

Spatial and temporal distribution of tropical phytoplankton species and biomass in the Gulf of Carpentaria, Australia

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ABSTRACT: The biomass and species composition of tropical phytoplankton in Albatross Bay, Gulf of Carpentaria, northern Australia, were examined monthly for 6 yr (1986 to 1992). Chlorophyll *a* (chl *a*) concentrations were highest (2 to 5.7 $\mu\text{g l}^{-1}$) in the wet season at inshore sites, usually coinciding with low salinities (30 to 33 ppt) and high temperatures (29 to 32°C). At the offshore sites chl *a* concentrations were lower (0.2 to 2 $\mu\text{g l}^{-1}$) and did not vary seasonally. Nitrate and phosphate concentrations were generally low (0 to 3.68 μM and 0.09 to 3 μM for nitrate and phosphate respectively), whereas silicate was present in concentrations in the range 0.19 to 13 μM . The phytoplankton community was dominated by diatoms, particularly at the inshore sites, as determined by a combination of microscopic and high-performance liquid chromatography (HPLC) pigment analyses. At the offshore sites the proportion of green flagellates increased. The cyanobacterium genus *Trichodesmium* and the diatom genera *Chaetoceros*, *Rhizosolenia*, *Bacteriastrum* and *Thalassionema* dominated the phytoplankton caught in 37 μm mesh nets; however, in contrast to many other coastal areas studied worldwide there was no distinct species succession of the diatoms and only *Trichodesmium* showed seasonal changes in abundance. This reflects a stable phytoplankton community in waters without pulses of physical and chemical disturbances. These results are discussed in the context of the commercial prawn fishery in the Gulf of Carpentaria and the possible effect of phytoplankton on prawn larval growth and survival.

KEY WORDS: Tropical phytoplankton · Biomass · Community · Seasonal changes · Gulf of Carpentaria

INTRODUCTION

The Gulf of Carpentaria is a large, shallow (<65 m depth) embayment in tropical northern Australia. Albatross Bay, a shallow (<20 m) bay in the northeast of the Gulf, has a distinct wet season in summer (October to March), with freshwater addition from flood plains and estuaries, and little or no rain for the rest of the year. Light northeast to northwest monsoons blow in the wet season, southeasterly winds in the dry season. Albatross Bay is within the coastal boundary layer, i.e. a zone of water with a particular physical, biological or chemical property that is different from the offshore water (Wolanski & Ridd 1990). It may trap freshwater runoff, nutrients and suspended sediments and inhibit mixing between the coastal and offshore waters.

The biomass and productivity of phytoplankton in the Gulf are comparable with those of other tropical

continental shelf regions (Hallegraeff & Jeffrey 1984, Rothlisberg et al. 1994). In short-term studies, chlorophyll *a* concentrations ranged from 0.78 to 2.34 $\mu\text{g l}^{-1}$ throughout the Gulf, but were generally higher and more uniform throughout the water column in shallow waters and stratified near the bottom in deeper waters (Hallegraeff & Jeffrey 1984). Primary productivity in coastal waters is considerably higher (1430 $\text{mg C m}^{-2} \text{d}^{-1}$) than offshore (660 $\text{mg C m}^{-2} \text{d}^{-1}$) (Rothlisberg et al. 1994). The phytoplankton community is dominated by the cyanobacterium genus *Trichodesmium* and large diatoms such as the genera *Rhizosolenia*, *Chaetoceros*, *Bacteriastrum* and *Thalassionema* (Hallegraeff & Jeffrey 1984).

In a 6 yr study of the recruitment dynamics of the prawns *Penaeus merguensis* and *P. semisulcatus* (Rothlisberg et al. 1988), the larval, juvenile and adult stages in the estuaries and waters of Albatross Bay

were examined. Our study focused on the seasonal, spatial and interannual changes in the phytoplankton community, which is both a direct food source for larvae and a link in the food web for juvenile and adult prawns. The biomass and species composition of the phytoplankton community were examined in conjunction with physical and chemical factors to find out which factors govern phytoplankton distribution and abundance in Albatross Bay.

METHODS

Sample collection. Temperature and salinity measurements were made with a submersible data logger each month between August 1986 and April 1992 at 4 sites in and near Albatross Bay (Fig. 1a). Measurements were not taken in some months and a few sites were not sampled during inclement weather. Wind speed and rainfall data from Weipa were obtained from the Australian Bureau of Meteorology. Wind speed was measured at 09:00 and 15:00 h daily, and rainfall was measured as a daily total. The data were pooled to give monthly totals.

In order to determine the phytoplankton pigment composition, water samples of 2 to 5 l were collected concurrently with the temperature and salinity measurements at the 4 sites over 6 yr (Fig. 1a) with a 5 l Niskin bottle, at mid-depth, i.e. 5 m at Site a, 6 m at

Site b, 18 m at Site c, 22 m at Site d. In the last 2 yr, bottom samples were also taken. Samples for pigment analysis were filtered on deck through GF/F glass fibre filters, which were frozen immediately. In the last year (August 1991 to April 1992), the supernatant was also collected and frozen for phosphate, nitrate+nitrite and silicate analyses.

Collections of larger phytoplankton species were made at the 4 sites (Fig. 1a) with surface-to-bottom drops of a 0.6 m diameter, 37 μ m mesh free-fall net (Heron 1982). They were immediately preserved with 2% formaldehyde buffered with hexamine.

In addition to the monthly sampling, 2 cruises were made to study vertical and horizontal changes in phytoplankton pigments in more detail, and size fractionation of the phytoplankton community. In February 1990, 21 sites in Albatross Bay were sampled at 2 m depth intervals for phytoplankton pigment analysis (Fig. 1b). Salinity and temperature profiles were recorded. Water samples of 2 to 5 l were collected, the phytoplankton filtered onto GF/F glass fibre filters and the filters immediately frozen for pigment analysis. In February 1991, water samples were filtered by size-fractionating through 0.6, 2 and 10 μ m pore size polycarbonate filters and the filters were immediately frozen.

Nutrient analyses. Frozen filtered samples were collected at 4 sites on 9 occasions in the final year of sampling, 6 of these during the wet season; they were

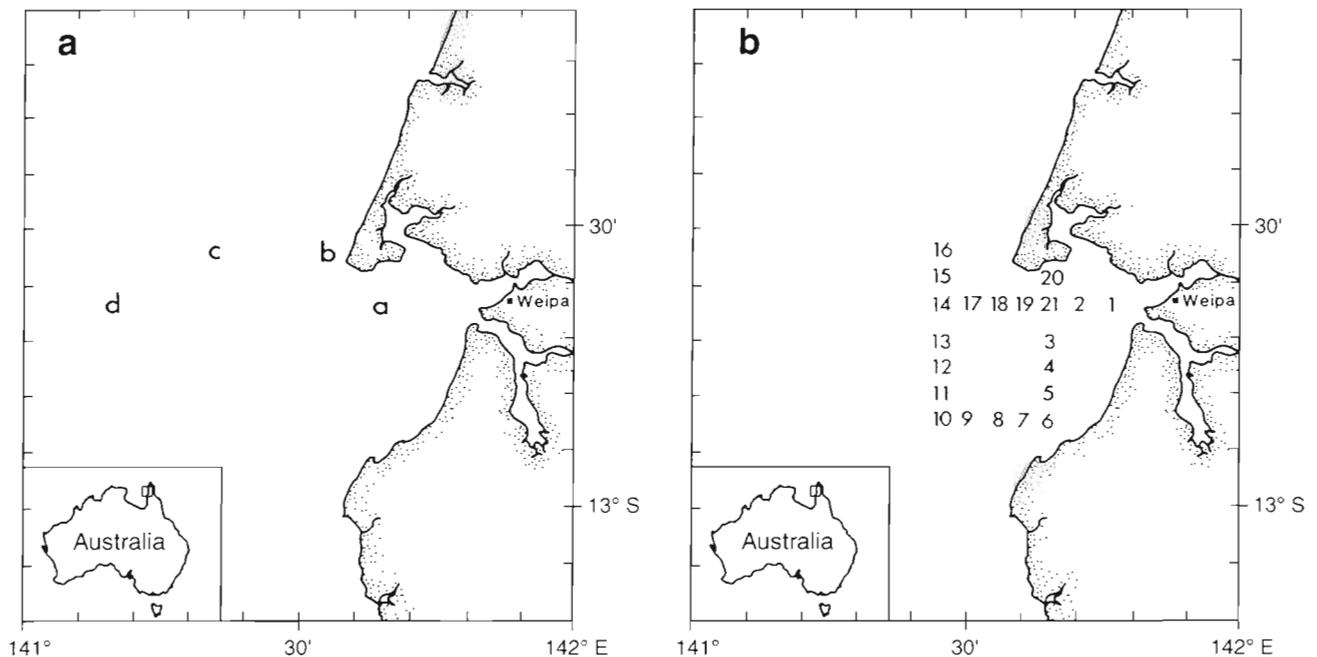


Fig. 1 Sites in Albatross Bay, Gulf of Carpentaria, Australia, sampled (a) monthly from August 1986 to April 1992, (b) in February 1990

thawed in the laboratory and analysed by the Strickland & Parsons (1972) method for nitrate+nitrite, phosphate and silicate.

Identification of 37 μm mesh net phytoplankton. In the laboratory, samples of phytoplankton caught by net were shaken and 2 subsamples removed for examination under a Wild Leitz M20 microscope under phase-contrast conditions at 200 and 400 \times magnification. Depending on the concentration of the sample, about 10 scans were done across two 22 \times 22 mm cover slips. Genera were identified from Allen & Cupp (1935) and Taylor (1976). The samples were quantified by grouping a list of potential genera (Hallegraeff & Jeffrey 1984) into 3 categories: dominant (genus present in every field of view at 200 \times magnification); common (genera observed more than 5 times in the total number of scans, but less frequently than dominant genera) and rare (genus observed no more than 5 times in the total number of scans).

The dominant, common and rare categories were classified numerically as 3, 2 and 1 respectively. The mean and standard deviation for each genus were calculated from 242 samples, the result of near-monthly sampling at 4 sites from May 1986 to April 1992. Mean abundances of >2 were classified as dominant, >1 to 2 were classified as common, and 1 or less as rare. Of 70 genera, 59 (83%) had a mean abundance of less than 1.0, and 27 (38%) had a mean abundance of less than 0.1. Only the 44 genera with a mean greater than 0.1 were used for further analysis (Field et al. 1982).

All statistical analyses were done with SAS software (Release 6.07). The Wilk's Lambda statistics were used to approximate *F* statistics for multivariate hypothesis testing (Seber 1984). The effect of site, season and year on the abundance of net phytoplankton was examined. The Bray-Curtis dissimilarity indices were used for cluster analysis and multidimensional scaling (MDS) of the net phytoplankton genera (Clarke 1993). Based on the dissimilarity indices, the genera were classified into groups. The dimension coefficients for 4 sites obtained from the MDS were then plotted to see how similar they were in terms of species abundance. The configuration of the MDS (the coordinates of each genera) was then plotted to determine the similarity of genera in terms of abundance.

Pigment analyses. The samples were analysed in the laboratory within 1 mo of sampling. Each sample was sonicated in 100% methanol on ice with a Branson ultrasonic probe for 1 min in the dark, and then

filtered through a 0.2 μm pore size filter to remove particulates. The extract was collected and injected into a high-performance liquid chromatography (HPLC) system. A methanol : water : ethyl acetate gradient solvent system and RP-C18 column were used. For details of this technique see Burford & Pollard (1994). The spectra and HPLC retention times of the various pigments were compared with those of pigments previously isolated from standard algal cultures (Burford et al. 1994).

RESULTS

Phytoplankton seasonality and hydrological environment

During the 6 yr of monthly sampling in Albatross Bay, water temperatures at the innermost site (Site a) ranged from 25 to 27°C in the dry season (April to September), to 30 to 32°C in the wet season (October to March) (Fig. 2a). The temperature range was the same at Site b, and at Sites c and d it was 25 to 31°C overall;

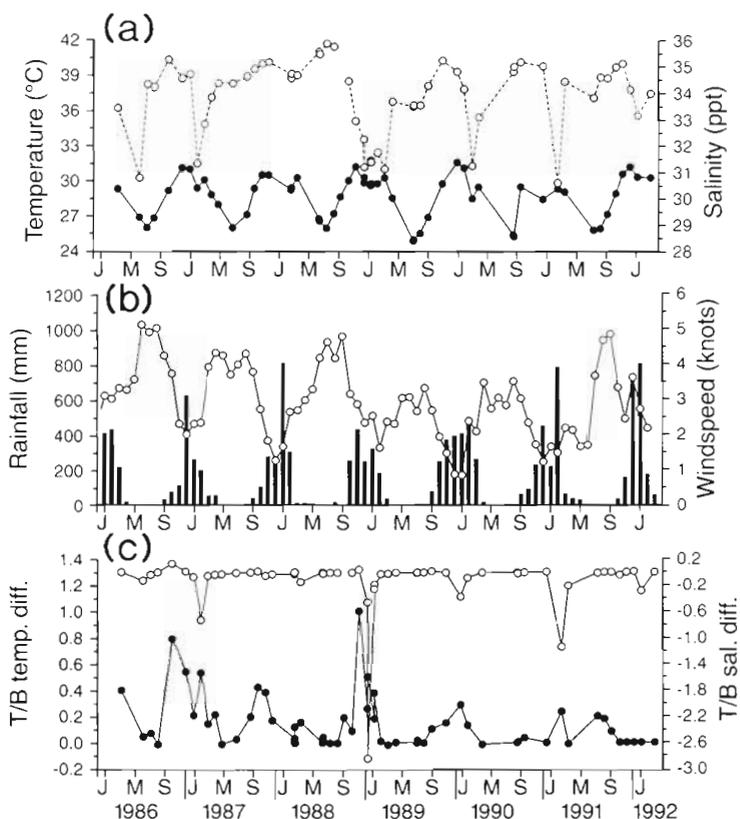


Fig. 2. Albatross Bay, Site a. (a) Mean temperatures (●) and mean salinity (○) in the water column. (b) Total monthly rainfall (bars) and windspeed (○) at Weipa, Gulf of Carpentaria. (c) Top/bottom temperature (●) and salinity (○) differences from April 1986 to April 1992. J: January; M: May; S: September

however, seasonal differences were not as great. The rainfall, most of which fell in the wet season, increased output from the rivers emptying into Albatross Bay and lowered salinities from 35 to 36 ppt in the dry season to 31 to 32 ppt in the wet season (Fig. 2a, b). The salinity range was the same at Site b, and at Sites c and d it was 32 to 36 ppt overall.

The stratification of the water column was generally weak, as indicated by the small differences in surface-to-bottom salinity and temperature (Fig. 2c); only in January 1989 and March 1991 was there a significant surface-to-bottom difference in salinity. Surface-to-bottom temperature differences did not necessarily coincide with surface-to-bottom salinity differences and reflect warming of surface waters in the summer wet season during periods of low wind mixing (Fig. 2b). The monthly wind speed at Weipa was higher in the dry season than in the wet season (Fig. 2b).

Chlorophyll *a* (chl *a*) concentrations at Site a ranged from 0.1 to 2.5 $\mu\text{g l}^{-1}$ over the 6 yr of sampling (Fig. 3). At Site b, they ranged from 0.5 to 5.7 $\mu\text{g l}^{-1}$, decreasing at the offshore sites to usually less than 1 $\mu\text{g l}^{-1}$ at Site d. Concentrations were highest in the wet season at the 2 innermost sites (a and b) but seasonality was not apparent at the 2 offshore sites (c and d) during the 6 yr.

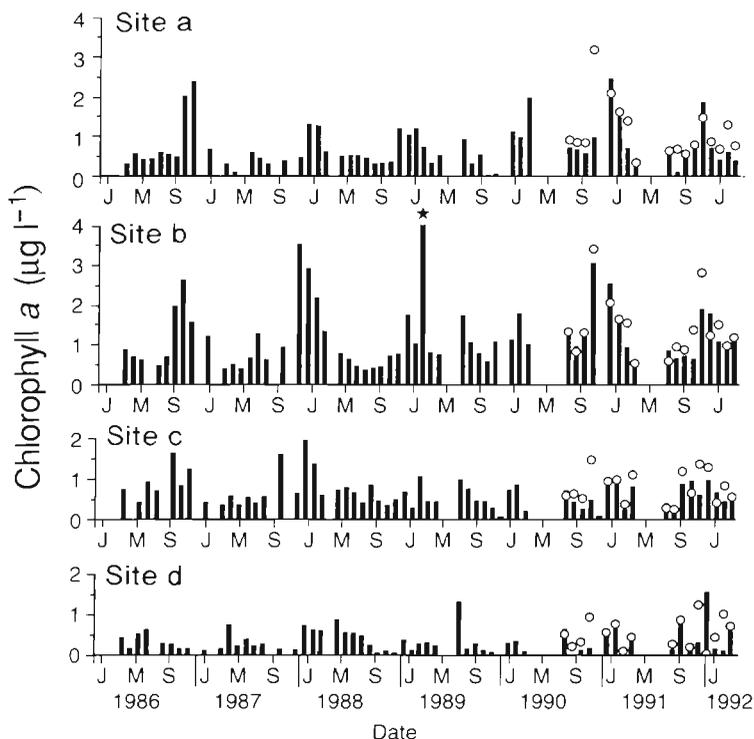


Fig. 3. Chl *a* concentrations at 4 sites in Albatross Bay, sampled monthly from April 1986 to April 1992. Bars are mid-depth concentrations; (o) bottom concentrations; (*) a chl *a* concentration of 5.7 $\mu\text{g l}^{-1}$. J: January; M: May; S: September. Months in which sampling did not occur have no bars

Table 1 Correlation between chl *a* and the environmental factors temperature, salinity and rainfall. * $p < 0.05$

	Chl <i>a</i>			
	Site a	Site b	Site c	Site d
Temperature	0.179	0.256	0.087	0.149
Salinity	0.068	0.223	0.249	0.102
Rainfall	0.490*	0.398	0.365	0.343

In the last 2 yr, samples were taken at the bottom of the water column in addition to mid-depth. In general, concentrations were only slightly higher than those at mid-depth, reflecting a well-mixed water column (Fig. 3). Occasionally concentrations were substantially higher at the bottom when the water column was stratified. For example, in November 1990 chl *a* concentrations were higher at the bottom than at mid-depth at all sites (Fig. 3).

The pattern of chl *a* concentrations over the 6 yr was compared with those of the environmental variables temperature, salinity and rainfall. Chl *a* was positively correlated with rainfall at Site a ($r = 0.49$, $p < 0.05$), but not at the sites further offshore (Table 1). While there was a trend of higher chl *a* concentrations in the wet season when temperature was higher and salinity was lower, there was no significant correlation with either temperature or salinity. The seasonal differences in temperature and salinity became less marked at consecutive sites offshore.

Nutrients

In 1991–92, nitrate plus nitrite, phosphate and silicate were sampled monthly for 9 mo, 6 mo of which were in the wet season. The concentrations varied considerably from month to month, but there were insufficient data to detect any seasonal trends (e.g. Site b; Fig. 4). The average phosphate concentrations over the period were similar at Sites a, c and d, ranging from 0.10 to 3.0 μM (Table 2). At Site b concentrations were somewhat lower, ranging from 0.09 to 0.47 μM . Nitrate/nitrite concentrations were highest at Site a, which had a mean of 1.15 μM ; the other sites had means of 0.55, 0.67 and 0.67 μM . Silicate concentrations were highest at the inshore sites (a and b), which had means of 6.66 and 7.40 μM respectively, and decreased offshore, where the means were 4.98 and 3.85 μM . The ratio of nitrate to phosphate

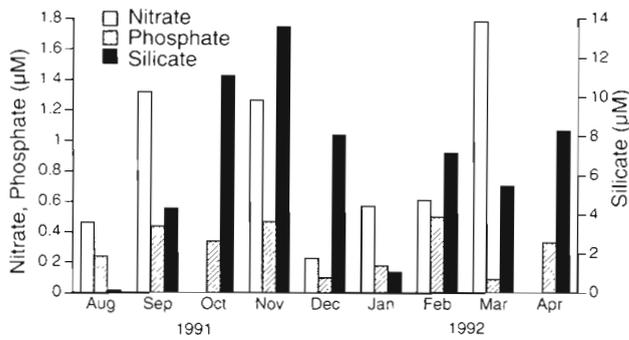


Fig. 4. Nitrate, phosphate and silicate concentrations at the bottom (12 m) of Site b in Albatross Bay from August 1991 to April 1992

ranged from 0.10 to 20, but was generally less than 4 (Table 2).

Net phytoplankton

A total of 70 phytoplankton genera were identified from the 37 µm mesh net samples taken at the 4 sites in Albatross Bay (Table 3). There were 44 diatom genera, 24 dinoflagellate genera, 1 cyanobacterium genus and 1 prymnesiophyte genus. Based on mean abundance (i.e. 2 to 3 = dominant, 1 to <2 = common and 0 to <1 = rare) of the 10 most common genera, 9 were diatoms, e.g. *Rhizosolenia*, *Chaetoceros* and *Thalassionema*, and 1 was a cyanobacterium, *Trichodesmium* (Table 3). The dinoflagellates were generally rare, i.e. a mean abundance of less than 1. A total of 42 genera with a mean abundance greater than 0.100 were examined in more detail.

There were significant differences ($p < 0.01$) in the relative abundance of phytoplankton from the inshore and offshore sites, at different seasons and years

Table 2. Mean and range of phosphate (PO_4), nitrate/nitrite (N_{OXY}), nitrate to phosphate ratio ($\text{N}_{\text{OXY}}:\text{PO}_4$) and silicate (SiO_3) concentrations (µM) at 4 sites in Albatross Bay, Gulf of Carpentaria, sampled at mid depth and bottom, monthly from August 1991 to April 1992

Site	PO_4	N_{OXY}	$\text{N}_{\text{OXY}}:\text{PO}_4$	SiO_3	n
a	0.38	1.15	3.0	6.66	17
	0.13–2.27	0.06–3.68		1.74–10.40	
b	0.26	0.67	2.6	7.40	18
	0.09–0.47	0–1.78		1.06–13.00	
c	0.34	0.67	2.0	4.98	18
	0.10–2.63	0–2.74		1.01–10.00	
d	0.37	0.55	1.5	3.85	18
	0.10–3.00	0–2.35		0.19–8.41	

(Table 4). However, the genera had a similar seasonal pattern at the 4 sites since the interaction between season and site was not significant. Analysis by MDS showed clear differences between inshore and offshore sites in the abundance of the 44 genera (Fig. 5). The MDS configuration plot for all genera (dimension 1 by dimension 2) appears to show 4 distinct groups (Fig. 6). All the most common genera present in the largest cluster were ubiquitous and did not appear to be closely linked with individual sites. One group was more closely associated with Sites c and d. Another group was more closely associated with Sites a and b. The ubiquitous diatom genus *Skeletonema* was not associated with any group.

Comparing the means of individual genera for the wet and dry seasons we found that the differences were explained by only a few genera, most notably the cyanobacterium *Trichodesmium*. It was one of the most abundant genera and bloomed in the wet season each year, becoming rare or absent during the dry season. *Trichodesmium* was dominant more frequently at the offshore sites, where blooming generally began in August/September and lasted for 3 to 6 mo (Fig. 7).

As a group, diatoms were more constant in their abundance and distribution. The dominant diatom genera varied considerably in the timing and duration of their blooms (Fig. 7). At the 2 inshore sites, *Rhizosolenia* had extended periods of dominance — up to 5 mo — but these were not related to season. This genus was often observed to have symbiotic relationships with the nitrogen-fixing cyanobacterium *Richelia*. In 1989–90, *Rhizosolenia* was particularly abundant, while the other diatom genera had a relatively low abundance. *Chaetoceros*, *Thalassionema* and *Bacteriastrium* had periods of dominance at all sites; however these were less prolonged than those of *Rhizosolenia* and the timing varied from genus to genus.

Dinoflagellates were the most stable group. Their overall abundance was lower, and did not vary markedly between seasons and inshore/offshore.

Principal component analysis was carried out to determine the degree of association between the genera. The association was not strong, i.e. the first 10 principal components explained only 52% of the variation, which suggests that the patterns of abundance for individual genera are relatively independent of each other.

Phytoplankton pigments

To obtain an indication of the components of the phytoplankton community that were not sampled in the 37 µm mesh net samples, phytoplankton pigments were also examined.

Fucoxanthin, the indicator pigment for diatoms, prymnesiophytes and chrysophytes, was detected at all 4 sites (Fig. 8). Concentrations ranged from 0 to $1.7 \mu\text{g l}^{-1}$ and were highest in the wet season at the 2 inshore sites (a and b), although they were often higher at Site b than at Site a. The fucoxanthin peaks mirrored the peaks in chl *a* (Fig. 3). Fucoxanthin concentrations at Sites c and d were considerably lower and did not vary seasonally. Concentrations at the bot-

tom of the water column were generally close to those at mid-depth at all sites. Size fractionation showed that fucoxanthin was not restricted to the larger size fractions. In February 1991, for example, it was present equally in the 2 to 10 and greater than 10 μm fractions (Table 5).

Zeaxanthin, the indicator pigment for cyanobacteria and prochlorophytes, was present at the 4 sites throughout the 2 yr of sampling (Fig. 8). Unlike fuco-

Table 3. Phytoplankton genera sampled with 37 μm mesh nets in Albatross Bay, and their mean abundance at 4 sites from April 1986 to April 1992. Abundance based on 3 = dominant, 2 = common, 1 = rare. Cyano: cyanobacteria; Dino: dinoflagellate; Prym: prymnesiophyte. *Subgenus. Codes for genera with a mean abundance > 0.100 are used in Fig. 6

Genus	Code	Class	Mean abundance	Genus	Class	Mean abundance
<i>Rhizosolenia</i>	Rhiz	Diatom	2.099	<i>Diploneis</i>	Diatom	0.079
<i>Chaetoceros</i>	Chae	Diatom	1.979	<i>Podolampas</i>	Dino	0.066
<i>Thalassionema</i>	Than	Diatom	1.901	<i>Heteraulacus</i>	Dino	0.066
<i>Trichodesmium</i>	Tric	Cyano	1.851	<i>Plagiogramma</i>	Diatom	0.054
<i>Bacteriastrium</i>	Bact	Diatom	1.702	<i>Mastogloia</i>	Diatom	0.054
<i>Nitzschia</i>	Nitz	Diatom	1.649	<i>Cerataulina</i>	Diatom	0.050
<i>Thalassiosira</i>	Thas	Diatom	1.541	<i>Asterolampra</i>	Diatom	0.037
<i>Coscinodiscus</i>	Cosc	Diatom	1.236	<i>Surirella</i>	Diatom	0.033
<i>Thalassiothrix</i>	That	Diatom	1.186	<i>Pyrocystis</i>	Dino	0.033
<i>Hemiaulus</i>	Hemi	Diatom	1.178	<i>Prorocentrum</i>	Dino	0.029
<i>Pleurosigma</i>	Pleu	Diatom	1.087	<i>Gymnodinium</i>	Dino	0.029
<i>Protoperidinium</i>	Prpe	Dino	1.083	<i>Ceratocorys</i>	Dino	0.029
<i>Lauderia</i>	Laud	Diatom	0.921	<i>Zygabikodinium</i>	Dino	0.025
<i>Paralia</i>	Para	Diatom	0.860	<i>Phaeocystis</i>	Prym	0.017
<i>Odontella</i>	Odon	Diatom	0.860	<i>Campyloneis</i>	Diatom	0.017
<i>Tripoceratium</i> *	Trce	Dino	0.855	<i>Amphipleura</i>	Diatom	0.017
<i>Blepharocysta</i>	Blph	Dino	0.810	<i>Bellerochea</i>	Diatom	0.012
<i>Detonula</i>	Deto	Diatom	0.806	<i>Epithemia</i>	Diatom	0.008
<i>Dinophysis</i>	Dinp	Dino	0.603	<i>Bernadinium</i>	Dino	0.008
<i>Amphiceratium</i> *	Amce	Dino	0.554	<i>Raphoneis</i>	Diatom	0.004
<i>Ceratium</i>	Crtem	Dino	0.525	<i>Plagiotropis</i>	Diatom	0.004
<i>Diplopsalis</i>	Dips	Dino	0.492	<i>Peridiniopsis</i>	Dino	0.004
<i>Eucampia</i>	Euca	Diatom	0.488	<i>Parahistioneis</i>	Dino	0.004
<i>Gonyaulax</i>	Gony	Dino	0.483	<i>Noctiluca</i>	Dino	0.004
<i>Leptocylindrus</i>	Lept	Diatom	0.463	<i>Histioneis</i>	Dino	0.004
<i>Palmeria</i>	Palm	Diatom	0.434	<i>Heterodinium</i>	Dino	0.004
<i>Amphiprora</i>	Ampr	Diatom	0.377	<i>Dissodinium</i>	Dino	0.004
<i>Bacillaria</i>	Bacp	Diatom	0.343	<i>Balechina</i>	Dino	0.004
<i>Planktoniella</i>	Plan	Diatom	0.335			
<i>Streptotheca</i>	Stth	Diatom	0.326			
<i>Asteromphalus</i>	Astp	Diatom	0.306			
<i>Skeletonema</i>	Skele	Diatom	0.298			
<i>Ditylum</i>	Dity	Diatom	0.264			
<i>Corethron</i>	Core	Diatom	0.260			
<i>Navicula</i>	Navi	Diatom	0.252			
<i>Asterionella</i>	Astn	Diatom	0.252			
<i>Climacodium</i>	Clim	Diatom	0.244			
<i>Ornithocercus</i>	Orni	Dino	0.227			
<i>Amphora</i>	Amra	Diatom	0.215			
<i>Stephanopyxis</i>	Step	Diatom	0.198			
<i>Actinoptychus</i>	Acti	Diatom	0.198			
<i>Guinardia</i>	Guin	Diatom	0.178			
<i>Pyrophacus</i>	Pyph	Dino	0.165			
<i>Amphisolenia</i>	Amph	Dino	0.107			

Table 4. Multivariate analysis of variance and significance tests for the effects of site, season and year on the abundance of 37 μm mesh net phytoplankton genera. F statistics were approximated based on the Wilk's Lambda values. Two seasons were used: May to November (dry), and December to April (wet). df_{num} : degrees of freedom for numerator; df_{den} : degrees of freedom for denominator

Variable	df_{num}	df_{den}	F	p
Site	132	555	2.777	0.0001
Season	44	185	3.669	0.0001
Year	264	1110	3.105	0.0001
Site \times Season	132	555	1.076	0.2842

xanthin, it did not show a seasonal trend, and its concentrations were considerably lower (0 to 0.23 $\mu\text{g l}^{-1}$). Values obtained at mid-depth and bottom were similar at most sampling times. Despite the predominance of the large cyanobacterium *Trichodesmium* in the 37 μm mesh net phytoplankton samples, size fractionation done in February 1991 showed that zeaxanthin was predominantly in the 2 to 10 μm fraction (Table 5). Therefore, in addition to *Trichodesmium*, cyanobacteria in the nanoplankton and possibly *Trichodesmium* fragments are contributors to the phytoplankton biomass.

Chl *b*, the indicator pigment for green algae and prochlorophytes, was present at the 4 sites with few exceptions throughout the 2 yr of sampling (Fig. 8). There appeared to be a seasonal pattern at the in-shore sites and concentrations were somewhat higher overall in 1991–92 than in 1990–91.

Chl *a* degradation products were not detected in any of the samples.

Spatial variation

The spatial variation in chl *a*, temperature and salinity was examined in more detail at 21 sites in Albatross Bay in February 1990. Temperature and salinity gradients were small, with a few exceptions, such as Sites 1 and 2 (Fig. 9). Chl *a* concentrations ranged from 0.06 to 4 $\mu\text{g l}^{-1}$ and increased with depth at all sites with few exceptions (e.g. Site 1; Fig. 9).

Fucoxanthin (the indicator pigment for diatoms, prymnesiophytes and chrysophytes) was the dominant pigment at all sites. The trends mirrored that of chl *a*: where chl *a* concentrations increased with depth, fucoxanthin concentrations also increased.

Size-fractionation studies were done in February 1991 to give an indication of the poten-

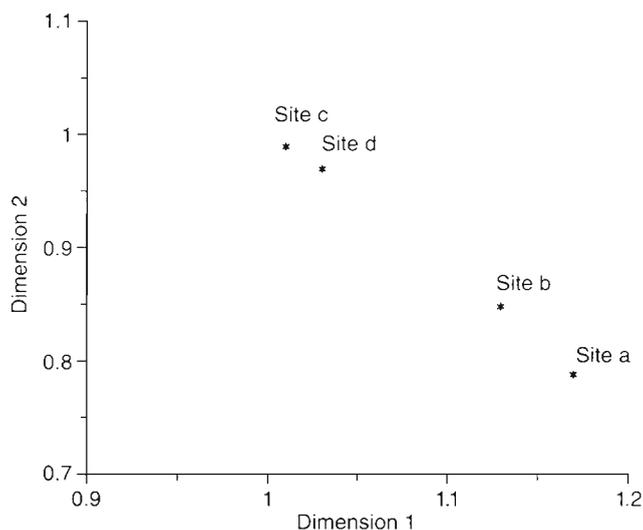


Fig. 5. Multidimensional scaling (MDS) ordination of Sites a, b, c and d based on the dimension coefficients (dimension 1 by dimension 2) of the abundance of large phytoplankton between April 1986 and April 1992

tial contribution of the smaller size fractions to the phytoplankton biomass that were not sampled with 37 μm mesh nets. The highest proportion of chl *a* (0.98 $\mu\text{g l}^{-1}$) was present in the 2 to 10 μm fraction (Table 5). There was also a significant proportion in the >10 μm fraction (0.35 $\mu\text{g l}^{-1}$).

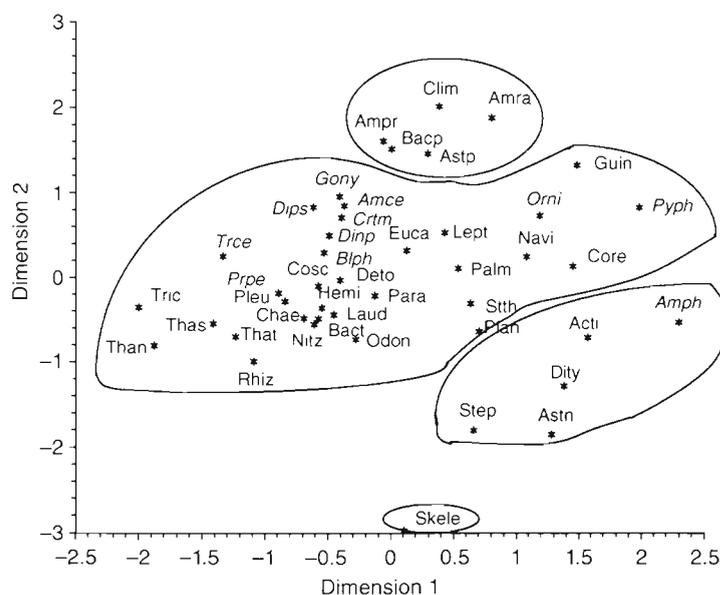


Fig. 6. MDS ordination of 44 phytoplankton genera based on the configuration of the scaling analysis for the period April 1986 to April 1992. For full genus names see Table 3. Dinoflagellate genera codes are italicised

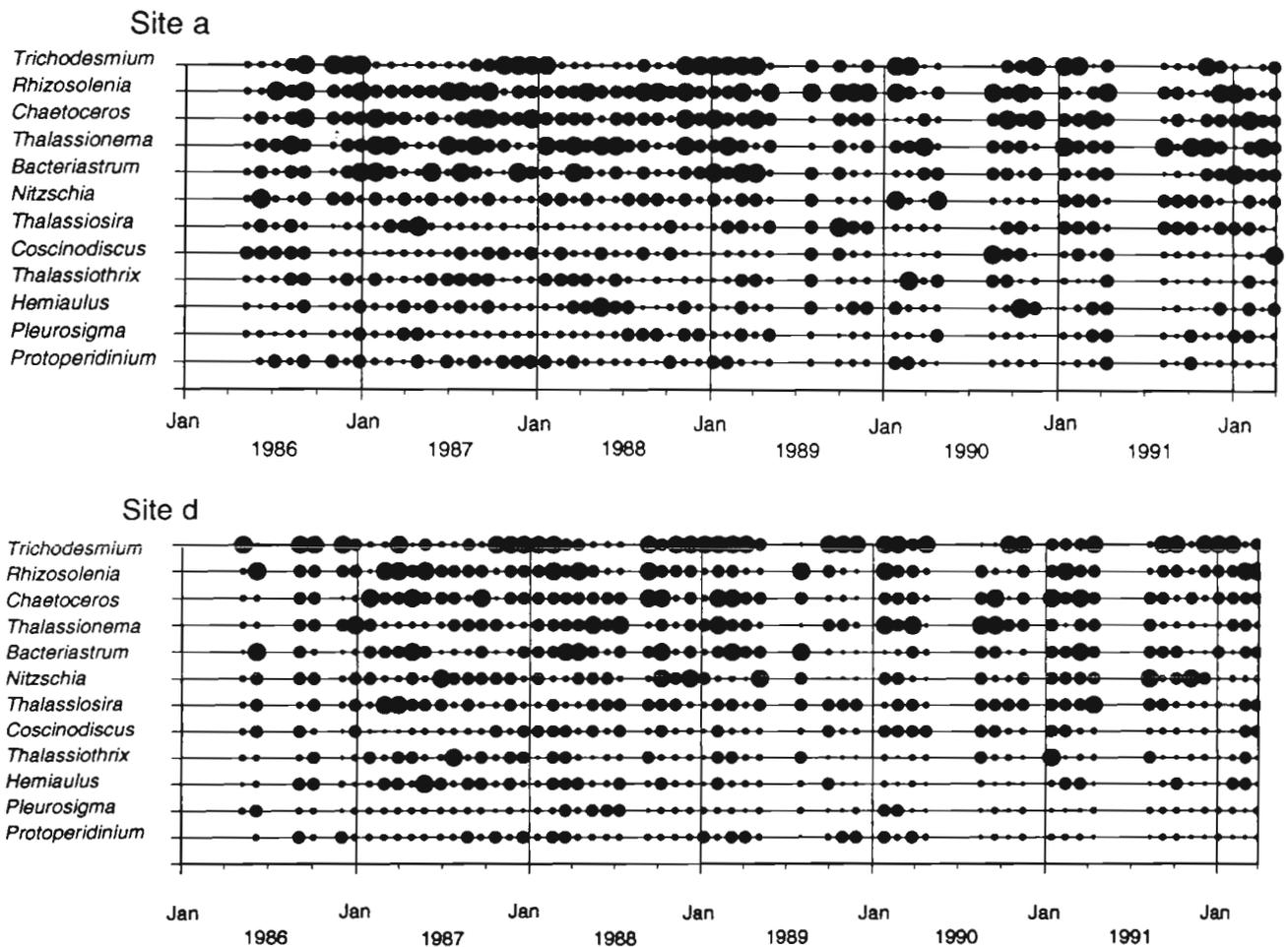


Fig 7. Bubble graph of the 12 dominant 37 μ m mesh net phytoplankton genera at 1 inshore (Site a) and 1 offshore (Site d) site in Albatross Bay from April 1986 to April 1992. Absent, rare, common and dominant categories are represented by 4 bubble sizes; bubble size increased with abundance. Months with no black dots denote no sampling

DISCUSSION

The Gulf of Carpentaria in northern tropical Australia has a climate characterised by wet and dry seasons. In Albatross Bay, in the east of the Gulf, around 70% of the rainfall falls between January and March. The rivers that empty into the Bay further reduce salinities. The winds in the wet season are lighter, moist northeast to northwest monsoons (Somers & Long 1994); in the dry season, they are relatively strong, dry southeasterlies. Water temperatures are higher in the wet than in the dry season.

A coastal boundary layer separates the stratified deeper waters, which receive no terrestrial runoff, from the well-mixed coastal waters (Wolanski & Ridd 1990, Rothlisberg et al. 1994). There is a region of upwelling, at 15 to 20 m depth, at this interface. It lies between the 2 inner sites (a and b) in Albatross Bay and the 2 outer sites (c and d).

The 2 inner sites had wet-season peaks in chl *a* and generally had a higher phytoplankton biomass than the outer sites. Site b generally had higher chl *a* values than Site a, which might be due to its closeness to the region of upwelling, where nutrient availability would be enhanced by mixing.

The wet season peaks in chl *a* at the inshore sites coincided with periods of higher rainfall and temperature. The increased freshwater input from estuaries might be providing nutrients for the phytoplankton; however, no systematic increase in nitrate+nitrite, phosphate and silicate concentrations was detected during the wet season and concentrations were not higher at the inshore sites. Wind mixing might also be stimulating phytoplankton growth by resuspending nutrients from the sediment, but the winds are stronger in the dry season than the wet season. Short-term events, such as storms, also mix the water column. The resulting pulses in nutrients might not be

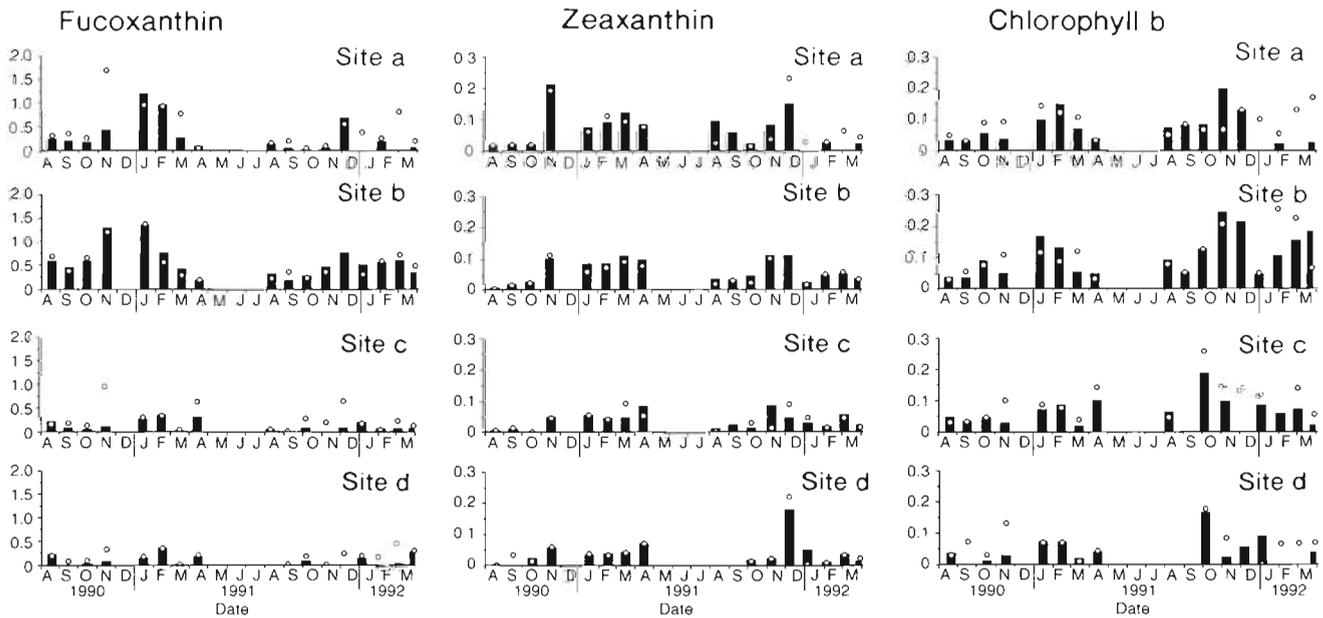


Fig. 8. Fucoxanthin, zeaxanthin and chl *b* concentrations (all in $\mu\text{g l}^{-1}$) at 4 sites from August 1990 to March 1992. Bars are mid-depth concentrations; (o) bottom concentrations. J: January; M: May; S: September. Months in which sampling did not occur have no bars

detectable by monthly sampling. Rainfall may also be stimulating growth by contributing micronutrients; however, they were not measured in this study.

Albatross Bay has low concentrations of nitrate+nitrite ($0.76 \mu\text{M}$); concentrations such as this are generally thought to be limiting to phytoplankton growth (Howarth 1988). In addition, the ratio of nitrate to phosphate present in the water column was low, i.e. 0.5 to 3, compared with the N:P ratio required for phytoplankton growth of 16:1 (Redfield 1958). Ammonia concentrations were not measured in this study and it is possible that when this component of the inorganic nitrogen pool is taken into account the N:P ratio may be higher. However it is unlikely that the ammonia concentrations would be high enough to increase the N:P ratio to 16:1. Howarth (1988) has suggested that many marine systems are nutrient-limited, for example, phytoplankton in the central Great Barrier Reef are nitrogen-limited (Furnas & Mitchell 1986).

Table 5. Albatross Bay, Gulf of Carpentaria: contributions of the >10, 2 to 10 and 0.6 to 2 μm size fractions to the chl *a*, fucoxanthin and zeaxanthin concentrations (mg l^{-1}) at the top (0 m) of the water column at Site a in February 1991

Size fraction (μm)	Chl <i>a</i>	Fucoxanthin	Zeaxanthin
0.6 to 2	0.08	0	0.01
2 to 10	0.98	0.21	0.07
>10	0.35	0.21	0

The concentration of chl *a* in Albatross Bay and its seasonal variation were similar to those found in other tropical coastal regions where eutrophication has not occurred. For example, on the west coast of Thailand, chl *a* concentrations are reported to range from 0.08 to $2.19 \mu\text{g l}^{-1}$ (Janekarn & Hylleberg 1989); in the central Great Barrier Reef phytoplankton populations have a seasonal progression of abundance, being highest in summer, $2 \mu\text{g l}^{-1}$, than in winter, 0.25 to $0.5 \mu\text{g l}^{-1}$ (Furnas & Mitchell 1986).

Chl *a* concentrations in Albatross Bay are higher (up to $5.7 \mu\text{g l}^{-1}$) than those in the deeper waters of the Gulf ($<1.5 \mu\text{g l}^{-1}$) (Hallegraeff & Jeffrey 1984, Rothlisberg et al. 1994). Many of the same 37 μm mesh net phytoplankton genera were observed in Albatross Bay and the central Gulf. For example, the cyanobacterium *Trichodesmium* was a dominant genus throughout the central Gulf (Hallegraeff & Jeffrey 1984) and Albatross Bay. With few exceptions, the dominant diatom genera in the central Gulf also coincided with those in Albatross Bay. However, 7 genera found in Albatross Bay were not found in the deeper waters of the Gulf: the diatoms *Amphipleura*, *Campyloneis*, *Epithemia* and *Plagiotropis*, dinoflagellates *Heteraulacus* and *Zygabikodinium*, and the prymnesiophyte *Phaeocystis*.

Trichodesmium is also an important component of the phytoplankton in other tropical and subtropical regions (Revelante & Gilmartin 1982, Hallegraeff & Jeffrey 1984, Devassy & Goes 1988). As in Albatross Bay, *Trichodesmium* has a high abundance in the

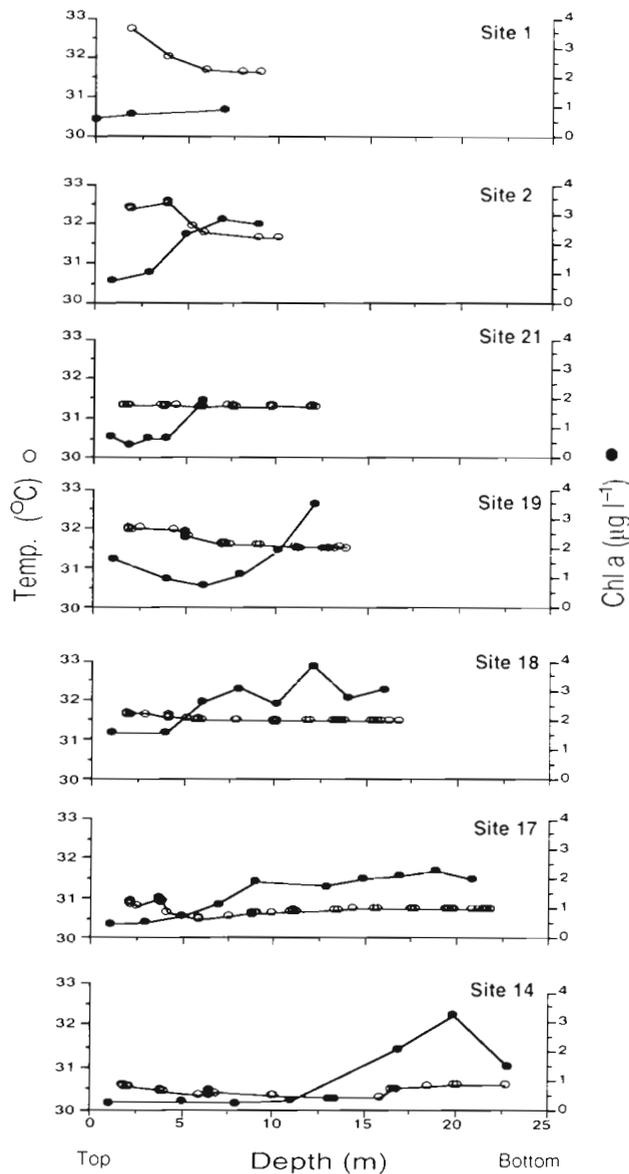


Fig. 9. Depth profiles of chl *a* concentrations (●) and temperature (○) with depth at 7 sites in the northern transect sampled in Albatross Bay in February 1990 (see Fig. 1b)

Great Barrier Reef lagoon from August to January. Extended calm conditions seem to be most conducive to the formation of *Trichodesmium* blooms since mixing created by wind tends to break up the bundles formed in the bloom (Revelante & Gilmartin 1982). These bundles appear to be necessary for nitrogen fixation (Bryceson & Fay 1981), however, the mechanism has not been established (Carpenter et al. 1990). In Albatross Bay *Trichodesmium* bloomed during relatively calm periods in the wet season, and the most extended blooms occurred in the years when there was little or no wind. Revelante & Gilmartin (1982) found

an inverse relationship between *Trichodesmium* and diatom numbers and diversity in the Great Barrier Reef, which suggested *Trichodesmium* has a negative impact on the growth of other phytoplankton. However, this relationship was not apparent in Albatross Bay and *Trichodesmium* may, in fact, be enhancing growth, because it is capable of fixing nitrogen and therefore provides a source of nitrogen for other phytoplankton (Carpenter & Romans 1991). Capone & Carpenter (1982) calculated that *Trichodesmium* contributes about 25% of the total estimated nitrogen fixation in the sea, illustrating its importance in the marine nitrogen cycle.

Continental shelf areas generally have higher proportions of large phytoplankton than small species (Malone 1980). These phytoplankton are generally highly variable in abundance, which may be related to pulses in the biomass of diatoms. In contrast to the distinct succession patterns observed in temperate regions of the world (Guillard & Kilham 1977, Hallegraeff & Jeffrey 1993), the diatoms in Albatross Bay did not show a seasonal pattern of succession. Succession patterns are often determined by the loss of diatoms through diffusion, sinking and grazing, as well as the rate of nutrient regeneration. The lack of a succession pattern in Albatross Bay may be related to the absence of major physical or chemical triggers which generally precipitate a systematic species succession, i.e. the temperature and salinity changes were not dramatic, and nutrient concentrations were generally low.

The abundance and successional changes in the phytoplankton community are generally linked to those of the herbivorous copepod community (Heinrich 1962). In temperate waters this has a seasonal cycle with the biomass of phytoplankton increasing in spring followed by a higher abundance of zooplankton. By contrast, in the Gulf of Carpentaria a 2 yr study of copepod abundance showed that there was no seasonal variation and a generally low abundance (J. G. Greenwood pers. comm.). This is consistent with the lack of a successional pattern in the phytoplankton community and hence the relatively stable nature of these waters.

The 37 µm mesh net phytoplankton was dominated by *Rhizosolenia*, a genus that is symbiotic with the nitrogen-fixing cyanobacterium *Richelia*, particularly when nutrient concentrations are low (Guillard & Kilham 1977), as they are in Albatross Bay. Possibly this strategy for obtaining nitrogen for growth explains the dominance of *Rhizosolenia*. The other dominant diatoms in the 37 µm mesh net phytoplankton fraction in Albatross Bay — *Chaetoceros*, *Thalassionema* and *Bacteriastrum* — also commonly dominate coastal waters in the tropics and subtropics throughout the world (Devassy & Goes 1988, Maclean 1989).

The dominance of diatoms in Albatross Bay may be related to the concentration of silicate. Egge & Aksnes (1992) showed that in experiments diatoms were dominant irrespective of season if the silicate concentration exceeded approximately 2 μM . Concentrations of silicate in Albatross Bay were generally higher than this. The high growth rate of diatoms, i.e. 5 to 50% faster than other algal classes (Egge & Aksnes 1992), and the abundant silicate in the Bay, may explain the preponderance of diatoms.

The size-fractionation and pigment studies showed that there is a significant nanoplankton fraction (i.e. <10 μm) in Albatross Bay. It includes *Navicula jeffreyi*, *Minutocellus polymorphus* and *Pseudostaurosira shiloi*, which were not observed in the 37 μm mesh net phytoplankton samples (Hallegraeff & Burford in press).

The relative contributions of the phytoplankton classes of all sizes to the overall algal biomass has been approximated by Everitt et al.'s (1990) method of using pigment indicator ratios: chl *a*:fucoxanthin of 1.4, chl *a*:zeaxanthin of 1.7, chl *a*:chl *b* of 0.75. By multiplying the pigment concentration by the ratios, its contribution to the chl *a* concentration can be determined. The average percentage contributions of fucoxanthin, zeaxanthin and chl *b* at the inshore sites were 66, 9 and 6% respectively over the 2 yr that HPLC pigment analysis was done. The high concentrations of fucoxanthin confirm that the phytoplankton community is diatom rich. In contrast, the offshore sites had a lower proportion of diatoms, a similar proportion of cyanobacteria and a higher proportion of green flagellates, i.e. 46% fucoxanthin, 11% zeaxanthin and 12% chl *b*. The deeper waters of the Gulf generally have a larger and more stable nanoplankton component (Hallegraeff & Jeffrey 1984, Rothlisberg et al. 1994). These small size fractions do not undergo marked seasonal cycles (Furnas & Mitchell 1986).

Albatross Bay is a spawning ground for the banana prawn *Penaeus merguensis* and tiger prawn *P. semisulcatus*. There are 2 peaks of reproduction in the Bay: spring and autumn. The resulting larvae feed exclusively on phytoplankton during the first week of growth (Dall et al. 1990). The abundance of larvae varies considerably from year to year (Rothlisberg et al. 1988), but in 6 yr larval numbers did not correlate with chl *a* concentrations. Short-term feeding experiments have also been conducted in Albatross Bay in spring and autumn with *P. merguensis* larvae reared in *in situ* cages (Preston et al. 1992). They confirmed that the diatom-rich community present at these times provided a nutritionally adequate food source for prawn larvae. In contrast the cyanobacterium genus *Trichodesmium*, which is dominant in Albatross Bay, appears to have a negative effect on larval growth (N. P. Preston unpubl. data). This effect was

examined by feeding a *Trichodesmium* species to prawn larvae in the laboratory and measuring larval growth and survival. In short-term trials, larval survival was not affected but the growth rate was reduced. However the extent to which *Trichodesmium* affects larval growth and mortality in Albatross Bay is unknown.

In summary, Albatross Bay has summer peaks in phytoplankton biomass, probably caused by a combination of higher temperatures, increased rainfall and calmer weather. It is a relatively stable environment in terms of the lack of seasonal succession in the diatom-dominated phytoplankton community, low nutrients and the dominance of phytoplankton species commonly associated with nutrient-depleted waters (e.g. *Rhizosolenia*). The cyanobacterium genus *Trichodesmium* was an exception: its seasonal peaks in abundance appear to be linked to calmer weather conditions. The deeper waters out from Albatross Bay had phytoplankton biomass and species composition similar to those in the central Gulf.

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