

Development of swimming abilities in reef fish larvae

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ABSTRACT: Recent studies have revealed that reef fish larvae have excellent sustained swimming capabilities and considerable potential for modifying their dispersal patterns by active swimming. However, these studies concentrate solely on the late pelagic phase. We examined the development of swimming abilities from hatching through to settlement in 3 reef fish species (*Pomacentrus amboinensis*, *Sphaeramia nematoptera*, *Amphiprion melanopus*). Larval rearing provided larvae at all stages of development. Experiments were conducted in flow chambers designed for measuring the critical and sustained swimming capability of young larvae. In all 3 species, critical swimming ability increased steadily with age, size, relative propulsive area and developmental stage of the larvae. In contrast, sustained swimming ability showed a marked inflection during development. Differences among species throughout development appear to reflect variations in the developmental patterns of the 3 species. Propulsive area was highly correlated with swimming ability and may prove useful for estimating swimming capabilities among species. The results suggest that some species have the potential to actively modify their dispersal patterns from an early age.

KEY WORDS: Larvae · Swimming · Dispersal · Coral reef fish · Development · Morphology

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INTRODUCTION

Coral reef fishes are generally sedentary as adults, with most possessing a dispersive larval phase. This phase lasts for several days or weeks, and during this time they may travel considerable distances. The distribution and abundance of reef fishes is significantly influenced by the dispersal of this planktonic larval stage (Doherty 1991). Knowledge of the extent of larval dispersal among reefs is an essential prerequisite for understanding reef-community dynamics and management (Williams et al. 1984). There has been considerable debate about the relative importance of active behaviour in determining dispersal and distribution patterns of reef fish larvae. The traditional view, which is still maintained by some authors, is that fish larvae have very little behavioural influence over their dispersal patterns and that currents are responsible for the dispersal of larvae to and from reefs (e.g. Roberts

1997). Other workers, however, have suggested that larval behaviour, including vertical and horizontal movement, may influence dispersal patterns (e.g. Cowen et al. 1993, Leis et al. 1996, Stobutzki & Bellwood 1997).

Recent work has shown that settlement-stage larval reef fishes have considerable swimming abilities over both short-term and prolonged periods. Using a current flume, Stobutzki & Bellwood (1994) showed that the short-duration swimming speeds of larvae are relatively high, *Pomacentrus amboinensis*, for example averaging 43.7 cm s^{-1} , or approx. 40 body lengths (BL) s^{-1} . Leis et al. (1996) and Leis & Carson-Ewart (1997) also recorded high swimming speeds, with capture-release methods recording speeds of $>60 \text{ cm s}^{-1}$ for reef fish larvae in the field. These speeds were maintained for several minutes. The sustained swimming abilities of many species were also remarkable. Stobutzki & Bellwood (1997) recorded some species swimming over 100 km in a single bout, lasting up to 10 d, at a speed of 13.5 cm s^{-1} . These studies describe different measures of swimming ability. The methods employed

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by Stobutzki & Bellwood provide data on endurance swimming which represents a maximal performance measure of maintained swimming ability. In comparison, the observations of Leis et al. were based in the field with unrestrained larvae, the larvae selecting their swimming speed over relatively short distances. Leis & Stobutzki (1997) suggested that similarities in the results of the 2 measures indicates that they provide comparable measures of overall abilities. Methodological differences notwithstanding, both approaches have shown that larval fish are capable of considerable horizontal movement.

The abilities of larval fishes to influence their dispersal remains a controversial topic and an area of considerable debate (Roberts 1997, 1998, Bellwood et al. 1998, Sale & Cowen 1998). While many authors admit that larval behaviour may be important (reviewed by Leis 1991a), few have considered these abilities in the theories and models put forward. Several authors have attempted to explain distribution patterns without considering active movement of larvae (Booth & Baretta 1994, Roberts 1997). In such cases, it has been assumed that there is no development of swimming ability throughout the larval period. The results of previous studies have clearly demonstrated that this assumption is not valid (Leis & Carson-Ewart 1997, Stobutzki & Bellwood 1997). An alternative assumption that has been made is that swimming ability develops rapidly immediately prior to the late-pelagic phase and that this ability is used to move towards reefs at the time of settlement (Wolanski et al. 1997). Such models attempting to include larval behaviour have been limited by the lack of knowledge of swimming abilities in younger larvae. The larvae used in the studies by Stobutzki & Bellwood (1994, 1997), Leis et al. (1996), Leis & Carson-Ewart (1997) and Stobutzki (1998) were all caught in light traps and represent late pelagic-stage larvae (Choat et al. 1993). If swimming ability develops earlier in the larval period, then the potential for horizontal swimming to modify dispersal patterns will be greatly underestimated in these dispersal models.

Before we can understand the potential importance of active swimming in larval reef-fish dispersal, we need to determine the swimming ability of different species of reef fishes throughout the entire larval period, rather than just the late-pelagic stage. Investigations into the abilities of younger stages of larval reef fishes has been limited by difficulties in obtaining live individuals in the field in good condition. Larval rearing techniques, however, provide a means of determining the biological capabilities of larval fishes throughout development. By hatching and rearing larvae under controlled conditions, it is possible to obtain live specimens of known age and parentage, in good condition, at all developmental stages. Using captive

rearing techniques, the aim of this study was to determine how swimming ability develops with age for several reef fish species. The specific questions to be addressed were: (1) How do critical and sustained swimming ability vary among species during larval development and (2) To what extent is swimming ability related to morphological attributes of the larvae?

MATERIALS AND METHODS

Three species of reef fishes were examined: the ambon damselfish *Pomacentrus amboinensis* (Pomacentridae; Pomacentrinae), the coral cardinal *Sphaeramia nematoptera* (Apogonidae), and the anemonefish *Amphiprion melanopus* (Pomacentridae; Amphiprioniinae). All were collected from the northern Great Barrier Reef. The species selected differed markedly in terms of their larval ecology. *P. amboinensis* and *S. nematoptera* have long larval durations of about 20 and 24 d respectively, whereas *A. melanopus* has a very short larval duration of approx. 9 d. Adult brood stock were kept in >1000 l aquaria and were fed a diet of chopped pilchards, prawns and *Ascetes* twice a day. Rearing methods follow Job & Bellwood (2000). Larvae were reared in 200 l aquaria at 27.5 to 29°C, under a 14:10 h light:dark photoperiod, throughout the study period. The alga *Nannochloropsis* sp. was used to green the water during the day. Larvae were fed >52 µm-filtered, wild-caught plankton, supplemented occasionally by rotifers and *Artemia* sp., at prey densities of between 2 and 6 ind. ml⁻¹.

Swimming experiments for older larvae were carried out using the technique and swimming flume described by Stobutzki & Bellwood (1997). Two additional swimming channels were designed for use with younger larvae (Fig. 1). These channels operated at slower speeds (between 0 and 15 cm s⁻¹) and had a much wider swimming area to minimise the area influenced by boundary layers. The swimming channels of the 2 chambers were 15 cm wide × 6.5 cm deep × 35 cm long and 10 cm wide × 5.5 cm deep × 30 cm long, for the 'slow' and 'medium' speeds, respectively. Both channels could be placed in aquaria with greened water so that there was no change in water quality between the experimental apparatus and the rearing tanks. Both swimming channels were calibrated using video and neutral-density particles.

Each batch of eggs from each species was raised from hatching through to settlement. Experiments were performed throughout the larval period, with sampling days depending on the species. The first swimming trial was conducted on Day 1, approx. 12 h after hatching. Three batches of each species were used for each swimming trial to allow for batch effects.

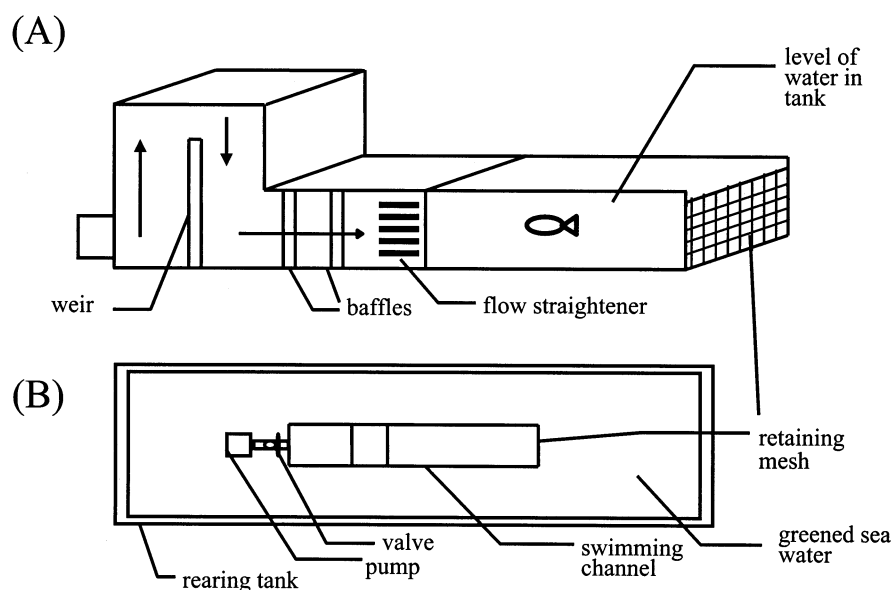


Fig. 1. Swimming channel for younger larvae. (A) Lateral view of apparatus; (B) top view of apparatus in experimental tank

At each experimental age, 8 to 12 fish were used from each batch for the swimming trials. The mean of these 8 to 12 fish was used for statistical analyses. In all experiments, larvae were transferred to the appropriate swimming chamber and allowed to acclimatise for 2 to 15 min. Fish that exhibited symptoms of stress such as erratic swimming behaviour, lying on the bottom or in corners, or clinging to the sides or surface of the swimming channel were excluded from the experiments.

The critical swimming speed gives an indication of the maximum sustainable swimming speed of a fish, and involves increasing the flow rate against which the fish is swimming in gradual increments. In the present study, larvae were subjected to incremental increases in flow rates equivalent to approx. 3 BL every 2 min until they could no longer maintain position for the full 2 min. The equation used to calculate the critical swimming speed (U_{crit}) of larvae followed Brett (1964):

$$U_{crit} = U + (t/t_i \times U_i)$$

where U = penultimate speed, U_i = velocity increment (3 BL), t = time swum in the final velocity increment, and t_i = set time interval for each velocity increment (2 min).

Allometric studies have demonstrated that the highest level of activity, achieved at the top speed within any one gait, is size-dependent, and thus at any given absolute speed different-sized fish may not be at similar relative levels of exertion (Drucker 1996). In order to compare the sustained swimming abilities of larvae

of different species at different ages, sustained swimming trial speeds were set as a multiple of body length. Fish were placed in the appropriate swimming channel and swam at a speed equivalent to approx. 11 body lengths (BL) per second until exhausted. Larvae were observed continuously for the first 5 min, every 10 min for the first hour, and every 2 to 4 h thereafter. The total duration recorded represented of the swimming time up to the last occasion on which an individual was observed swimming. Actual swimming durations may be up to 4 h longer for the older larvae.

Fishes that had swum, or siblings of the same age, were anaesthetised in chilled water and then fixed in 10% buffered formalin. After 12 to 48 h, larvae were transferred to 70% alcohol and stored. Measurements were taken of total length (from the tip of the caudal fin to the tip of the upper jaw), body depth (the height of the fish measured at the deepest region), body area (the entire area of the fish in lateral view excluding the fins) and total propulsive area (the area of the fish including the fins but excluding the head and gut region). Although larvae exhibit some degree of shrinkage (approx. 10%), this varies among ages and parameters (Job & Bellwood 1996). For consistency, all values are based on raw (non-corrected) measurements. The Reynolds numbers (Re) for the sustained swimming trials were calculated for the mean of each batch at each age using the formula: $Re = U \times L/\nu$ (after Webb & Weihs 1986), where U = sustained swimming speed ($m\ s^{-1}$), L = body depth (m), and ν = kinematic viscosity of sea water ($1.03 \times 10^{-6}\ m^2\ s^{-1}$).

Scatterplots and regressions were used to investigate the relationship between swimming ability and age. Analyses were based on both age since hatching and developmental age. Developmental age provides a way of comparing larvae between species at similar ontogenetic stages and was calculated following Job & Bellwood (2000): developmental age = total age/total larval duration, where total age = age of fish (days post-hatch) + egg duration and total larval duration = larval duration (days post-hatch) + egg duration.

Scatterplots and regressions were also used to describe the development of swimming ability with increasing size. To investigate the relationship between swimming ability and changes in the shape of larvae, measurements were standardised by dividing each variable by total length. Zero-order, as well as partial correlation coefficients were used to determine which of the relative morphological variables were most useful for modelling swimming ability. For all regressions, dummy variable analysis using corner-point parameterisation was used to determine if there were significant differences in either the slopes or intercepts for each species (Silk 1976). This technique involves comparing the sum of squares obtained for the most complex model (each species has different slopes and intercepts) with simpler models (2 or 3 species have the same slope and/or intercept) using *F*-tests. Where multiple *F*-tests were used, a Bonferroni correction was applied. The assumptions of all regressions were tested graphically, and data were \log_{10} -transformed when appropriate. In the case of sustained swimming time, transformation was unable to linearise the relationship with all of the parameters investigated. Consequently, statistical analyses based on the assumption of linearity were not used and the development of sustained swimming ability was described using scatterplots.

RESULTS

Critical swimming ability increased with age for all 3 species (Fig. 2A). Initially, critical swimming ability increased slowly with age, then improved rapidly for older stages, with the exception of *Sphaeramia nematoptera*, in which critical swimming ability increased steadily throughout development (Fig. 2A). At 12 h after hatching, all 3 species had some critical swimming ability, with mean critical swimming speeds ranging from 1.8 cm s^{-1} for *S. nematoptera* to 4.6 cm s^{-1} for *Amphiprion melanopus* (Table 1). The mean critical swimming speeds obtained just prior to settlement ranged from 10.9 cm s^{-1} for *S. nematoptera* to 30.3 cm s^{-1} for *Pomacentrus amboinensis* (Table 1).

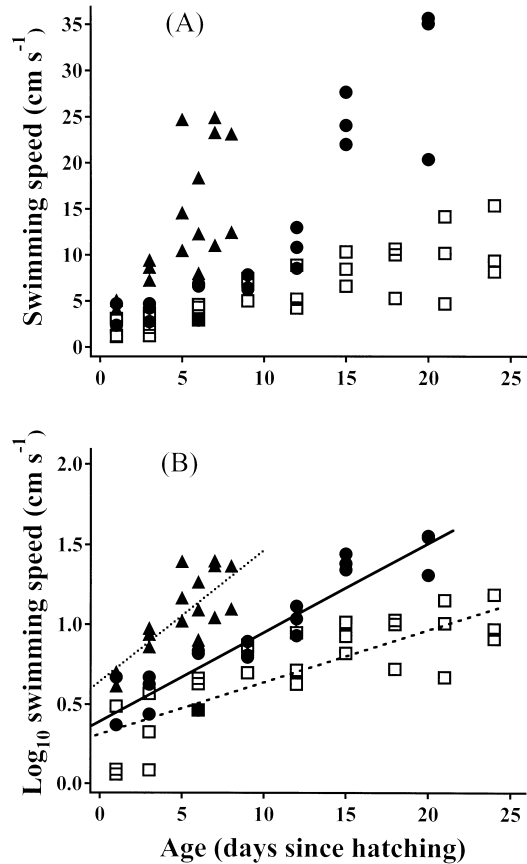


Fig. 2. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Development of critical swimming speed with age. (A) Raw values; (B) \log_{10} -transformed values

Table 1. *Pomacentrus amboinensis*, *Sphaeramia nematoptera*, *Amphiprion melanopus*. Mean and maximum critical swimming speed and sustained swimming time 12 h after hatching and at settlement. Values in parentheses = SE (n = 3 batches of 8 to 12 fish)

Species	Critical speed (cm s^{-1})		Sustained swimming time (h)	
	Mean	Max.	Mean	Max.
At hatching				
<i>P. amboinensis</i>	3.5 (1.2)	6.4	0.11 (0.04)	0.5
<i>S. nematoptera</i>	1.8 (0.6)	4.4	0.02 (0.02)	0.2
<i>A. melanopus</i>	4.6 (0.2)	8.2	0.26 (0.12)	1.1
At settlement				
<i>P. amboinensis</i>	30.3 (5.0)	48.1	90.4 (31.9)	234
<i>S. nematoptera</i>	10.9 (2.2)	24.2	4.3 (4.3)	39.9
<i>A. melanopus</i>	14.6 (4.5)	35.7	8.6 (4.8)	44.25

The relationship between critical swimming ability and age differed significantly among the 3 species. In terms of critical swimming ability, *Pomacentrus amboinensis* and *Amphiprion melanopus* have a very

similar developmental rate with age (Fig. 2B), with no significant difference between the slopes ($F_{3,60} = 1.15$, $p = 0.34$). *A. melanopus*, however, hatches with significantly greater critical swimming abilities than the other 2 species ($F_{1,58} = 218.1$, $p < 0.0001$) (Fig. 2B). *Sphaeramia nematoptera*, although hatching at a similar state of development in terms of critical swimming ability as *P. amboinensis* ($F_{3,60} = 0.15$, $p = 0.34$), has a significantly slower rate for the development of swimming ability with age ($F_{1,58} = 194.5$, $p < 0.0001$), taking longer to reach much slower final speeds than *P. amboinensis* (Fig. 2B).

All 3 species hatched with very little sustained swimming abilities, and each developed this ability quite differently with age. *Sphaeramia nematoptera* had the poorest sustained swimming ability throughout development, exhibiting very little sustained swimming at any age (Fig. 3A). *Pomacentrus amboinensis* had the greatest sustained swimming ability, and this developed rapidly 12 to 15 d after hatching (Fig. 3A). *Amphiprion melanopus* had shorter sustained swimming times than *P. amboinensis*, with a mean of 8.6 h at the oldest age compared to 90.4 h for *P. amboinensis* (Table 1). *A. melanopus*, however, did display the earliest development of swimming ability, with some

degree of sustained swimming ability between Days 3 and 5 after hatching (Fig. 3A). All 3 species had considerably higher maximum swimming times, and the mean values displayed here are clearly a conservative estimate of the potential swimming abilities of the 3 species (Table 1). Although most sustained swimming values for *S. nematoptera* were near zero, the maximum values obtained indicated that at 24 d some individuals were able to swim for up 39.9 h (Table 1). A similar pattern is obtained if the equivalent distance that the larvae have travelled is calculated, based on the speeds at each age and the overall time swum (Fig. 3B). All 3 species swam very short distances immediately after hatching. *A. melanopus* was only able to swim very short distances throughout the larval period, although this species was able to swim around 4 km just 5 d after hatching (Fig. 3B). In contrast, *P. amboinensis* exhibited no sustained swimming abilities until 15 d after hatching. Although developing sustained swimming ability at a much slower rate, *P. amboinensis* was capable of swimming 12 km at 15 d post-hatch and 40 km at 20 d post-hatch (Fig. 3B). *S. nematoptera*, on average, was able to swim only very short distances throughout the larval period (Fig. 3B).

Critical swimming ability increased exponentially with developmental age in all 3 species (Fig. 4A). There was no significant differences in the slopes for all species, or in the intercepts for *Sphaeramia nematoptera* and *Amphiprion melanopus* ($F_{3,61} = 2.17$, $p = 0.10$), indicating that all 3 species develop critical swimming abilities at a similar rate in terms of developmental age. Although *Pomacentrus amboinensis* was found to have a significantly different intercept ($F_{3,61} = 4.9$, $p = 0.004$), this difference was much less than that seen when the development of critical swimming ability is compared using age since hatching.

The 3 species exhibited different sustained swimming abilities, even at similar developmental ages. *Pomacentrus amboinensis* had the greatest sustained swimming ability, followed by *Amphiprion melanopus* and then *Sphaeramia nematoptera*. The sustained swimming ability of *S. nematoptera* was very poor at all ontogenetic stages (Fig. 4B). Although the magnitudes of sustained swimming ability were different for *P. amboinensis* and *A. melanopus*, these species developed sustained swimming abilities at a similar stage of ontogenetic development (Fig. 4B), in marked contrast to the onset of abilities in terms of age since hatching (Fig. 3A).

Critical swimming speed increased proportionally with total length for all 3 species, but the rate of development and the values of critical swimming ability for a given length differed among species. *Amphiprion melanopus* increased swimming ability with increasing length at a significantly faster rate than the other

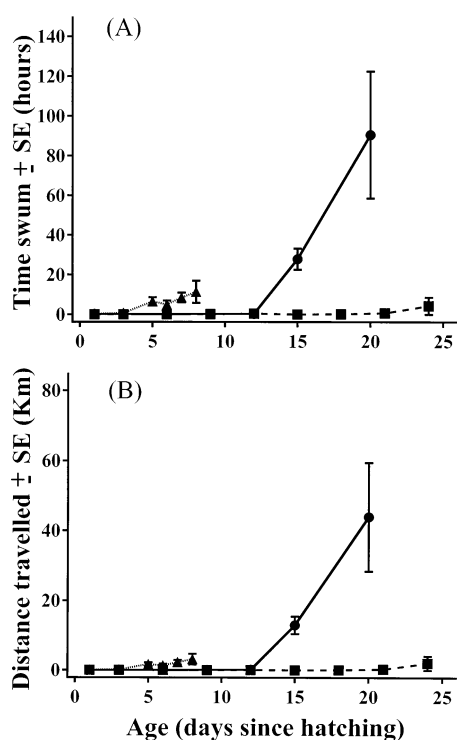


Fig. 3. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (■) and *Amphiprion melanopus* (▲). Development of sustained swimming ability with age. (A) Sustained swimming time in hours; (B) equivalent distance travelled

2 species ($F_{1,58} = 10.09$, $p < 0.001$) (Fig. 5B). *Pomacentrus amboinensis* and *Sphaeramia nematoptera* developed swimming ability at a similar rate with increasing length (Fig. 5B), with no detectable difference between these rates ($F_{1,58} = 0.37$, $p = 0.54$). However, there was a significant difference in the intercepts ($F_{2,59} = 11.62$, $p < 0.0001$), with *P. amboinensis* having considerably better swimming abilities for a given length than *S. nematoptera* (Fig. 5B). Sustained swimming time also increased with increasing total body length, but only for *P. amboinensis* and *A. melanopus*. *S. nematoptera* exhibited very little sustained swimming ability at any length (Fig. 6). *P. amboinensis* developed sustained swimming ability at a body length of around 11.5 mm, with shorter larvae exhibiting very little sustained swimming ability (Fig. 6). *A. melanopus* appeared to develop sustained swimming abilities smoothly with increasing length, and showed some sustained swimming abilities even at a length of only 5 or 6 mm, which is at a considerably shorter body length than *P. amboinensis* (Fig. 6).

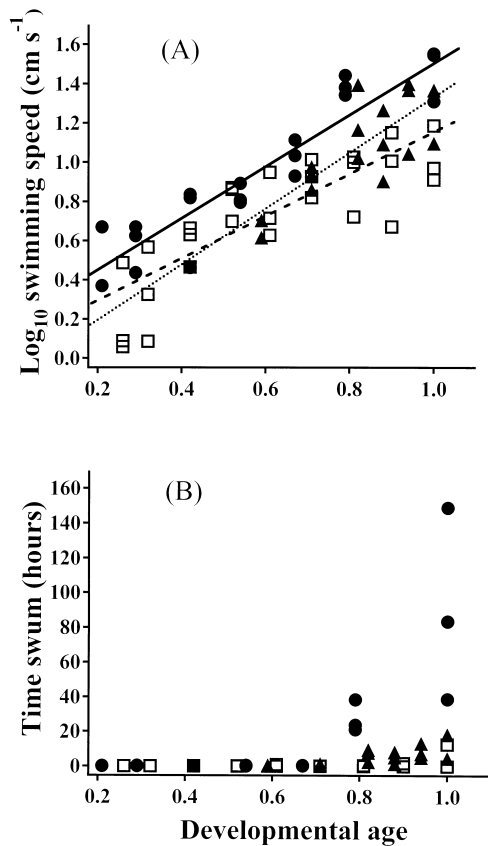


Fig. 4. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Swimming ability vs developmental age. (A) Critical swimming speed; (B) sustained swimming time

Partial correlation coefficients for the 3 morphological variables indicated that for sustained and critical swimming ability, relative propulsive area described the greatest amount of unique variation in nearly all cases (Table 2). A plot of relative propulsive area against critical swimming ability suggested a similar relationship for all 3 species (Fig. 7A). Statistical analysis indicated that there was only a marginal difference between the slope for *Pomacentrus amboinensis* and the other 2 species ($F_{4,61} = 2.71$, $p = 0.04$). Overall, critical swimming ability increased similarly with increasing relative propulsive area for all species. While there was no simple linear relationship between sustained swimming time and relative propulsive area, there was a similar pattern for all 3 species (Fig. 7B). There appeared to be a common value of propulsive area relative to length at which sustained swimming ability first occurs for all 3 species. The Reynolds numbers experienced by larvae throughout their developed increased steadily from hatching through to settlement. For all species, a significant increase in sus-

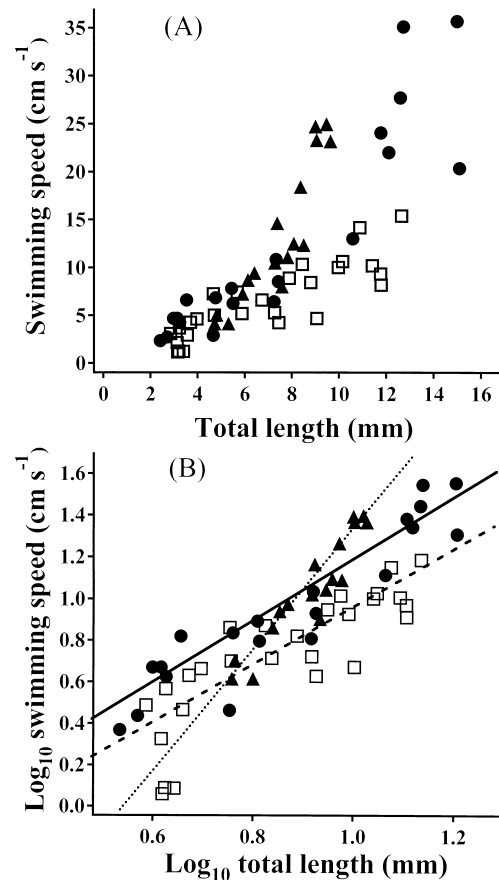


Fig. 5. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Mean critical swimming speed vs total length. (A) Raw values; (B) \log_{10} -transformed values

tained swimming ability was not recorded at Re values below 500 (Fig. 8). However, Re values at the time when sustained swimming ability first occurs differed for each species, with *Amphiprion melanopus* exhibit-

ing sustained swimming at Re values considerably lower than those for *P. amboinensis* (Fig. 8).

DISCUSSION

The 2 measures of swimming ability, critical and sustained, provide different estimates of swimming performance. Critical swimming speed provides a maximum performance measure of the speeds at which fishes can swim, as well as a means of directly comparing the development of swimming capabilities among species. In *Pomacentrus amboinensis*, *Sphaeramia nematoptera*, and *Amphiprion melanopus*, critical swimming speeds increased with age, size, developmental age and relative propulsive area. The development of critical swimming ability is probably based largely on the development of muscles and other features related to locomotion, as well as the ability to use readily available energy reserves. The biological implications of critical swimming speeds relate to the ability of the larvae to cover short distances (<500 m) at

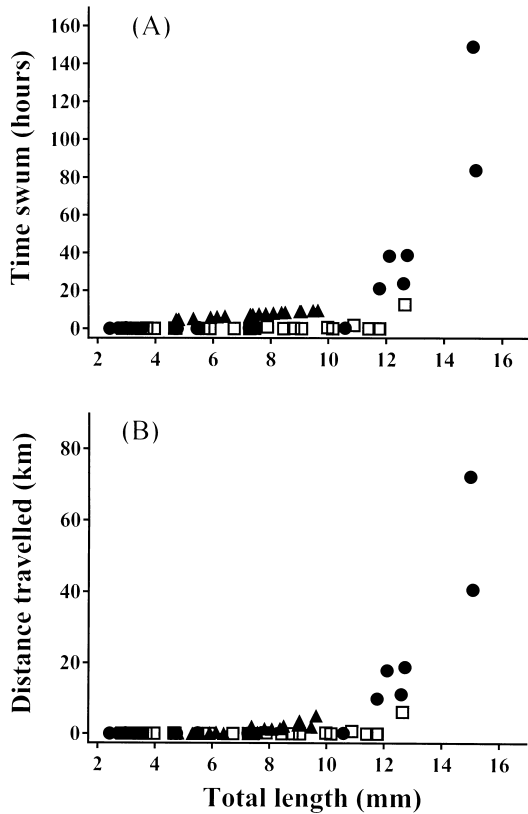


Fig. 6. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Mean sustained swimming ability vs total length. (A) Sustained swimming time in hours; (B) equivalent distance travelled

Table 2. *Pomacentrus amboinensis*, *Sphaeramia nematoptera*, *Amphiprion melanopus*. Partial correlation coefficients for multiple regression between critical swimming speed and sustained swimming time versus body area, body depth, and propulsive area. All variables were standardised relative to total length. Values in **bold** indicate morphological variables with the highest partial correlation coefficients for each species. Data were log₁₀-transformed prior to analysis

Species	Body area	Body depth	Propulsive area
Critical swimming speed			
<i>P. amboinensis</i>	0.22	-0.07	0.44
<i>S. nematoptera</i>	0.67	-0.45	0.17
<i>A. melanopus</i>	0.09	0.13	0.73
Sustained swimming time			
<i>P. amboinensis</i>	-0.10	-0.08	0.46
<i>S. nematoptera</i>	-0.05	0.08	0.21
<i>A. melanopus</i>	-0.32	-0.32	0.72

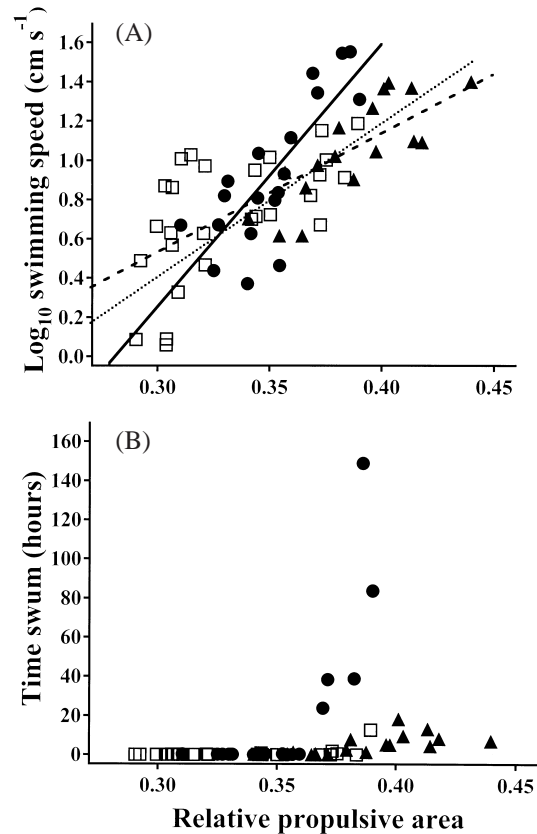


Fig. 7. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Swimming ability vs relative propulsive area. (A) Critical swimming speed; (B) sustained swimming time

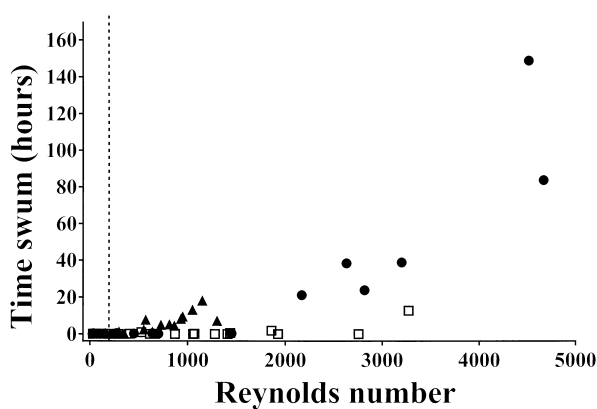


Fig. 8. *Pomacentrus amboinensis* (●), *Sphaeramia nematoptera* (□) and *Amphiprion melanopus* (▲). Sustained swimming time vs Reynolds number. Dotted line: $Re = 200$

high speeds (up to 48 cm s^{-1}), over short time scales (< 30 min). These abilities may be important in terms of the potential for larvae to move between locations on a small scale (e.g. within slicks), to vertically migrate to access different current regimes or prey, to move away from reefs at hatching, or to move between habitats at settlement. High abundances of larval fishes have been found in areas of internal waves, slicks and fronts, and this may be due to active migration of larvae to these areas, possibly because of increased food availability (Franks 1992). Critical swimming capabilities of larval fishes are likely to be important for such small-scale movement, and possibly in the modification of broad-scale dispersal patterns via vertical migration (see Job & Bellwood 2000).

In contrast to critical swimming speed, sustained swimming increased very differently among the species with both age and size, suggesting that the factors involved are more complex than those required for the development of critical swimming abilities. The development of sustained swimming ability varied from no development at all in *Sphaeramia nematoptera*, a fairly smooth transition in *Amphiprion melanopus*, to a rapid jump in *Pomacentrus amboinensis*. The development of sustained swimming may be dependent upon both physiological and behavioural changes during ontogeny. Sustained swimming experiments provide a measure of the long-term swimming abilities of larvae. However, this method may underestimate the swimming capabilities of larvae in the field (Stobutzki & Bellwood 1997). In the trials, larvae were not permitted to feed or rest and were unable to select their optimal swimming speeds. Leis & Carson-Ewart (1998) have observed larvae feeding whilst maintaining such high speeds. Given food, rest and the ability to select preferred swimming speeds, the swimming times might have been much higher. Although there is a great

deal of variation among species, this method provides a maximum performance measure of the sustained swimming capabilities of larvae, which may reflect relative abilities of larvae to modify dispersal patterns in the field.

Among-species differences — Is it possible to predict swimming ability across different species?

The development of both critical swimming speeds and sustained swimming times of larvae showed clear differences among the 3 species. Comparable differences in the swimming ability among taxa have been reported by other authors (Leis et al. 1996, Stobutzki & Bellwood 1997, Stobutzki 1998), and have been attributed to differences in developmental stage and morphology at the end of the pelagic stage (Stobutzki & Bellwood 1997). The differences observed in the present study appear to reflect the larval strategies of the 3 species. They hatch at varying stages of development and go through the larval period at different rates. However, if one considers the development of critical swimming speeds in terms of developmental age, which provides a way of comparing the species at similar stages of ontogenetic development (Job & Bellwood 2000), critical swimming ability increased similarly for all 3 species. Interestingly, Job & Bellwood describe a comparable pattern in the development of visual abilities of these species.

The differences between the swimming abilities of the 3 species can also be explained by the morphology of the larvae. All 3 species display a similar pattern, with critical swimming speeds increasing with increasing relative propulsive area. Other studies have reported that various measures of body muscle or propulsive area may, along with length, explain differences in swimming abilities. These include differences in the percentage of muscle relative to other body tissue (Bainbridge 1960) and the sagittal area of the body musculature (Ryland 1962). Furthermore, the length of the body that is undulated is known to vary among species (Jayne & Lauder 1995) and may explain differences in swimming ability. Propulsive area, as defined in the present study, approximates the area of the fish that can be undulated during swimming. This variable appears to give a good indication of the swimming capabilities of larval fishes, probably because it provides a measure of the development of the functional region of the fish used for swimming. If the relationship between relative propulsive area and swimming ability is similar in a number of additional species, it is possible that this variable may prove useful for general predictions of the swimming capabilities of larval fishes, including taxa which are difficult to rear in captivity.

Reynolds numbers (Re) have also been identified as an important factor in the development of swimming abilities in larval fishes (Webb & Weihs 1986). Re values give a measure of the ratio of forces arising from inertial and viscous effects as a fish moves through water. At small Re values (<200), the interaction between the fish and the water changes so that swimming becomes energetically expensive. Considering that all the species in this study did not develop sustained swimming until they had attained Re values considerably larger than 200, this may be a constraining factor only in the development of sustained swimming in very young larvae. It is unlikely that Re values are important for the development of sustained swimming ability, particularly as all 3 species developed sustained swimming at very different values of Re.

Implications for dispersal

The 3 species investigated in this study displayed quite different patterns of development of swimming ability. Although some species, such as *Sphaeramia nematoptera*, appear to exhibit very little swimming ability, and may be considered 'passive', others are clearly capable of exerting considerable influence over their dispersal patterns. The 2 species that did exhibit sustained swimming, however, developed these abilities considerably faster than expected. Critical swimming abilities for these species also developed rapidly. *Pomacentrus amboinensis* rapidly develops excellent sustained swimming abilities somewhere between Days 12 and 15 after hatching. Thus, for nearly half of the larval phase, this species is capable of significantly influencing its dispersal patterns by active swimming. This changes the potential distance that larvae of this species are able to cover before settlement from about 34 km (based on the ability to swim for the last 2 d of the pelagic period) to 93 km (based on the ability to swim for the last 6 d of the pelagic period, at a speed of 13.5 cm s^{-1}). These values may be considerably greater if we further assume that larvae are able to delay metamorphosis until they find a suitable settlement site (see McCormick 1999) and that they are able to feed whilst swimming (see Leis & Carson-Ewart 1998). *Amphiprion melanopus* also develops sustained swimming abilities earlier than expected, but differs from *P. amboinensis* in increasing its sustained swimming ability gradually with age. *A. melanopus* was capable of some sustained swimming starting between 3 and 5 d after hatching, and this ability increased consistently throughout the larval period. The development of significant swimming abilities relatively soon after hatching, along with well-developed sensory systems (Job & Bellwood 2000), and a very short larval duration

suggests that there is a high potential for this species to remain near to the natal reef. Preliminary results for *Amphiprion percula* (Fisher unpubl. data) indicated that the rapid development of swimming ability observed in *A. melanopus* may be common to other anemonefish species. These findings are consistent with the observation that there appears to be a high degree of genetic differentiation between some populations of anemonefish, an indication of limited dispersal (Bell et al. 1982). Job & Bellwood (2000) suggest that the precocious development in anemonefishes may be related to low post-settlement mortality rates and the need for well-developed sensory systems rather than size at settlement.

In at least 2 of the 3 species investigated, swimming abilities develop at an early enough age to substantially influence their dispersal patterns. If they use their swimming capabilities to modify dispersal, genetic evidence suggests that they are using them to retard rather than enhance dispersal (Doherty et al. 1995). There is considerable evidence from hydrodynamic studies suggesting that young larvae will not necessarily be transported very far from reefs (Black et al. 1991, Black 1993), particularly in the summer months (Frith et al. 1986). Local retention, along with the early development of swimming ability in some demersal-spawning reef fishes, suggests that many populations of reef fishes may have a significant potential for self-seeding¹.

The 3 species investigated in this study were all demersal brooders. Fish larvae which hatch from demersal eggs are more developed in terms of their sensory and swimming abilities than larvae which hatch from pelagic eggs (Kobayashi 1989, Leis 1991a), and are more likely to be able to remain near the reef of origin (Leis 1991b, Brogan 1994). Apparently anomalous distribution patterns have been observed in larvae with demersal eggs, with studies often reporting higher abundances near to shore (Leis 1986, Kingsford & Choat 1989). This may indicate that these larvae are using the early development of their motor skills to influence their dispersal patterns or that they are very adept at using currents to reduce advection. It is yet to be determined how swimming ability develops in pelagic spawners, but it is likely to follow a period of relative passivity during early development.

Larval fishes exhibit excellent swimming abilities that develop, especially for critical swimming ability, early in larval life and improve consistently throughout

¹Since submission of this paper 3 articles have been published highlighting the potential for self seeding in reef fishes. See Jones et al. (Nature 402:802–804, 1999), Swearer et al. (Nature 402:799–802, 1999) and Cowen et al. (Science 287: 857–860, 2000)

the larval period. However, swimming is only one aspect of larval biology that will enable them to exhibit behaviour potentially important in influencing dispersal. Job & Bellwood (2000) indicate that larvae of the study species are able to feed at considerable depths (50 to 200 m) even from an early age, and that this may provide access to a range of current regimes. The combination of prolonged access to deep water and the early development of swimming abilities suggest that even the youngest larvae of these taxa have considerable potential to actively modify their dispersal and reduce advection from reefs. Clearly, the potential of larvae to modify their dispersal is not restricted to the last few days of the pelagic period.

The present study has demonstrated that there are great differences in the development of swimming ability in the larvae of reef fishes and that these differences are apparent shortly after hatching. In order to fully understand the potential capabilities of larvae in modifying their dispersal, the whole of the larval period must be considered. Information based solely on the swimming abilities of older stages may be misleading because it does not take into account how quickly or slowly their relative swimming abilities develop during the pelagic period. Overall, it appears that there is no such thing as a typical fish larva at any stage and that some species of fish have considerably more potential for influencing their dispersal than others.

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