

Influence of phytoplankton concentration and wave exposure on the ecophysiology of *Mytilus californianus*

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ABSTRACT: The relationship between nearshore food availability and physiological state of the suspension-feeding mussel *Mytilus californianus* was examined along a wave-exposure gradient at 2 rocky intertidal sites on the Oregon coast, USA, differing in nearshore concentrations of phytoplankton. The RNA:DNA ratio (an indirect measure of protein synthetic capacity) of adductor muscle was used as an indicator of dietary status and growth. The activities of 2 metabolic enzymes, citrate synthase (CS) and malate dehydrogenase (MDH), were measured for gill (CS) and adductor (MDH) as indicators of metabolic activity. Mussels were sampled at wave-exposed and wave-protected areas at Strawberry Hill (SH), a site with relatively high phytoplankton concentrations, and Boiler Bay (BB), a site with relatively low phytoplankton concentrations. Individuals at wave-exposed areas at both SH and BB had higher RNA:DNA ratios in spring and summer than those at wave-protected areas. RNA:DNA ratios in spring and summer were higher at SH than at BB, and these differences were especially pronounced following coastal upwelling events. Increases in RNA:DNA ratio during phytoplankton blooms after upwelling suggest that mussels respond rapidly to short-term changes in nearshore oceanographic conditions. Both CS and MDH activities were higher at SH than at BB many times throughout the year, suggesting metabolic activity was higher overall for SH mussels. Reciprocal transplant experiments were performed to evaluate the degree of physiological plasticity in relation to site- and exposure-related factors (e.g. phytoplankton concentration, heat and/or desiccation stress, wave forces). Between-site transplants were done between wave-exposed areas at SH and BB; within-site transplants were done between areas of different wave-exposure at each of the 2 sites. RNA:DNA ratios of transplanted mussels converged on those of the site or area to which they were transplanted, suggesting that mussel physiology is highly plastic in response to site and area-related variations in environmental factors. The strong apparent response of mussels to variations in phytoplankton concentration suggests that physiological indices may be useful tools in assessing the magnitude of 'bottom-up' effects in rocky intertidal communities.

KEY WORDS: *Mytilus californianus* · Rocky intertidal · Ecophysiology · Food availability

INTRODUCTION

Organisms living in rocky intertidal regions experience spatial and temporal variability in environmental conditions due to steep gradients in wave impact, food availability, temperature, and desiccation (Paine 1977, Newell 1979, Menge & Farrell 1989). Biochemical and

physiological processes of intertidal organisms can vary over a wide range of values in response to this environmental variation, and organismal performance (growth, reproduction, mobility, etc.) may therefore be modified in response to environmental conditions (Hawkins & Bayne 1984, Stickle et al. 1985, Menge & Sutherland 1987, Stickle & Bayne 1987, van Erkom Schurink & Griffiths 1992, Hofmann & Somero 1995). These physiological adjustments may have profound effects on the distribution and abundance of a single species, as well as on that species' interactions with

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other species in its habitat, and therefore on the entire structure of a given rocky intertidal community. Because key species interactions often vary significantly along these same environmental gradients (Connell 1961a, b, Dayton 1971, Paine 1974, Lubchenco 1978, Menge 1978a, b, 1991, Menge & Sutherland 1987, Menge et al. 1994), it is likely that physiological mechanisms underlie environmentally induced variation in community structure. To gain a more detailed understanding of the link between one environmental variable, the availability of food, and the physiological ecology of the rocky intertidal region, we examined biochemical indices of nutritional status in an ecologically important species of marine invertebrate, the suspension-feeding mussel *Mytilus californianus*.

Variation in food and nutrient availability has profound effects on the physiology of intertidal suspension feeders (Bayne & Widdows 1978, Hawkins & Bayne 1984, Stickle & Bayne 1987, Bertness et al. 1991, van Erkom Schurink & Griffiths 1992, Bayne et al. 1993, Kreeger et al. 1995). Suspension-feeding invertebrates living in the rocky intertidal region may experience variation in food quality and quantity, nutrients, and in feeding opportunities within and between sites due to strong environmental gradients in phytoplankton productivity (see below), quality of food, wave exposure, relief of substratum, and tidal level (Navarro & Winter 1982, Bayne & Hawkins 1990, Hawkins & Bayne 1992). Suspension-feeding animals living in wave-exposed areas may have more or longer feeding opportunities than do individuals in wave-protected areas because of greater water flow at wave-exposed sites. Although mussels may compensate for lower feeding time at wave-protected sites with higher feeding efficiency (Kreeger et al. 1991), thermal and desiccation stress at wave-protected areas may suppress metabolism (Roberts et al. in press).

Variation in the availability of food to suspension-feeding invertebrates resulting from variation in nearshore phytoplankton concentration or feeding time may influence the structure of rocky intertidal communities (Menge & Olson 1990, Menge et al. 1995, 1996). Although the importance of bottom-up effects (so named because regulating processes originate at the base of the food web) on community structure has long been debated (e.g. Oksanen et al. 1981, Fretwell 1987, Carpenter 1988, McQueen et al. 1989), recent studies along the Oregon coast suggest that rocky intertidal community structure and regulation may vary with nearshore phytoplankton concentration and primary productivity (hereafter PP) (Menge 1992, Menge et al. 1994, 1995). Because energy input of nearshore PP into intertidal communities occurs through suspension-feeding invertebrates (primarily mussels and barnacles), examination of the effects of variation in PP and

feeding opportunity on the dietary physiology of suspension feeders is a logical step to elucidating the mechanisms by which variation in PP input affects rocky intertidal community structure.

We examined mussels living at 2 rocky intertidal habitats on the Oregon coast, Strawberry Hill (hereafter SH: 44° 15' N, 124° 07' W) and Boiler Bay (hereafter BB: 44° 40' N, 124° 03' W). Patterns of species distribution and abundance differ strikingly between these 2 sites (Menge et al. 1994). At BB, macrophyte abundance in the low zone is relatively high while abundance of sessile suspension feeders (e.g. mussels and barnacles), and invertebrate herbivores and predators (e.g. seastars and whelks) is relatively low. Conversely, at SH, macrophyte abundance is relatively low while abundance of suspension-feeding invertebrates, invertebrate herbivores and predators is relatively high.

Phytoplankton abundance and productivity, indexed by chlorophyll *a* (hereafter chl *a*) and ¹⁴C respectively, were consistently higher at SH than at BB (Menge et al. 1995, 1996). From February to June in 1993 and 1994, for example, monthly water samples taken showed that the average concentration of chl *a* at SH was $3.37 \pm 2.00 \mu\text{g l}^{-1}$ and $5.00 \pm 1.22 \mu\text{g l}^{-1}$ respectively, whereas during those same times, chl *a* at BB was $0.55 \pm 0.20 \mu\text{g l}^{-1}$ and $1.87 \pm 0.90 \mu\text{g l}^{-1}$. These patterns are even more pronounced during the summer after periods of coastal upwelling, when chl *a* concentrations at SH are up to 5-fold greater than those at BB. For example, on 2 July 1993, chl *a* concentration was $50 \pm 4.8 \mu\text{g l}^{-1}$ at SH and $11.0 \pm 2.9 \mu\text{g l}^{-1}$ at BB; on 16 July 1994, chl *a* concentrations were $29 \pm 3.5 \mu\text{g l}^{-1}$ at SH and $7.2 \pm 5.0 \mu\text{g l}^{-1}$ at BB (Menge et al. 1995). These data suggest that suspension feeders living at SH have more food available in most months than do suspension feeders living at BB, and that this difference is extreme during and after coastal upwelling.

Adjustments in rates of physiological processes may be necessary to bring metabolic demands of organisms into alignment with available energy supplies (Hawkins & Bayne 1984, Hochachka & Somero 1984, van Erkom Schurink & Griffiths 1992, Bayne et al. 1993). Therefore, variation in food availability may be reflected in variation in metabolic activity. Previous studies of marine organisms have shown the potential for using biochemical data to develop links between metabolic activity and food availability. For example, depriving some organisms of food, or significantly reducing their abilities to feed, results in a concomitant decrease in the activities of metabolic enzymes relative to fed controls (Sullivan & Somero 1983, Yang & Somero 1993, Stillman et al. 1994, Dahlhoff & Menge 1996). The activities of metabolic enzymes may therefore be considered an approximate index of dietary-

dependent changes in metabolic activity *in situ*. Another physiological index that is sensitive to changes in dietary status of many organisms is the RNA:DNA ratio, as it reflects the protein synthetic capacity of a given tissue at any time (Buckley 1984, Wright & Hetzel 1985, Martinez et al. 1992, Foster et al. 1993, Stillman et al. 1994). In this study, the activities of metabolic enzymes (citrate synthase and malate dehydrogenase) and RNA:DNA ratios were used to monitor changes in metabolic activity of mussels exposed to seasonal and site-related differences in food and nutrient availability at SH and BB.

MATERIALS AND METHODS

Study organism. *Mytilus californianus* was selected for study as it is a major component of rocky intertidal communities along much of the west coast of North America and is abundant at BB and SH. *M. californianus* inhabits the mid-intertidal zone on exposed coasts from southern Baja California to the Aleutian Islands. It is a major competitor for space along wave-swept rocky shores, is a favored prey of certain predators, particularly the seastar *Pisaster ochraceus*, and is thus a central species in community dynamics (Paine 1966, 1974, 1984). Furthermore, the distribution, abundance, and relationships between mussels and other species at SH and BB have been well-studied (Menge 1992, Menge et al. 1994, Navarrete & Menge 1996).

Mussel collection. Starting in May 1993, mussels were collected every 90 d at wave-exposed and wave-protected areas at SH and BB. Sampling during July (1993 and 1994) was conducted 5 to 10 d after the peak of the large, upwelling-induced increases in chl *a* (Menge et al. 1995; see also 'Introduction'). Ten individuals for each site-exposure treatment were collected at randomly determined locations along a 10 m transect line laid parallel to the shore at each wave-exposed and wave-protected area. Individuals were placed in a cooler on ice and brought to the laboratory, where they were immediately dissected, weighed, and frozen on dry ice. Dissected tissues were stored at -70°C until biochemical analyses were conducted.

Upwelling measurements. Water samples were collected during and after coastal upwelling in June and July 1994. Upwelling was evidenced by strong north-west winds, as well as a decrease in nearshore water temperature. The concentration of chl *a* in nearshore water was determined fluorometrically (Menge et al. 1995). Water samples were collected directly at each site and were processed within 14 h of sampling. Mussels were collected from the exact location of water sampling before, during and after upwelling. Individuals were placed in a cooler on ice and brought to the

laboratory, where they were immediately dissected, weighed, and frozen on dry ice. Dissected tissues were stored at -70°C until analyzed.

RNA:DNA ratios. Concentrations of RNA and DNA in adductor muscle were determined by ethidium bromide fluorescence following the method of Bentle et al. (1981) as modified by Stillman et al. (1994). Tissues were thawed on ice, weighed and homogenized in 20 to 30 volumes 2M NaCl (dilution depended on tissue type) with a hand-driven glass homogenizer (Kontes Duall, Vineland, NJ, USA). From each sample 50 μl was incubated in 1.5 ml of 0.005 mg ml^{-1} ethidium bromide and 0.10 mg ml^{-1} proteinase K at 37°C for 90 min. After incubation, 0.5 ml buffer (80 mM Tris-Cl, pH 7.5 at 20°C) was added, and fluorescence was recorded at 365 nm excitation and 590 nm emission using a Perkin-Elmer LS-5B luminescence spectrofluorometer. RNA and DNA concentrations of tissues were estimated from a standard curve calculated by measuring the fluorescence of known quantities of RNA and DNA (Sigma calf thymus DNA, 1 to 4 μg ; Sigma calf liver RNA, type IV, 2 to 8 μg).

Citrate synthase and malate dehydrogenase activities. Tissues were thawed on ice, weighed and immediately homogenized in 10 volumes ice-cold 50 mM potassium phosphate buffer (pH 6.8 at 20°C) with a glass homogenizer, as described above. The homogenate was centrifuged at 4°C in a fixed rotor microfuge for 5 min at $14000 \times g$, and the supernatant was collected and used for enzymatic activity determinations. Enzymes were assayed spectrophotometrically following the methods of Yang & Somero (1993) and Stillman et al. (1994). Enzymatic activities are reported in International Units per gram wet mass (IU g^{-1}).

Mussel transplants. Mussels were collected in May 1994 from wave-exposed and wave-protected areas at both SH and BB and transplanted using the methods of Menge et al. (1994). Mussels were collected at comparable tidal heights at both SH and BB and were reciprocally transplanted across sites and wave exposures [n (number of transplanted clumps) = 4]. To control for transplant effects, mussels were also transplanted within their site of origin. A single set of transplants ($n = 4$) served as the control for both site and wave-exposure manipulations.

Clumps of 20 mussels each were held against the rock with cages of Vexar plastic mesh to allow the mussels to reattach byssal threads to the rock. After 14 d, most of the mussels had reattached and cages were loosened to free the mussels from the restraint of the mesh. Cages were left in place over the mussel clumps in order to exclude *Pisaster ochraceus*. Until October 1994, 3 individuals were removed from each cage every 30 d. Specimens were handled for biochemical analysis as described previously.

Statistical analyses. One-way, univariate and multivariate ANOVA were used to compare seasonal and site-related variation of RNA:DNA ratios and enzymatic activities of mussels. Univariate and multivariate repeated-measures ANOVA and 1-way ANOVA were used to analyze data from mussel transplant studies. Statistical analyses were conducted on non-transformed data using SYSTAT (Release 5.0 for Macintosh; Wilkinson 1990). Assumptions of normality, independence of error terms, and homogeneity of variances were evaluated and found to fit the assumptions of the models used.

RESULTS

Biochemical variation in natural populations

RNA:DNA ratio: differences between sites and exposures

In field-collected individuals, RNA:DNA ratios for adductor muscle of mussels from wave-exposed areas at SH were significantly higher than RNA:DNA ratios of mussels from BB in July of 1993 and 1994 (compare circle data points for July in Fig. 1A, B; $p < 0.050$, 1-way ANOVA; probabilities adjusted with Bonferroni approximation for multiple comparisons). RNA:DNA ratios were also significantly greater at the SH wave-protected area than at the BB wave-protected area in July of 1993 and 1994 (compare triangle data points for July in Fig. 1A, B; $p < 0.050$). With the exception of higher RNA:DNA ratios at the BB wave-exposed area in autumn 1993 ($p < 0.050$), RNA:DNA ratios of *Mytilus californianus* did not differ between SH and BB in other seasons; that is, differences between sites were time-dependent (Site \times Time interaction was significant; Table 1). The July collections that resulted in the significant differences between SH and BB were made

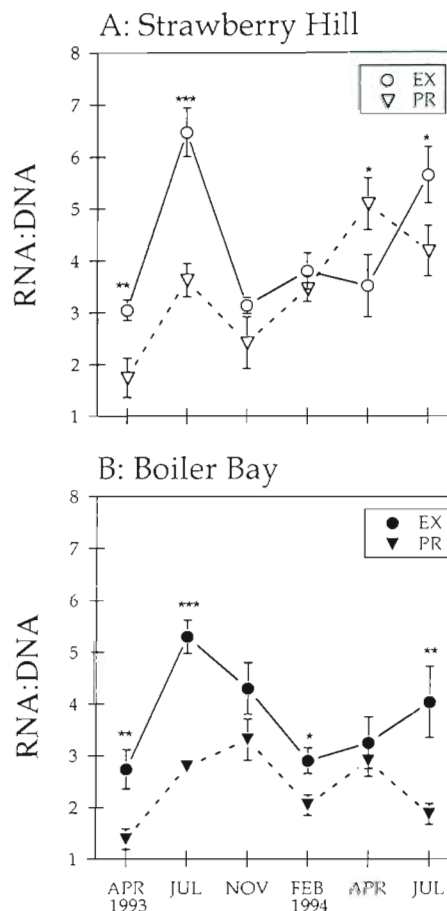


Fig. 1 *Mytilus californianus*. Changes in protein synthetic capacity, as indexed by RNA:DNA ratio, of mussels from wave-exposed (EX) and wave-protected (PR) areas at (A) Strawberry Hill and (B) Boiler Bay (Oregon, USA). Data points are means (± 1 SE) of 10 individuals that were collected 90 d apart. Asterisks indicate significance of comparison between exposures by date (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). Additional statistical analysis in Table 1

Table 1 Three-way ANOVA on the effect of site, exposure, and time on the RNA:DNA ratio of *Mytilus californianus*. Site: Strawberry Hill and Boiler Bay; exposure: wave exposed, wave protected; time: every 90 d from May 1993 to July 1994. Site, exposure, and time were treated as fixed variables. ns: $p > 0.050$

Source of variation	df	MS	F	p
Site	1	43.15	19.79	<0.0005
Exposure	1	71.51	32.80	<0.0005
Time	5	24.89	11.41	<0.0005
Site \times Exposure	1	1.468	0.673	ns
Site \times Time	5	11.60	5.322	<0.0005
Exposure \times Time	5	11.81	5.417	<0.0005
Site \times Exposure \times Time	5	0.343	0.158	ns
Error	202	2.180		

5 to 10 d following coastal upwelling events that occurred in late June to early August (Menge et al. 1995; Fig 2).

RNA:DNA ratios of *Mytilus californianus* also differed between wave exposures independent of site (Fig. 1A, B; Site \times Exposure interaction was not significant; Table 1). At SH, RNA:DNA ratio was greater at wave-exposed than at wave-protected areas in April and July 1993 and 1994; values in other seasons were not different (Fig. 1A; p-values for 1-way ANOVA shown). Similarly, RNA:DNA ratios were greater at wave-exposed than at wave-protected areas at BB in April

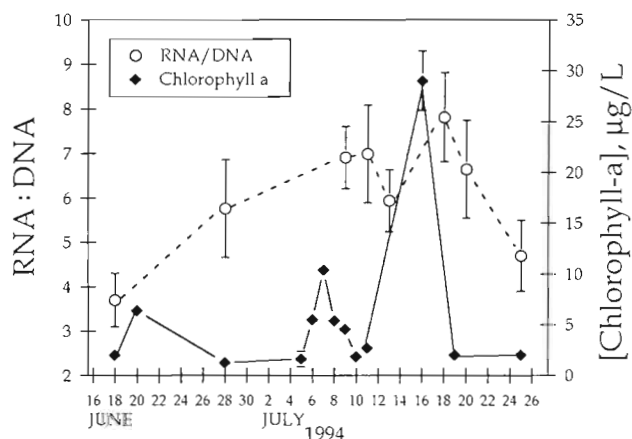


Fig. 2. *Mytilus californianus*. Changes in RNA:DNA ratio of mussels and chl *a* concentrations during coastal upwelling at Strawberry Hill. Data points are means (± 1 SE) of values for 10 individuals. * $p < 0.050$ for a 1-way ANOVA of RNA:DNA ratio between mussels collected on that day and on 18 June 1994, before coastal upwelling began. Concentrations of chl *a* are in $\mu\text{g l}^{-1}$ (\pm SD) of seawater ($n = 5$) collected at wave-exposed sites at SH on those days

and July 1993 and February and July 1994; values for other times were not different (Fig. 1B; p values for 1-way ANOVA shown). Exposure differences were time-dependent (Exposure \times Time interaction is significant; Table 1). Therefore, during much of the year, but especially during the summer months, mussels in wave-exposed areas had higher RNA:DNA ratios than did individuals in wave-protected areas.

RNA:DNA ratio: rapid response during coastal upwelling

RNA:DNA ratios of adductor muscle for *Mytilus californianus* from wave-exposed areas at SH were significantly higher than those for mussels collected on 18 June, 2 to 8 d after rapid increases in chl *a* concentrations (7 July and 16 July) (p -values for 1-way ANOVA are shown in Fig. 2). RNA:DNA ratios returned to 'baseline' levels within 4 to 8 d after a drop in chl *a* concentrations (Fig. 2). The increase in RNA:DNA ratio was correlated with increases in chl *a* at least 3 times in 1994, although not all increases in

chl *a* resulted in significant increases in mussel RNA:DNA ratio.

Metabolic activity: differences between sites

Metabolic activity of *Mytilus californianus*, as indexed by enzyme activities of gill and adductor muscle, differed with site and exposure by season [Table 2: Site \times Exposure \times Time interactions were significant for citrate synthase (CS) and malate dehydrogenase (MDH) in multivariate and univariate analyses]. Some of these differences are illustrated in Fig. 3. MDH activity of adductor muscle (Fig. 3A) was higher for mussels from wave-exposed areas at SH than at BB in both April and July 1993 and April 1994. CS activity of gill (Fig. 3B) was higher for SH mussels in April 1993 and February, April, and July 1994. While differences in metabolic activity between mussels at SH and BB were less pronounced in fall and winter, clear seasonal patterns in these indices were not evident (Fig. 3). Instead, CS and MDH at SH tended to remain rela-

Table 2. ANOVA and MANOVA of the effect of site, exposure, and time on the enzymatic activity of *Mytilus californianus*. Site, exposure and time: as in Table 1. ns: $p > 0.050$

ANOVA	df	MS	F	p
CS activity				
Site	1	0.518	3.716	ns
Exposure	1	0.217	1.553	ns
Time	5	2.577	18.48	<0.0005
Site \times Exposure	1	0.556	3.990	0.0470
Site \times Time	5	0.322	2.378	0.0400
Exposure \times Time	5	1.889	13.55	<0.0005
Site \times Exposure \times Time	5	0.434	3.111	0.0100
Error	186	0.139		
MDH activity				
Site	1	8194.7	63.89	<0.0005
Exposure	1	10258	79.98	<0.0005
Time	5	1448.7	11.29	<0.0005
Site \times Exposure	1	63.84	0.498	ns
Site \times Time	5	1390.6	10.84	<0.0005
Exposure \times Time	5	751.48	5.859	<0.0005
Site \times Exposure \times Time	5	1017.5	7.932	<0.0005
Error	186	128.27		
MANOVA				
Source of variation	df	Wilks' lambda	F	p
Site	2185	0.7344	33.45	<0.00005
Exposure	2185	0.6959	40.42	<0.00005
Time	10370	0.5150	14.56	<0.00005
Site \times Exposure	2185	0.9766	2.216	ns
Site \times Time	10370	0.7332	6.209	<0.00005
Exposure \times Time	10370	0.6525	8.805	<0.00005
Site \times Exposure \times Time	10370	0.7610	5.413	<0.00005

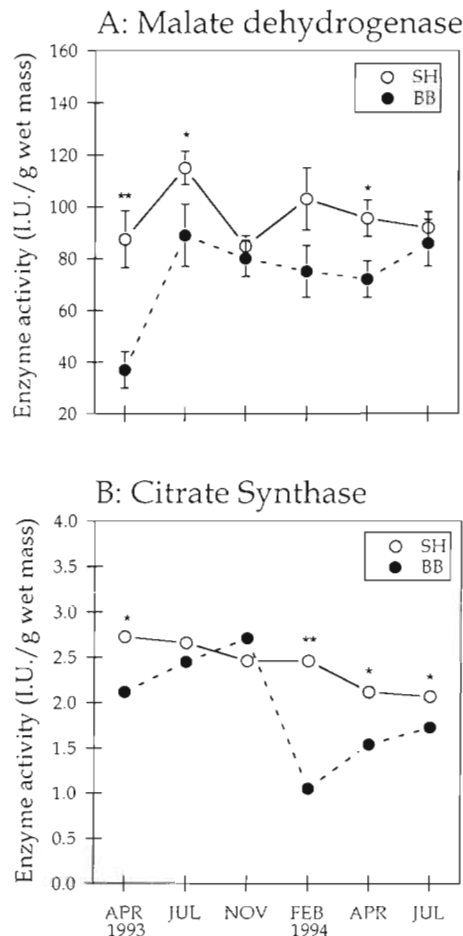


Fig. 3. *Mytilus californianus*. Changes in (A) adductor muscle malate dehydrogenase and (B) gill citrate synthase activities of mussels from wave-exposed areas at Strawberry Hill (SH) and Boiler Bay (BB). Data points are means (± 1 SE) of 10 individuals collected 90 d apart. Asterisks indicate significance of comparison between sites by date (* $p < 0.05$, ** $p < 0.01$).

Additional statistical analysis of these data in Table 2

tively constant, while values for animals at BB tended to fluctuate. No differences were observed in enzymatic activities for mussels at wave-protected sites (data not shown).

Biochemical variation under experimental conditions: mussel transplants

The mussel transplant experiments tested the hypothesis that differences in mussel physiology were influenced by environmental factors (see 'Materials and methods'). Prior to transplantation in May 1994, RNA:DNA ratios of adductor muscle of mussels at SH were higher than those of mussels at BB (Fig. 4; 1-way ANOVA, $p < 0.0001$). After transplantation,

RNA:DNA ratios of mussels transplanted between sites (wave-exposed only) changed rapidly (Fig. 4). Within 4 wk (June), RNA:DNA ratios of mussels transplanted from BB to SH (between-site transplants) were indistinguishable from those of mussels transplanted from SH back to SH (SH controls), and were significantly different from RNA:DNA ratios of mussels from the site of origin, the BB controls (June; Fig. 4, Table 3; 1-way ANOVAs). Similarly, RNA:DNA ratios of mussels transplanted from SH to BB were indistinguishable from BB control mussels within 8 wk (July; Fig. 4, Table 3). These differences persisted through August, after which RNA:DNA ratios for between-site transplants inexplicably converged, as did those for within-site transplants (October; Fig. 4).

The effects of wave exposure on RNA:DNA ratios of transplanted mussels was much smaller than effects of site (Fig. 5). Repeated-measures analysis indicated that there was no significant effect of exposure transplant over time at either SH or BB, except for August 1994 at SH ($p > 0.050$ for all cases except August). In August at SH, RNA:DNA ratios of mussels transplanted from exposed to protected areas were significantly lower than RNA:DNA ratios of exposed transplant control mussels and indistinguishable from those of protected transplant control mussels (1-way ANOVA, $p < 0.0001$). Similarly, RNA:DNA ratios of protected to exposed mussels were significantly higher than those of protected transplant control mussels, and were indistinct from RNA:DNA ratios of exposed transplant control mussels (Fig. 5A; 1-way ANOVA, $p < 0.0001$).

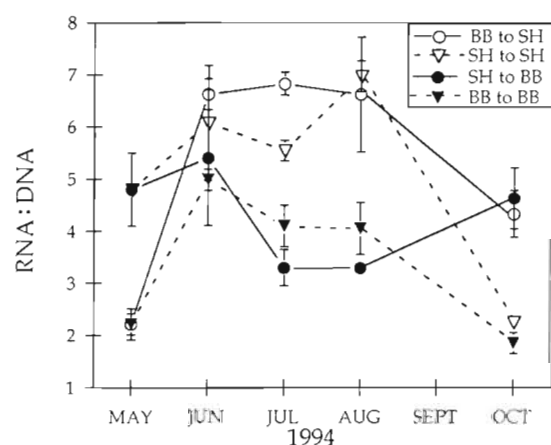


Fig. 4. *Mytilus californianus*. Changes in RNA:DNA ratios of mussels transplanted between wave-exposed areas at SH and BB. Data points are means (± 1 SE) of values for 12 individuals, obtained by combining 3 mussels from each of 4 cages for each time point. Statistical analysis of these data shown in Table 3

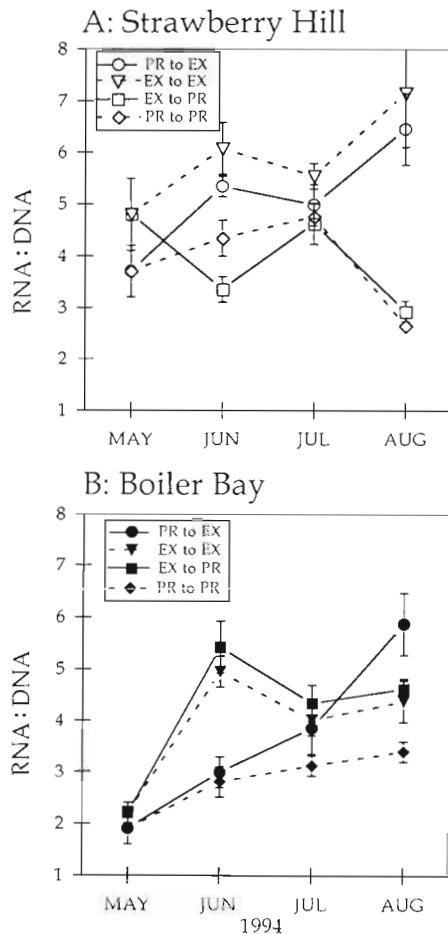


Fig. 5. *Mytilus californianus*. Changes in RNA:DNA ratios of mussels transplanted between wave-exposed and wave-protected areas at (A) SH and (B) BB. Data derived as for Fig. 4

and RNA:DNA ratios decreased after chl *a* levels dropped to pre-upwelling values (Fig. 2); (3) RNA:DNA ratios of transplanted mussels were indistinguishable from those of nearby control mussels and were significantly different from those of control mussels at the site of origin within 4 to 8 wk of transplantation, depending on site of origin and transplant treatment (Fig. 4, Table 3)

These data suggest that mussels may be highly acclimatable in response to site-related differences in nearshore PP. This high degree of physiological plasticity may have significant ramifications for community structure. Mussels are a major ecological player in wave-swept intertidal communities. Given that PP is different between BB and SH (Menge et al. 1995), and that SH is dominated by suspension-feeding invertebrates (Menge et al. 1994), the observation that mussels respond rapidly to changes in nearshore PP supports the hypothesis that 'bottom-up' effects may be an

important determinant of differences in community structure between BB and SH. While such a mechanistic link between environmental conditions and ecological performance is still untested, the observation that the physiological status of a major competitive dominant in the wave-swept rocky intertidal region appears to be linked to site- and exposure-related differences in nearshore food availability suggests one mechanism by which community structure may be influenced by environmental gradients in food availability.

Differences between SH and BB had a strong seasonal component; that is, the highly significant effects of site on differences between RNA:DNA ratio and enzyme activities are time-dependent (Tables 1 & 2). Seasonal effects are illustrated by the observation that mussels collected in July 1993 and 1994 had higher RNA:DNA ratios and enzyme activities than mussels collected in November 1993 or February 1994 (Figs. 1 & 3, Tables 1 & 2). Not surprisingly, chl *a* concentrations have been higher in summer (June to August) than in winter (November to February) each year from 1992 to 1996 (Menge et al. 1995, 1996, 1997). During times of low productivity (November to February), chl *a* and nutrient concentrations were slightly higher at SH than BB. During times of high productivity (June to August), the differences between site were much greater, with chl *a* concentrations up to 5-fold higher at SH than at BB (e.g. Menge et al. 1995, 1996). The observation that differences in the physiology of suspension-feeding mussels at BB and SH are most pronounced during periods of upwelling and high productivity, and are weak or non-existent during times of low productivity, supports the hypothesis that there is a link between nearshore productivity and mussel physiology.

Seasonal variation in the quantity of food available is likely to have strong effects on mussel physiology, as our data suggest. However, seasonal and spatial variation in the composition of phytoplankton and the availability of other food sources can influence mussel physiology as well, as illustrated by the following examples. (1) There is a relationship between composition of the diet and metabolic activity of mussels (Hawkins & Bayne 1992, Kreeger 1993, Kreeger & Langdon 1993, Fidalgo et al. 1994). For example, *Mytilus galloprovincialis* and *M. trossulus* fed phytoplankton with high protein content had significantly higher growth rates than conspecifics fed low protein phytoplankton (Kreeger & Langdon 1993, Fidalgo et al. 1994). (2) Mussels can adjust metabolism depending upon the composition of their diet (Hawkins & Bayne 1992). (3) Mussels have additional food sources besides phytoplankton, such as detritus and dissolved organic matter, and mussels may utilize these sources during times of low PP (Manahan 1990, Bayne & Hawkins

1990). (4) Mussels may alter feeding behavior significantly in response to variation in diet (Willows 1985, Bayne et al. 1993, Sanford et al. 1994). For example, *M. edulis* that were fed on diets low in organic content increased both feeding rate and absorption efficiency relative to individuals fed on a high organic content diet (Bayne et al. 1993). These examples illustrate the fact the composition of the diet, as well as feeding behavior, must be considered when discussing the effects of variation in food availability *in situ*. Because the composition of phytoplankton may be different at different sites, further studies of the links between nearshore productivity and the physiology of suspension-feeding invertebrates at these sites will be necessary to dissect differences caused by the composition versus the quantity of food.

A major physiological factor that may differ between mussels at any given time or site is reproductive state. Some suspension feeders (most notably the mussel *Mytilus edulis*) show annual fluctuations in gonad development and spawning in response to a number of factors, including variation in food availability, and gonad development can affect RNA:DNA ratio of tissues (Seed 1975, Robbins et al. 1990, Seed & Suchanek 1992). However, variation in gonad development was probably not a confounding factor in this study. Gonad development was monitored, and no significant differences in gonad production (as indexed by gonad mass g^{-1} total wet mass) were observed between SH and BB at any time of the year (data not shown). These observations are in agreement with studies which demonstrated that gonad development in *M. californianus* does not show the strong cyclicity that is observed in other species (Suchanek 1981, Seed & Suchanek 1992). While there are factors that alter gonad production in *M. californianus* (notably disturbance), neither season nor site in the present study seemed to influence gonad production, so that differences in RNA:DNA ratio observed between BB and SH were likely due to other factors.

Wave exposure effects

RNA:DNA ratios were higher for adductor muscle of mussels from wave-exposed areas than for conspecifics from wave-protected areas (Fig. 1). These significant effects of wave exposure on RNA:DNA ratio were observed for individuals from both SH and BB and were most pronounced during spring and summer, when differences in nearshore productivity, as well as potential differences in thermal stress, are greatest (Fig. 1, Table 1). Comparisons of mussels at the top (high-exposed) and bottom (low-exposed) of a wave-exposed bed at SH showed no significant differences

in RNA:DNA ratio or enzyme activities in July 1993 and 1994, so the effects of wave exposure on mussel physiology is probably more related to degree of wave exposure, rather than tidal height per se (data not shown). That is, high-exposed mussels get splashed almost as much as low-exposed mussels, and have lower body temperatures than wave-protected mussels (Roberts et al. in press). Changes in the RNA:DNA ratios of adductor muscle of mussels transplanted between wave-exposed and wave-protected areas at SH and BB showed that wave-protected mussels had significantly lower RNA:DNA ratios in August, independent of site of origin. Mussel body temperatures preceding the August collection were high (31°C) relative to June (21°C) and July (22°C) (authors' unpubl. obs.). These observations implicate elevated temperatures as a major factor influencing differences in physiology for mussels living at different wave exposures.

Recent studies on mussels at SH show that differences in body temperature that result from differences in wave exposure are associated with differential expression of stress-inducible proteins, specifically heat-shock protein 70 (hsp-70) (Roberts et al. 1996). Mussels transplanted to wave-protected areas had significantly higher concentrations of hsp-70 than those at wave-exposed areas (Hofmann & Somero 1996, Roberts et al. in press). Also, the amount of thermally damaged protein, measured by the levels of ubiquitin conjugates in tissues of mussels from SH, is greater for mussels living at wave-protected areas than for individuals at wave-exposed areas (Hofmann & Somero 1996). The repair of thermally damaged proteins (using hsp-70 as a catalyst to facilitate re-folding of thermally damaged proteins) or removal of irreversibly damaged proteins from the total protein pool (via ubiquitination of damaged proteins and subsequent degradation) utilizes ATP that could be used for anabolic processes (cf. Lindquist 1986). Mussels living at wave-protected areas at SH and BB do grow more slowly and are smaller than those at wave-exposed areas (Menge et al. 1994). The observation that wave-protected mussels have lower RNA:DNA ratios than wave-exposed mussels may be because wave-protected mussels 'turn down' anabolic processes to compensate for energy used to repair thermally damaged proteins (Roberts et al. in press).

Another major factor that is likely to influence the physiological status of mussels living at distinct wave exposures is the amount of time available for mussels to feed. Previous studies have suggested that suspension feeders will experience greater delivery of food and greater feeding opportunities in wave-exposed areas due to high flow and greater 'splash' at wave-exposed areas (Menge 1978a, b, 1983, Denny 1988, Bertness et al. 1991). The results of the present study

suggest that, on average, wave-exposed mussels are making more new protein than wave-protected individuals. However, from these data it is impossible to distinguish physiological variation due to amount of time available for the mussels to suspension feed and the effects of elevated body temperatures.

We investigated the relationship between nearshore food availability and the physiology of the mussel *Mytilus californianus* to gain insight into mechanism(s) underlying variation in community structure between rocky intertidal regions experiencing differences in nearshore phytoplankton productivity. The data presented in this paper suggest a strong link between nearshore phytoplankton productivity and the physiology of a dominant suspension feeder, *M. californianus*, that is reflected by differences in site, season, and wave exposure. We therefore conclude that the examination of the metabolic activity of ecologically important species can provide valuable insight when examining links between nearshore oceanic processes and rocky intertidal community structure.

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