

COMMENT

Evaluating the 'reliability' of filtration rate measurements in bivalves

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The study of the physiology of bivalve suspension feeding has resulted in an extensive and diverse literature, but remains controversial as a result of divergent interpretations of the large body of data (Jørgensen 1990, 1996, Hawkins & Bayne 1992, Bayne 1993, Bayne 1998). This controversy has recently been heightened by Riisgård (2001) in a review of published filtration rate measurements in which he attempts to identify 'reliable' data within what he describes as 'heaps of conflicting data'. Whether or not these data are actually conflicting is central to the controversy. The wide range of filtration and clearance rate measurements reported in the literature are clearly in conflict with the view that bivalve feeding is a highly automated and unregulated process that greatly limits the range of potential filtration rate responses (Riisgård & Larsen 1995, Clausen & Riisgård 1996, Jørgensen 1996). However, others interpret the large variance in experimental and observed responses as evidence of direct physiological control linked to variability in the food supply (e.g. Hawkins & Bayne 1992, Bayne 1993, 1998). It is important for new readers of this literature to keep this underlying context in mind when following Riisgård's (Riisgård 2001) search for 'reliable' data.

Riisgård (2001) scrutinized the literature with the intent 'to point out some reliable (i.e. no methodological flaws) filtration rate data obtained under optimal laboratory conditions' in an attempt to dismiss what he calls 'erroneous interpretations'. A similar critical examination was undertaken by Jørgensen (1996). Although a large body of work has been called into question, I will confine my comments to Riisgård's criticisms of the reliability of my own data. For an insightful discussion on the merit of Jørgensen's critique, I refer readers to Bayne (1998). My reluctant foray into this debate extends primarily from the development of the *in situ* biodeposition approach for autonomously

and continuously measuring feeding and digestion processes in bivalves (Cranford & Hargrave 1994). This methodology was subsequently utilized to study short- (Cranford et al. 1998) and long-term (Cranford & Hill 1999) variations in bivalve feeding behaviour. The availability of *in situ* measurements on undisturbed bivalves exposed to natural dietary conditions was viewed as an opportunity to test clearance rate predictions based on a wide variety of available *in vivo* clearance rate models, including the widely applied models of Riisgård & Möhlenberg (1979) and Möhlenberg & Riisgård (1979). The observation of relevance here is that clearance rate predictions based on laboratory measurements using only algal cell diets (i.e. optimal laboratory conditions) were, on average, 320 to 1365% higher than the *in situ* measurements (Cranford & Hill 1999). This result confirmed similar observations reported in numerous cited studies, which collectively indicate that while bivalves in nature do fully exploit their clearance capacity for short periods (Cranford et al. 1998), they more often feed at a much reduced rate (Cranford & Hill 1999).

As for all experiments with living organisms, filtration and clearance rate measurements require close attention to the potential negative effects of experimental conditions (Jørgensen 1996). It was with these concerns in mind that the *in situ* biodeposition approach was applied. Riisgård (2001) suggests that the biodeposition method produces errors induced by pseudofaeces production and methods of diet characterization. These potential sources of error have been thoroughly and quantitatively addressed elsewhere (Cranford & Hargrave 1994, Cranford et al. 1998, Cranford & Hill 1999) and cannot explain the often large difference in clearance rates measured *in situ* and observed under optimal laboratory conditions. An important component of our work has been to carefully and objectively assess the assumptions of the method (Cranford et al. 1998) as well as determining if the estimates are 'reliable' (Cranford & Hill 1999). Precision

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and accuracy, defined as how much measurements deviate from the central and true values, respectively, are the primary criteria by which the 'reliability' of any data must be based. Riisgård (2001) has chosen not to present published data on the high precision and accuracy of our clearance rate estimates (Cranford & Hill 1999). We used the approach outlined in Clausen & Riisgård (1996) to assess measurement accuracy and showed that tissue growth predictions based on our clearance rate estimates were comparable with the observed growth of the animals used in the feeding experiments (Cranford & Hill 1999).

If data reliability is not based on an analysis of data precision and accuracy, how can it be judged objectively? The approach of Riisgård (2001) is to judge reliable data by searching for potential methodological pitfalls and flaws and by determining if the methods meet 'the necessary prerequisites'. In general, his examination and description of available methods is clear and insightful and will aid new researchers in selecting and properly applying many laboratory techniques for feeding experiments. However, his prerequisite that reliable filtration rate data must come from methodologies that provide results comparable to those measured for fully open bivalves under 'optimal laboratory conditions' (see Table 1 in Riisgård 2001) lacks scientific objectivity and is designed to exclude equally valid reasons for data variability. This prerequisite was used as the primary basis for questioning the reliability of any published filtration rates that were lower than he predicted, including many data provided by the biodeposition method. A suggested explanation for the observed low filtration rates was that 'sub-optimal conditions may be prevailing'. Considering that the definition of optimal conditions includes a carefully controlled laboratory diet consisting of an algal cell monoculture, optimal conditions can never prevail in nature. The prerequisite that measurements be performed on fully open bivalves is also questionable as *in situ* video observations show large natural temporal variations in shell gape (e.g. Newell et al. 1998), a measure that is correlated with filtration rate (Jørgensen et al. 1988).

After suggesting that the biodeposition method may be invalid, Riisgård used the relatively high clearance rates reported for sea scallops *Placopecten magellanicus* by Cranford et al. (1998) to question the reliability of lower values reported by MacDonald & Thompson (1986). While Riisgård is correct to point out that the geometry of flow-through chambers, such as used in the latter study, can provide erroneously low results, the potential for mixing within the chamber is easily tested and is routinely checked at each flow rate employed. A low clearance rate is not a valid reason in itself for concluding that a methodological problem

exists. For example, Riisgård implies that the relatively high clearance rates measured using flow-through chambers by Cranford & Gordon (1992) is evidence of a recirculation problem in the MacDonald & Thompson (1986) study. What he failed to mention was that Cranford & Gordon (1992) showed that these high rates immediately decreased by a factor of 3 when the algal cell diet was replaced with a natural seston diet. As the flow conditions in the chamber were identical for both diets, the latter result cannot be explained by potential methodological problems, but strongly supports the results of MacDonald & Thompson (1986), which were also based on a seston diet. Selective literature reviews and inconsistent interpretations of the validity of methodologies and results must always be viewed with caution.

Bivalve growth has been extensively studied but the relationship with environmental conditions is often poorly understood (Grant 1996). An understanding of the potential interplay between the physiological processes that limit food utilization and all potential exogenous and endogenous forcing functions is fundamental to the prediction of bivalve growth. Ample filtration rate data can be found in the literature that appear incompatible with any of the present theories of bivalve suspension feeding behaviour. The way to move forward is not to evaluate data reliability based on anticipated responses, but to focus attention on establishing standard and objective means of quantitatively addressing data accuracy under environmentally relevant experimental conditions. Research efforts to provide filtration rate measurements in the field have increased considerably in the last few years and we need to continue to encourage this effort rather than promoting the use of optimal laboratory conditions that constrain the potential for further insight.

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