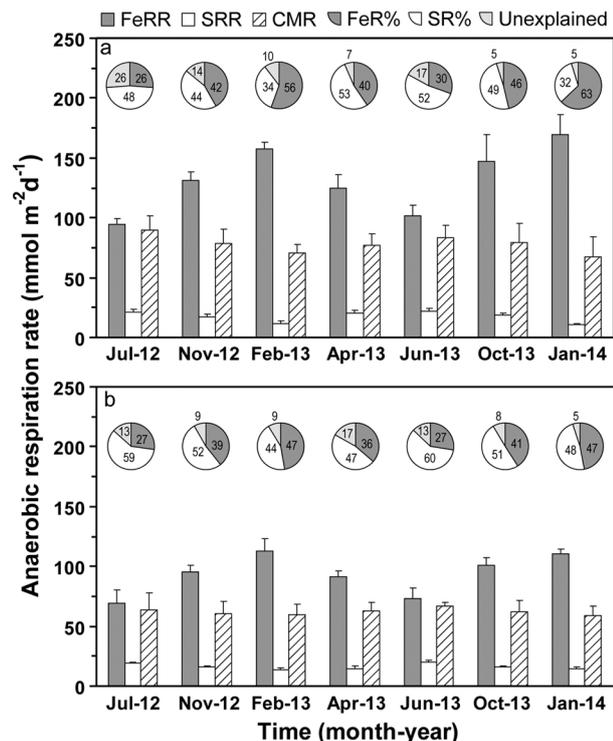


Fig. 5. Seasonal changes in depth-integrated rates of microbial iron reduction (FeRR), microbial sulfate reduction (SRR), and carbon mineralization (CMR) (bar charts); and FeR%, SR%, and discrepancy (pie charts) in (a) vegetated and (b) unvegetated mesocosms in tidal marsh sediments of the Min River estuary, East China Sea, during 2012–2014. Unexplained proportion denotes a difference between 100% and FeR% + SR%. Error bars indicate SE

(Fig. 5). Both season and vegetation significantly affected FeR% and SR%, and accounted for 86% and 11% of FeR% variability, respectively, whereas they accounted for 55% and 31% of SR% variability, respectively (Table 3).

DISCUSSION

The mesocosm approach has previously been used for studying SR in saltmarsh and mangrove sediments (Gribsholt & Kristensen 2002, Nielsen et al. 2003), as well as FeR in mangrove sediments (Kristensen & Alongi 2006). However, to the best of our knowledge, this is the first time *in situ* mesocosms have been used to examine the role that sedge plants play in FeR in tidal marsh sediments.

Seasonal change in anaerobic metabolism

SR tracked seasonal temperature variations in all of the mesocosms. Similar seasonal trends in SR have been observed in other coastal systems, such as saltmarshes (Abdollahi & Nedwell 1979, King 1988, Moeslund et al. 1994, Kostka et al. 2002a,b), mangrove forests (Holmer et al. 1999), and marine fish farms (Holmer & Kristensen 1996). Warm seasons tend to support a greater SO_4^{2-} -reducing activity and

Table 3. Summary of a 2-way ANOVA of the effects of plant, season, and their interaction on inter-depth rates of microbial iron reduction rate (FeRR), microbial sulfate reduction rate (SRR), carbon mineralization (CMR), the contribution of iron reduction (FeR%), and the contribution of sulfate reduction (SR%) in the Min River estuary, East China Sea, during 2012–2014. Results in bold are statically significant ($p < 0.05$)

Source	df	FeRR		SRR		CMR		FeR%		SR%	
		F	p	F	p	F	p	F	p	F	p
Plant	1	90.281	<0.001	12.994	0.001	25.143	<0.001	5.795	0.022	8.024	0.008
Season	3	36.999	<0.001	39.590	<0.001	2.780	0.056	47.167	<0.001	14.086	<0.001
Plant × Season	3	2.967	0.046	6.913	0.001	0.718	0.548	1.602	0.207	3.446	0.027
Total		130.247		59.497		28.641		54.564		25.556	
r^2		0.869		0.813		0.513		0.819		0.662	

Table 4. Multiple regression parameters explaining the variance associated with microbial sulfate reduction rate (SRR). Adjusted $r^2 = 0.735$. Multiple regression $F = 38.866$; $p < 0.001$. Models were derived using the stepwise selection method and included seasonal (temperature and precipitation) and plant (above- and belowground biomasses, primary productivity, total organic carbon, total organic nitrogen, C:N ratios, and integrated-depth dissolved organic carbon) components

Response variable	Independent variable	Unstandardized regression coefficient (mean ± SE)	Standardized regression coefficient	T	p
SRR	(Constant)	11.232 ± 1.102		10.194	<0.001
	Temperature	0.230 ± 0.055	0.412	4.190	<0.001
	Primary productivity	0.455 ± 0.099	0.472	4.607	<0.001

higher levels of bioavailable organic substrates than cold seasons (Howarth 1993, Holmer & Kristensen 1996, Koretsky et al. 2003, Beck et al. 2008, Al-Raei et al. 2009). The large quantities of Fe sulfides and negative sediment ΔSO_4^{2-} observed in the present study also indicate that SR increased during the spring and summer (Figs. 3f & 4e,f). In contrast, more positive sediment ΔSO_4^{2-} and smaller accumulations of Fe sulfides were observed during the autumn and winter (Fig. 3e), indicating that sulfide oxidation is stronger than SR in cold months (Howarth & Giblin 1983, Hines et al. 1989, Hyun et al. 2007, Kostka et al. 2002a).

The inhibitory effect of increasing temperature on FeRR was clear in both types of mesocosm (Fig. 5), which was in contrast to simultaneous increases in Fe(III) oxide levels and organic substrate availability during warm months (Table 1, Figs. 3c & 4c,d). This also disagrees with previous studies reporting increased growth of isolated Fe(III)-reducing microorganisms with increasing temperature (King & Garey 1999, Meier et al. 2005). Furthermore, a positive correlation exists between electron acceptors or organic substrates and FeRR in freshwater systems (Roden & Wetzel 1996, Neubauer et al. 2005) and rice wetlands (Yao & Conrad 2000). However, a culture experiment on saltmarsh sediments collected from Sapelo Island, USA, provided results similar to ours (Koretsky et al. 2003). The same study showed that Fe(III)-reducing microorganisms were adversely affected by the addition of $\Sigma\text{H}_2\text{S}$ (Koretsky et al. 2003). This is in accordance with the negative relationship observed between FeRR and SRR in both types of mesocosm in the present study. Importantly, both field and laboratory data support the sulfide-mediated limitation of FeR (Koretsky et al. 2003, 2005, Meier et al. 2005). How sulfides suppress FeR, even with abundant electron acceptors or organic substrates, is unclear. However, previous studies have found abundant SO_4^{2-} -reducing microorganisms in the tidal wetlands of the Min River estuary e.g. *Desulfovibrio*, *Desulfobacterium*, and *Desulfobulbus* (Tong et al. 2015), which can reduce Fe(III) and SO_4^{2-} simultaneously (Lovley et al. 1993). Therefore, the inhibition of FeR by sulfides may also be caused by a shift in the microbial metabolic pathway (Weston et al. 2006), which means that more facultative microorganisms tend to use SO_4^{2-} rather than Fe(III) in the presence of sulfides.

Previous studies have shown that the annual precipitation pattern over the course of a year (i.e. wet and dry seasons) also has a large impact on Fe and S biogeochemistry (Hines et al. 1989, Holmer et al.

1999, Kristensen et al. 2011, Nóbrega et al. 2013), given that the hydrology and hydroperiod of tidal marsh sediments are linked to oxygen (O_2) availability and redox conditions (Alongi 1995, Kostka & Luther 1995, Holmer et al. 1999, Kostka et al. 2002a, Nóbrega et al. 2013). During the wet season, rainfall could dilute SO_4^{2-} and thus suppress the activity of SO_4^{2-} -reducing microorganisms (Hines et al. 1989, Holmer et al. 1999, Alongi et al. 2001, Nóbrega et al. 2013). However, our results suggest that SRR and FeRR were similar in early spring ($15.2 \pm 2.0^\circ\text{C}$; precipitation: 122 ± 21 mm) and mid-autumn ($17.5 \pm 3.3^\circ\text{C}$; precipitation: 55 ± 13 mm), when temperatures were more or less similar but moisture regimes differed considerably (Fig. 5). These results, combined with the insignificant correlation observed between precipitation and both SRR and FeRR, suggest that these 2 anaerobic respiration rates in the tidal marsh sediments of the Min River estuary are less sensitive to changes in precipitation than to changes in temperature. This was most likely because our study site in the low-tide zone was inundated with seawater on a semi-daily basis, which may have offset any potential precipitation effects. Kostka & Luther (1995) reported a similar pattern in the Great Marsh, Delaware, USA.

Influence of plants on sediment geochemistry

SRR in the vegetated mesocosms were correlated with both temperature and primary productivity. Temperature regulates SRR not only directly, but also indirectly via sedge plants, given that temperature is a key factor in controlling the physiological development of plants (Alongi et al. 2001, Koretsky et al. 2003). SRR were low in cold months, possibly because the *Cyperus malaccensis* plants had reached their frutescent and withering phases (September to February). During this period, plants generally allocate more organic matter to reproductive tissues or reduce exudations to withstand the cold, and consequently less plant-derived organic matter enters the sediment (Hines et al. 1999, Tobias & Neubauer 2009; Table 1, Fig. 3c).

DOC is the main source of labile organic substrate for benthic microorganisms in tidal marshes (Hines et al. 1989, 1991, 1999). In addition to microbial hydrolysis and fermentation of organic matter, root exudates and detrital inputs also contribute to the DOC pools in the vegetated mesocosms (Holmer et al. 1999, Armstrong et al. 2000, Kristensen et al. 2000, 2011). This is probably the reason why the

DOC pools were larger and exhibited greater fluctuations (due to constantly changing root activity) in the vegetated than in the unvegetated mesocosms (Fig. 3c). However, plant-derived DOC in the vegetated mesocosms may also have contained structural macromolecules (e.g. lignin), which would inhibit microbial respiration (Kristensen et al. 2008). Consequently, a certain fraction of low-quality DOC in the vegetated mesocosms may explain the lack of a clear relationship between respiration rates and the DOC pool.

Fe(III)-reducing microorganisms in the vegetated mesocosms were provided with higher Fe(III) oxide concentrations than those in the unvegetated mesocosms (Fig. 4c,d). The initial Fe(III) pools were identical in the 2 kinds of mesocosm, but roots induce a progressive and long-term increase in Fe(III) oxides in rhizospheres due to radial O₂ loss (Crowder & St-Cyr 1991, Frenzel et al. 1999, Neubauer et al. 2008), which creates oxic and suboxic mosaics (Gribsholt & Kristensen 2002, Holmer et al. 2002, Kostka et al. 2002a,b). The accumulated reduced Fe(II) is then reoxidized into a rusty-colored Fe plaque, which is composed of large amounts of amorphous Fe(III) and smaller amounts of crystalline Fe(III) (Weiss et al. 2004, 2005, Neubauer et al. 2008). Although Fe(III)-reducing microorganisms can reduce a variety of Fe(III) oxide minerals, they prefer amorphous Fe(III) to crystalline Fe(III) because of the larger surface area of the former (Bonneville et al. 2009, Li et al. 2012). The importance of the mineralogical composition of Fe(III) oxides for FeR has been demonstrated in both field and laboratory settings (Weiss et al. 2004, Luo et al. 2015). In our experiment, 42 ± 4% (n = 147) of reactive Fe(III) (the sum of amorphous and crystalline Fe(III)) in the vegetated mesocosms was amorphous Fe(III), but this proportion was only 30 ± 3% (n = 147) in the unvegetated mesocosms. A relatively high abundance of amorphous Fe(III) may also account for the higher FeRR in the vegetated than in the unvegetated mesocosms.

Porewater Fe²⁺ levels were higher in the vegetated than in the unvegetated mesocosms during the sedge-growing seasons (Fig. 4a). One possible explanation for this is that Fe sulfides are oxidized by the O₂ that is transported to the roots (Gribsholt & Kristensen 2002, Holmer et al. 2002), which leads to an accumulation of porewater Fe²⁺ and an acidification of the sediments (Berner 1970, Luther & Church 1988, Otero & Macías 2002). This is in accordance with the fact that sulfide concentrations and the pH were lower in the vegetated than in the unvegetated mesocosms (Figs. 3a & 4e,f). Another explanation is

that higher DOC concentrations in the vegetated mesocosms resulted in higher levels of organic substrate adsorption onto amorphous Fe(III) under reducing conditions, which generated dissolved organic Fe(II) complexes (Jones et al. 2011). Dissolved organic Fe(II) complexes may be rapidly reoxidized by O₂ or Fe(III) minerals and yield dissolved organic Fe(III) (Luther et al. 1992). Dissolved organic Fe(III) is a preferred electron acceptor for Fe(III)-reducing microorganisms, and it accelerated Fe cycling in the vegetated mesocosms because dissolved organic Fe(III) complexes are generally reduced more rapidly than solid-phase Fe(III) minerals (Beckler et al. 2015). Although dissolved organic Fe(III) was not measured in our study, relatively high concentrations of dissolved organic Fe(III) have been found in saltmarsh creek sediments in previous studies (Koretsky et al. 2008, Jones et al. 2011, Beckler et al. 2015).

Total anaerobic metabolism

Our results suggest that the dominant organic C mineralization pathway changed from SR in summer to FeR in winter in both vegetated and unvegetated mesocosms (Fig. 5). Similar seasonal trends in the partitioning between SR and FeR have been reported in the brackish marshes of the Patuxent River, Maryland, USA, and saltmarshes on Sapelo Island, Georgia, USA (Giblin & Howarth 1984, Kostka et al. 2002a, Neubauer et al. 2005). However, the opposite trend was observed in a fresh tidal marsh on the Patuxent River, Maryland, USA (Neubauer et al. 2005), and in a freshwater wetland in Talladega, Alabama, USA (Roden & Wetzel 1996). These discrepancies may have been caused by low levels of SO₄²⁻ limiting SR in tidal freshwater marshes, and an insufficient generation of sulfides to inhibit FeR.

Our results also suggest that the presence of plants increases the competitiveness of FeR% and lowers SR%, which is in accordance with that found in other studies, including mangrove mesocosms in North Queensland, Australia (Kristensen & Alongi 2006), tidal marshes on Sapelo Island, USA (Kostka et al. 2002a) and Skidaway Island, USA (Hyun et al. 2007), Han River estuary, Korea (Hyun et al. 2009), Patuxent River, USA (Neubauer et al. 2005), and mangrove forests on Phuket Island, Thailand (Kristensen et al. 2000).

Both FeRR and SRR may be underestimated in sealed laboratory incubations, because the rapid reoxidation of Fe(II) and sulfides by sedge root activity, advective groundwater processes, and tidal

pumping are not taken into account (Kristensen et al. 2000, 2011, Tallifert et al. 2007, Beckler et al. 2015). In addition, dissolved organic substrate-mediated Fe(II)–Fe(III) cycling cannot occur in closed containers because roots are excluded (Beckler et al. 2015).

When combined, FeR% and SR% on average accounted for $88 \pm 5\%$ of total anaerobic organic C mineralization in the 2 types of mesocosm (Fig. 5). The unexplained proportion may have been caused by underestimated SR% in both types of mesocosm, but particularly for the vegetated mesocosms due to rapid sulfide reoxidation (Luo et al. 2016). The unexplained proportion might also have been caused by CO₂ formation via fermentation (Jørgensen 2000, Hyun et al. 2009) or by other terminal electron-accepting processes, such as denitrification and manganese (Mn) reduction. However, the contributions of NO₃⁻ and Mn(IV) to organic C mineralization may have been small, because their concentrations are much lower than concentrations of SO₄²⁻ and Fe(III) in sediments of the Min River estuary tidal marsh (Luo et al. 2014).

CONCLUSIONS

Comparative data collected from *in situ* vegetated and unvegetated mesocosms from 2012 to 2014 clearly demonstrate that seasonal change and the presence of plants both have an effect on Fe cycling and the organic C mineralization pathway in tidal marsh sediments of the Min River estuary. A shift in the dominant organic matter mineralization pathway from SR in summer to FeR in winter occurred in both the vegetated and unvegetated mesocosms. The presence of sedge plants reduced the relative importance of SR in favor of FeR as the primary organic C mineralization pathway. When directly compared, seasonal change clearly had a more pronounced effect on the organic C mineralization pathway than the presence of plants. SR influenced FeR in both vegetated and unvegetated mesocosms, while SR variability was mainly caused by seasonal cycles of temperature and primary productivity. In addition to temperature, precipitation, and plants, other factors not examined here, including light intensity, tidal flushing, salinity, bioturbation, freshwater inputs, artificial interference, typhoons, alien species invasions, and sea-level rises, may also affect the dynamics of plant-mediated processes and oxidative events in tidal marsh sediments. Therefore, redox-mediated Fe dynamics should be further investigated in future studies.

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