Nitrogen-limited primary productivity of coral reef algal turfs: potential contribution of ammonium excreted by *Diadema antillarum* 

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ABSTRACT: Algal turfs account for the majority of the considerable primary productivity exhibited by coral reefs. Algal turfs grazed by the sea urchin *Diadema antillarum* Philippi are 2 to 10 times more productive per unit chlorophyll a than turfs not grazed by *D. antillarum*. We investigated one of several hypotheses to account for this effect: primary productivity of algal turfs is nutrient-limited and excretion by *D. antillarum* provides a nitrogen supplement to algal turfs. Algal turfs on natural settling plates were fertilized with ammonium, phosphorus, or both, for 17 d on a backreef in St Croix, US Virgin Islands. Primary productivity per unit chlorophyll a was significantly greater in the ammonium treatment than in the control treatment, although algal biomass in each treatment was not significantly different from the control. Biomass-specific ammonium excretion by *D. antillarum* was correlated with sea urchin size and respiration rates and averaged 115 µg N ind⁻¹ h⁻¹. Excretion was significantly greater during the day than at night. In the field, ammonium concentrations were significantly higher under *D. antillarum* than ambient concentrations, indicating that dissipation of sea urchin excretions was not immediate or complete. Calculations of the nitrogen required for the observed primary productivity of algal turfs indicate that, prior to the mass mortality, up to 19% could have been supplied by *D. antillarum* excretions.

INTRODUCTION

Coral reefs exhibit rates of primary productivity that are among the highest reported for any ecosystem (Lewis 1977). Algal turfs are responsible for the majority of whole reef primary productivity (Johannes et al. 1971, Marsh 1976, Wanders 1976, Hatcher 1981, Carpenter 1985a) due to the high coverage of substratum by this component and very high biomass-specific productivity rates [e.g. 6.4 to 8.0 µg O₂ (µg chl a)⁻¹ h⁻¹; Carpenter 1985a]. Algal turfs are densely packed assemblages of unicellular and filamentous algae from at least 5 algal divisions and are characterized by diminutive canopy heights (generally <5 mm; Wanders 1977, Steneck & Watling 1982).

Numerous studies have demonstrated the widespread importance of grazing by the sea urchin *Diadema antillarum* Philippi on benthic algal commun-

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Diadema antillarum at natural densities of 6 sea urchins m\(^{-2}\) (Ogden 1977, Carpenter 1986, Foster 1987) to achieve fairly uniform algal turf biomass among the plates. After 15 d, 12 plates were selected randomly to estimate the light-saturated rate of O\(_2\) production in acrylic (Plexiglas) chambers. Duplicate incubations (15 min) were conducted and averaged for each of the 12 plates. The methodology for primary productivity estimates, algal biomass, and chlorophyll \(a\) (chl \(a\)) followed Carpenter (1985a).

Six additional plates were assigned randomly to each of 4 treatments: (1) unfertilized control; (2) fertilized with ammonium (+N); (3) fertilized with phosphorus (+P); (4) fertilized with ammonium and phosphorus (+N +P). Plates were affixed to aluminum strips which rested on cement blocks ca 30 cm above the sediments. Nutrient-enrichment treatments were located downstream of the control treatment and were assigned randomly to positions 2 m apart on a transect perpendicular to the normal net unidirectional water flow to minimize cross-fertilization of the treatments.

Nutrients were released slowly from 250 ml polycarbonate bottles drilled with several small holes and filled with 165 g NH\(_4\)Cl or 60 g KH\(_2\)PO\(_4\), or both, contained in nylon mesh bags. Two bottles were placed in each treatment at the level of the algal canopy and the fertilizer was replaced every 6 d. Concentrations of ammonium (Koroleff 1976) and phosphorus (Parsons et al. 1984) were determined in duplicate daily for 6 d for each treatment and averaged over the time between successive replacements of nutrient enrichments. After 17 d, rates of primary productivity of algal turfs were estimated. Pairs of plates (1 per chamber) from each treatment were incubated twice for 15 min and the rates per plate were averaged. The mean rate of the 2 plates was used as the primary productivity estimate. To detect temporal changes over 17 d, algal biomass in the control treatment was compared to the initial biomass using a 2-tailed t-test. We tested our hypothesis that nutrient enrichment would not stimulate primary productivity using paired t-tests between the control and each treatment.

Algal tissue was removed for carbon and nitrogen content from randomly selected 1 cm\(^2\) areas on 5 plates grazed by Diadema antillarum and 5 plates not grazed by sea urchins. Carbonates were removed from the samples with 1% HCl followed by a rinse with deionized water. Decalcified samples were filtered onto pre-combusted glass fiber filters, and dried at 60°C for 24 h. Carbon and nitrogen content were determined in a Perkin-Elmer 240B elemental analyzer using acetonitrile as a standard. The nitrogen required to support the measured O\(_2\) production was calculated from the C:N content assuming a 10 h light period and a photosynthetic quotient of 1.0.

Respiration and ammonium excretion by Diadema antillarum may enhance nutrient supply to algal turfs in several ways. For example, grazing may favor growth of cyanobacteria (blue-green algae) in the algal turf community and result in increased rates of N\(_2\) fixation per unit area, similar to the effects of fish grazing (Wilkinson & Sammarco 1983, Wilkinson et al. 1984). Also, soluble excretions from D. antillarum may provide a nutrient supplement to algal turfs. Ammonium comprises >60% of the nitrogenous excretions of D. antillarum (Lewis 1967) which exit from the respiratory surfaces (tube feet, gills). If water exchange with the surroundings is reduced under sea urchins, nitrogenous excretions may remain available for uptake by algae. Nitrogen-fixing bacteria have been isolated from the gut of D. antillarum (Guerinot & Patriquin 1981); thus, excretions may represent a source of both recycled and new nitrogen (sensu Dugdale 1976).

We present results from an experiment designed to test the hypothesis that the primary productivity of coral reef algal turfs is nutrient-limited, and, if it is, whether nitrogen or phosphorus is limiting. In addition, we assess the contribution that ammonium excretions from Diadema antillarum may make to algal turf primary productivity.

**METHODS**

Fertilization effects on algal turf primary productivity. Algal turfs were grown on settling plates (8 x 8 x 1 cm) cut from dead branches of the reef coral Acropora palmata (Carpenter 1985a). Plates were placed at a depth of 1.5 to 2 m on the backreef of Tague Bay, St Croix, US Virgin Islands, for 10 mo prior to the fertilization experiments. This period is more than sufficient for turfs on the settling plates to resemble those on the surrounding natural substrata (Carpenter 1986). Eighteen plates then were placed in cages with Diadema antillarum at natural densities of 6 sea urchins m\(^{-2}\) (Carpenter 1985b), which may minimize algal self-shading and/or facilitate diffusion of carbon and nutrients to the plants. Other hypotheses concern possible nutrient limitation of algal turfs and the role of D. antillarum in enhancing nutrient input to algal turfs.

It is currently under debate whether the primary productivity of coral reefs is nutrient-limited and, if so, what nutrient is limiting (Entsch et al. 1973, Smith 1984, Gladfelter & Kinsey 1985). Increased rates of whole reef primary productivity have resulted from experimental enrichment of a reef with nitrogen and phosphorus, suggesting that reef algae are nutrient-limited (Kinsey & Domm 1974). Following nitrogen enrichment of another reef, algal turf community structure changed, suggesting nitrogen limitation of at least the algal turf component (Hatcher & Larkum 1983).

Diadema antillarum may enhance nutrient supply to algal turfs in several ways. For example, grazing may favor growth of cyanobacteria (blue-green algae) in the algal turf community and result in increased rates of N\(_2\) fixation per unit area, similar to the effects of fish grazing (Wilkinson & Sammarco 1983, Wilkinson et al. 1984). Also, soluble excretions from D. antillarum may provide a nutrient supplement to algal turfs. Ammonium comprises >60% of the nitrogenous excretions of D. antillarum (Lewis 1967) which exit from the respiratory surfaces (tube feet, gills). If water exchange with the surroundings is reduced under sea urchins, nitrogenous excretions may remain available for uptake by algae. Nitrogen-fixing bacteria have been isolated from the gut of D. antillarum (Guerinot & Patriquin 1981); thus, excretions may represent a source of both recycled and new nitrogen (sensu Dugdale 1976).
antillarum. *Diadema antillarum* were collected from a patch reef and the fore reef at 3 to 5 m depth from 07:00 to 09:00 h local time. The sea urchins were placed in seawater in buckets and immediately returned to the laboratory where they were transferred to tables with flowing seawater.

Experiments began after a minimum acclimation period of 2 h. The maximum test diameter (MTD) of *Diadema antillarum* was measured to the nearest millimeter using long-jawed calipers and 1 sea urchin was placed in each of 2 aquaria (22 l volume). A third aquarium served as control. *D. antillarum* were not fed during incubations, although they had access to algal turfs during the acclimation period. Lids fitted with an oxygen probe/stirrer combination (Yellow Springs Instruments) and water sampling ports were sealed over the aquaria with unvulcanized rubber. The aquaria were placed in tables with flowing seawater shaded from direct sunlight. Temperature inside the aquaria was maintained at 26 ± 2°C. Changes in dissolved O2 were recorded every 30 min and water for ammonium and nitrate plus nitrite analyses (Parsons et al. 1984) was withdrawn in triplicate every 2 h for 8 to 10 h beginning at 08:00 to 10:00 h local time or at 20:00 to 21:00 h for dark experiments. Water was replaced simultaneously through a second sampling port.

Respiration rates of 38 sea urchins were estimated from the decline in dissolved O2 concentration over time. During initial experiments, activity patterns of sea urchins were observed for correlation with respiration rates. Activity was ranked on a scale from 1 to 3, corresponding to sea urchins climbing aquarium walls (1), moving along the bottom of the aquarium (2), and not moving except for spines (3). Biomass-specific respiration rates were calculated based on the equation:

\[
Y = -0.41198 + 0.057108 X
\]

where \(X = \text{MTD}\), \(Y = \text{g dry weight of soft and hard parts of } D. \text{antillarum} \) (Carpenter unpubl.).

Ammonium excretion rates of 40 sea urchins were calculated as the slope of a best-fit regression between ammonium concentration and time. If the change in ammonium concentration over time in the control was significant \((p < 0.05; 1\text{-way analysis of variance})\), the excretion rate was corrected for the control rate.

Ammonium concentrations under *Diadema antillarum* resting during the day in the open or in crevices open to light were estimated in water samples taken from under *D. antillarum* with a modified syringe. The needle was placed at the end of a polycarbonate tube that extended 40 cm from the syringe attachment point and was held rigid by a thin metal rod. Each sample under a sea urchin was paired with a control sample taken with a separate syringe above an algal turf, >20 cm away from a sea urchin. Samples were fixed immediately with reagents and shielded from light in a cooler before being returned to the laboratory for analyses.

**RESULTS**

Effects of nutrient enrichment on primary productivity of algal turfs

Ammonium and phosphate concentrations were substantially elevated over control levels in the nutrient-enrichment treatments (Table 1). There were no significant differences in biomass or rates of primary productivity of algal turfs on the plates at the start of the nutrient-enrichment experiment \((p > 0.05; 1\text{-way analysis of variance})\). The algal biomass of the control plates did not change significantly from the original biomass after 17 d \((p > 0.05)\) and was representative of previously reported values (Carpenter 1986). Algal bio-

<table>
<thead>
<tr>
<th>Ammonium (a) (µM)</th>
<th>Control (n)</th>
<th>+ N</th>
<th>+ P</th>
<th>+ N + P</th>
</tr>
</thead>
<tbody>
<tr>
<td>12</td>
<td>0 ± 0</td>
<td>1.46 ± 2.41</td>
<td>0 ± 0</td>
<td>1.50 ± 1.45</td>
</tr>
<tr>
<td>Phosphorus (b) (µM)</td>
<td>12</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>0.27 ± 0.15</td>
</tr>
<tr>
<td>Algal biomass mg dry cm(^{-2})</td>
<td>3</td>
<td>1.8 ± 0.7</td>
<td>2.0 ± 0.7(^*)</td>
<td>2.6 ± 0.9(^*)</td>
</tr>
<tr>
<td>Ch l a cm(^{-2})</td>
<td>3</td>
<td>4.2 ± 0.6</td>
<td>2.7 ± 0.4(^*)</td>
<td>3.9 ± 1.4(^*)</td>
</tr>
<tr>
<td>Net productivity µg O(_2)/µg ch l a(^{-1}) h(^{-1})</td>
<td>3</td>
<td>7.54 ± 1.07</td>
<td>13.47 ± 2.41(^*)</td>
<td>10.63 ± 6.02(^*)</td>
</tr>
<tr>
<td>g O(_2)/m(^{-2}) h(^{-1})</td>
<td>3</td>
<td>0.31 ± 0.01</td>
<td>0.36 ± 0.05(^*)</td>
<td>0.35 ± 0.03(^*)</td>
</tr>
</tbody>
</table>

\(a\) Detection limit = 0.1 µM; \(b\) detection limit = 0.03 µM
\(^*\) Not significant at \(p > 0.05\); \(*\) significant at \(p < 0.05\); \(**\) significant at \(p < 0.01\)
mass in each treatment was not significantly different from the control after 17 d \( (p > 0.01; \text{Table 1}) \), a time substantially greater than the turnover time of \( \geq 1 \) d for algal turfs (Carpenter 1986). Net primary productivity per unit chl \( a \) was significantly greater \( (p < 0.05) \) in the +N treatment than in the control treatment. Rates of primary productivity in each of the remaining treatments did not differ significantly from the control \( (p > 0.05) \). Rates of primary productivity per unit area were stimulated significantly only in the +N+P treatment \( (p < 0.05) \), suggesting a possible synergistic effect of N and P. In this treatment, chl \( a \) cm\(^{-2} \) was higher, although algal biomass was similar to other treatments, indicating an increased concentration of chl \( a \) in the algal tissue. These results suggest that chlorophyll-specific primary productivity was depressed in the +N+P treatment or that a portion of the chl \( a \) was inactive in photosynthesis.

Respiration and ammonium excretion by *Diadema antillarum*

The decline of \( O_2 \) in aquaria with sea urchins was always linear, indicating that disturbance of *Diadema antillarum* had no observable effect on their rates of respiration during the experiments (Fig. 1). Respiration rates ranged from 2.4 to 8.6 mg \( O_2 \) ind.\(^{-1} \) h\(^{-1} \) and were not significantly different between day \( (4.47 \text{ mg } O_2 \text{ ind.}^{-1} \text{ h}^{-1} \pm 1.48 \text{ SD}, n = 28) \) and night \( (3.93 \text{ mg } O_2 \text{ ind.}^{-1} \text{ h}^{-1}) \). Respiration rates increased with *D. antillarum* size in the range from 49 to 90 mm MTD (Fig. 2); however, biomass-specific respiration rates declined with increasing sea urchin biomass (Fig. 3).

![Fig. 1. Example of changes in dissolved \( O_2 \) over time in aquaria with and without (control) *Diadema antillarum*. m: slope of least squares regression line; r: regression coefficient. Activity of *D. antillarum* is indicated at 0.5 h intervals: 1 = climbing walls, 2 = moving over aquarium floor, 3 = spine movement only.](image1.png)

![Fig. 2. *Diadema antillarum*. Respiration as a function of maximum test diameter. r: correlation coefficient.](image2.png)

![Fig. 3. *Diadema antillarum*. Respiration rates as a function of sea urchin dry weight. r: correlation coefficient.](image3.png)

A typical time course used to calculate ammonium excretion is shown in Fig. 4. Mean ammonium excretion rates were significantly higher during the day \( (141 \mu g \text{ N ind.}^{-1} \text{ h}^{-1} \pm 84 \text{ SD}, n = 30) \) than at night \( (38 \mu g \text{ N ind.}^{-1} \text{ h}^{-1} \pm 18 \text{ SD}, n = 10) \) \( (p < 0.05; \text{2-tailed } t\text{-test}) \). Ca 79% of the estimated daily excretion occurred during the day when *Diadema antillarum* is relatively inactive but in proximity to algal turfs. Biomass-specific rates of ammonium excretion were negatively
correlated with sea urchin biomass ($r = 0.42$, $p < 0.075$). Partial correlation analysis, with sea urchin biomass held constant, indicated that biomass-specific excretion rates were positively correlated with biomass-specific respiration rates ($r = 0.88$, $p < 0.01$).

Changes in ammonium concentration in the control were relatively small ($0.2 \pm 1.1 \mu g \, N \, l^{-1} \, h^{-1}$). The mean nitrate-plus-nitrite production rate of $0.4 \pm 0.9 \mu g \, N \, l^{-1} \, h^{-1}$ in aquaria with *D. antillarum* was only 1% of the total combined production of dissolved inorganic nitrogen (Fig. 5). There were no consistent patterns between changes in ammonium and nitrate-plus-nitrite concentrations, suggesting that there were no systematic, and presumably microbially-mediated, transformations of dissolved inorganic nitrogen (e.g. nitrification).

In the field, dissipation of *Diadema antillarum* excretions was not immediate or complete, suggesting that nitrogenous excretions are likely to be available for uptake by algae. Mean ammonium concentrations of $2.28 \mu M \pm 1.40 \, SD$ under *D. antillarum* were significantly higher than adjacent ambient concentrations of $1.04 \mu M \pm 1.03 \, SD$ ($p < 0.025$, paired 1-tailed t-test). The ambient values reflected contamination of syringes and glassware during the 1 h period of sampling from a small open boat. Ammonium was undetectable when sampled with a clean bottle and was also undetectable in the initial control sample, but each successive pair of ambient and sea urchin samples contained more ammonium. Because paired comparisons were made, the relative differences between ambient concentrations and those under sea urchins were unaffected. The increase of $1.24 \mu M$ attributable to the presence of *D. antillarum* was similar to the average ammonium concentration in the nutrient enrichment experiment (Table 1).

**Potential contribution of *Diadema antillarum* excretion to primary productivity of algal turfs**

Based on the above evidence that in nature *Diadema antillarum* represent microenvironments of elevated ammonium concentrations and that algal turf primary productivity is nitrogen-limited, we calculated the potential contribution of ammonium excretion by *D. antillarum* to the gross primary productivity of algal turfs. We used pre- and post-mortality population densities of *D. antillarum* (Carpenter 1985b) and the mean day (= 10 h) plus night excretion rates. We did not use size-specific excretion rates because *D. antillarum* size-class frequencies were not available. The mean C:N content of sea urchin-grazed algal tissue ($15.0:1 \pm 1.8:1 \, SD$) for the pre-mortality conditions and that of algal tissue not grazed by sea urchins ($11.4:1 \pm 1.6:1 \, SD$) for post-mortality, were used to calculate algal turf nitrogen requirements. Ammonium excretion from *D. antillarum* could supply 6 to 19% of the estimated nitrogen required for pre-mortality algal turf primary productivity but < 1% of post-mortality primary productivity (Table 2).
Table 2. Percentages of the nitrogen requirement of algal turfs that are supplied by *Diadema antillarum* excretions, based on pre- and post-mortality population densities, gross primary productivity, and C:N ratios of algal turfs

<table>
<thead>
<tr>
<th>Site</th>
<th>Water depth [m]</th>
<th>Diadema density (no. m⁻²)</th>
<th>Primary productivity [g C m⁻² d⁻¹]</th>
<th>Nitrogen requirement [mg N m⁻² d⁻¹]</th>
<th>Nitrogen provided [mg N m⁻² d⁻¹]</th>
<th>N supplied by excretion [%]</th>
</tr>
</thead>
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<tr>
<td></td>
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<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pre-mortality (December 1983)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Backreef</td>
<td>2</td>
<td>6.4</td>
<td>3.18</td>
<td>212</td>
<td>12</td>
<td>6</td>
</tr>
<tr>
<td>Forereef</td>
<td>2</td>
<td>13.4</td>
<td>2.02</td>
<td>135</td>
<td>26</td>
<td>19</td>
</tr>
<tr>
<td>Forereef</td>
<td>5</td>
<td>9.0</td>
<td>1.70</td>
<td>113</td>
<td>17</td>
<td>15</td>
</tr>
<tr>
<td>Forereef</td>
<td>10</td>
<td>5.8</td>
<td>1.13</td>
<td>97</td>
<td>11</td>
<td>13</td>
</tr>
<tr>
<td>Post-mortality (December 1985)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Backreef</td>
<td>2</td>
<td>0.1</td>
<td>2.32</td>
<td>205</td>
<td>0.2</td>
<td>&lt;1</td>
</tr>
<tr>
<td>Forereef</td>
<td>2</td>
<td>1.1</td>
<td>2.18</td>
<td>246</td>
<td>2</td>
<td>&lt;1</td>
</tr>
<tr>
<td>Forereef</td>
<td>5</td>
<td>0.3</td>
<td>2.36</td>
<td>207</td>
<td>0.6</td>
<td>&lt;1</td>
</tr>
<tr>
<td>Forereef</td>
<td>10</td>
<td>0.3</td>
<td>1.84</td>
<td>161</td>
<td>0.6</td>
<td>&lt;1</td>
</tr>
</tbody>
</table>

a Data from Carpenter (1985a, b, 1988)
b C:N of algal turf = 15:1 (pre-mortality), 11.4:1 (post-mortality)
c Based on mean day (=10 h) plus night excretory rates

**DISCUSSION**

Few data on sea urchin respiration, and fewer on excretion, are available (for review, see Jangoux & Lawrence 1982). Our main purpose in measuring respiration was to verify that *Diadema antillarum* metabolism was stable during measurement of excretion. Respiration rates of *D. antillarum* in our study are within the upper range of values reported for this and other Caribbean regular echinoids (Lewis 1968a, b, Jangoux 1982, Lawrence & Lane 1982). Ammonium excretion rates are also similar to those previously reported for *D. antillarum* of a similar size class (Lewis 1967, Hawkins & Lewis 1982). Respiration and excretion rates on a biomass-specific basis are higher for smaller *D. antillarum*, consistent with patterns between size and metabolic activity demonstrated for most organisms (Schmidt-Nielsen 1984).

Higher rates of excretion during the day are not correlated with higher respiration rates but probably are related to the nocturnal feeding activity and diurnal digestion of *D. antillarum*. Food first appears in the hind gut or intestine, where absorption probably occurs (Lewis 1964, Lawrence 1982), 4 h after ingestion (Lewis 1964). Food passes through the gut in 8 to 12 h (Lewis 1964), suggesting that the majority of digestion occurs during the day in *D. antillarum*. This most likely explains the concomitant higher diurnal rates of excretion. An alternative explanation may be that the sea urchins did not graze the algal turfs provided in the laboratory before the night experiments commenced and had empty guts. *D. antillarum* rest in proximity to algal turfs during the day and cover only 1 to 2 m² at night when they sit for periods probably sufficient to provide a buildup of ammonium similar to that measured (Carpenter 1984, pers. obs.). Multicellular marine algae take up ammonium in the dark at a rate that may or may not be lower than in the light, depending on the species and length of the dark period (Haines & Wheeler 1978, Hanisak & Harlin 1978, Ryther et al. 1981). Data on kinetics of nutrient uptake by coral reef algal turfs are presently unavailable.

The input of nutrients to plant communities that is mediated by animals is increasingly recognized as an important ecological feature of many plant-plant interactions. Stimulation of primary productivity of grazed plants by herbivore-regulated nutrient inputs has been documented in terrestrial and aquatic ecosystems (Flint & Goldman 1975, McNaughton 1979, Seale 1980, Newbold et al. 1982, Carpenter & Kitchell 1984, Sterner 1986). Marine primary producers, such as the giant kelp *Macrocystis pyrifera* and the coral *Acropora palmata*, that provide resting areas for fishes benefit from nutrients that the fishes obtain during migrations to adjacent feeding areas (Bray et al. 1981, Meyer et al. 1983). Nitrogenous excretions from fauna associated with temperate seaweeds can provide a critical source of nitrogen for seaweed growth when ambient nitrogen is depleted (Kautsky & Wallentinus 1980, Probyn & Chapman 1983). Nitrogenous excretions from *Diadema antillarum* may provide an example of a herbivore-plant nutrient transfer that fosters high plant productivity in a nutrient-poor environment.

We have demonstrated that primary productivity of algal turfs is limited by nitrogen (Table 1). Potential sources of nitrogen for reef primary productivity are
advection, N₂ fixation, and recycling. To the extent that recycled nitrogen is used, more new nitrogen (sensu Dugdale 1976) will be available for net algal production. Ammonium excreted by Diadema antillarum is available for algal uptake and may potentially supply up to 19% of the total nitrogen requirements of algal turfs (Table 2). Although it has not been demonstrated that excreted ammonium is taken up by algal turf species, it is reasonable to assume rapid uptake of ammonium when it becomes available in a nitrogen-limited environment (Rosenberg & Probyn 1984, Thomas & Harrison 1987). This nitrogen supplement may explain partly why algal turfs grazed by D. antillarum are more productive per unit chl a than algal turfs not grazed by D. antillarum. Grazing by herbottomous fishes, the other dominant functional group of herbivores on algal turfs (Carpenter 1986, Lewis 1986), does not stimulate the chl a-specific productivity of algal turfs. One obvious difference between herbottomous fishes and D. antillarum is that D. antillarum excretes in much closer proximity to algal turfs. Although recycled nitrogen may not increase the amount of net production available for export from reef ecosystems, it is another mechanism that maintains the high rates of primary productivity of algal turfs on coral reefs. The relative amounts of nitrogen supplied to algal turfs by advection, N₂ fixation, and recycling remain to be determined.

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LITERATURE CITED


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