

Significance of Temperature to Larval Survival and Length of Development in *Balanus eburneus* (Crustacea: Cirripedia)* **

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ABSTRACT: Mortality and length of development of *Balanus eburneus* nauplii is modified in laboratory culture by both temperature and concentration of unicellular algal food. At 20 and 25°C mean larval survival was 63.5% and 66.0%, respectively, at an algal concentration of 1×10^5 cells ml^{-1} . When the algal concentration was doubled to 2×10^5 cells ml^{-1} mean survival increased significantly to 92.5 and 96.5% at 20 and 25°C, respectively ($p = 0.002$ at both temperatures). At 30°C the difference in mortality related to algal food concentration disappeared and mean survival was above 90% at both concentrations. Over the interval of 20 to 30°C, time to complete development to the cyprid stage decreased from a median of 7.6 to 3.9 d at 2×10^5 cells ml^{-1} and from 10.8 to 3.9 d at the lower food level of 1×10^5 cells ml^{-1} . At 30°C there was no difference between the 2 food concentrations in the median time to reach the cyprid stage. The hypothesis is proposed that increased algal cell concentration at 20 and 25°C compensates for lower feeding efficiency and consequently results in better survival. To account for the similarity in development time at 30°C at either food concentration, it is proposed that maximum possible food intake at 30°C is already attained at 1×10^5 cells ml^{-1} and that therefore increase in the number of algal cells will not further affect development rate to the cyprid stage. Experiments on feeding efficiency of *B. eburneus* nauplii in relation to temperature will be needed to test this hypothesis.

INTRODUCTION

The research described here undertakes to show by laboratory experiments how differences in seawater temperature may affect mortality and length of planktonic development of the 6 nauplius larvae belonging to the common estuarine barnacle *Balanus eburneus* Gould, a species indigenous to the region between Massachusetts (USA) and northeastern South America and subsequently introduced into many other parts of the world including the Mediterranean Sea, Black Sea and various areas of the Pacific Ocean (Henry and McLaughlin, 1975, p. 60).

Even though temperature is known to be a factor regulating mortality and length of planktonic larval life in a variety of marine invertebrates, only few previously published studies consider its role through all 6 naupliar stages. Hirano (1962) in a laboratory experiment showed for *Balanus amphitrite hawaiiensis* the relationship between water temperature and

length of development to the cyprid stage. Tighe-Ford et al. (1970) considered the effect of temperature on mortality and the time required for complete development through the 6 naupliar stages of *Elminius modestus*, but presented only few quantitative data. The remaining published accounts dealing with the role of temperature on the development of nauplii are restricted to experiments using either the embryos and first nauplii readily obtained from breeding adults or alternatively late larval stages taken from the plankton (e.g. Barnes and Barnes, 1958; Crisp and Ritz, 1967). In such studies the intermediate naupliar stages were not considered.

* This paper is dedicated to the memory of Dr Harold Barnes who made so many important contributions to the ecology and physiology of Cirripedia

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METHODS

The larvae of *Balanus eburneus* were first reared and described by Costlow and Bookhout (1957). Subsequently they have been grown with varying success in a number of other laboratories (Freiberger and Cologer, 1966; Landau and d'Agostino, 1977, 1978).

The earliest stages of barnacle development occur within the ovigerous lamellae of the parent, and gravid adults collected during the breeding season readily release first stage nauplii when held in 4-l jars. The short first nauplius stage does not feed; the remaining 5 planktonic nauplii are phytoplanktonic and develop when fed upon a diet of unicellular algae. The final free-swimming cyprid is a non-feeding larva which settles and subsequently attaches onto a hard surface.

Several unicellular algae grown in single-species cultures were initially tried as food for the nauplii of *Balanus eburneus*. The algal species tried included *Thalassiosira pseudonana* (W.H.O.I.*, Clone 3H = *Cyclotella nana* used by Freiberger and Cologer, 1966), *Skeletonema costatum*, *Isochrysis galbana*, *Pavlova lutheri* (= *Monochrysis lutheri*), *Dunaliella tertiolecta* and *Platymonas* sp. (W.H.O.I., Clone 1). Differences were found in the survival and growth of nauplii with each species offered. For example, in an experiment using *Dunaliella tertiolecta* at 25°C at either 1×10^5 or 2×10^5 cells ml⁻¹, survival to the cyprid stage was only 8 to 10%, whereas *T. pseudonana* at a similar temperature and concentration gave a survival of 36 and 80%, respectively, even though cells of *D. tertiolecta* have on average twice the volume of *T. pseudonana* (i.e. 400 μm³ and 203 μm³, respectively).

Combinations of more than one algal species consistently resulted in greater survival and faster development of nauplii than when any single species was used as food. Combinations of *Thalassiosira pseudonana* at a ratio of 1:1 with either *Dunaliella tertiolecta*, *Pavlova lutheri* or *Platymonas* sp. gave uniformly high survival approaching 90% or better. Cultures of nauplii offered 3 species of algae, e.g. *P. lutheri*, *Platymonas* sp. and *T. pseudonana* at ratios of 1:1:1 or 1:1:2, respectively, and a total combined concentration of 2×10^5 cells ml⁻¹ also showed very high survival. Enhancement of growth and survival when more than one algal species is used as food also has been shown for bivalve veligers (Davis and Guillard, 1958) and a number of other phytoplanktonic invertebrate larvae. However, it has not been our purpose here to define the precise relative food value of the different algal species or combination of algal

species (*vide* Moyses, 1963; Lang, 1977), but rather to find a source of food that will sustain the five successive feeding naupliar instars to the cyprid stage.

Initially wide variations in survival and growth of larvae resulted from one culture to the next, even when the same species of alga were used as food. Apparently such variations were not related to differences in the condition or 'quality' of the alga used (e.g. age of the algal culture or growth medium used) but rather to differences between different broods of larvae. Progeny from different parents show larger variations in survival than siblings from the same parent even when all cultures are grown simultaneously under identical conditions. As much as a 35% difference in survival occurred between larvae arising from different parents from the same population.

To minimize the variables inherent in our experiments we established the following prerequisites: (1) Larvae were always siblings from the same parents (some may be 'half' siblings since it is not known whether or not the parents were promiscuous); (2) experiments at all temperatures tested were conducted concurrently using algal food from the same cultures; (3) algal food organisms with wide spectra of temperature tolerance were selected since we wished to know the direct effect of temperature upon the larvae themselves and not the indirect effects resulting from the effect of temperature upon the algal food (Scheltema 1967, p. 262).

The experimental procedure was as follows. Nauplii were grown in 100 ml of 25 ppt seawater in covered dishes 9 cm in diameter and 5 cm high and fed from unialgal cultures of *Pavlova lutheri*, *Platymonas* sp. and *Thalassiosira pseudonana* at ratios of 1:1:1 and 1:1:2 and at total concentrations of 1×10^5 and 2×10^5 cells ml⁻¹. The larvae were obtained from adult *Balanus eburneus* found in Eel Pond at Woods Hole and in nearby Green Pond in the town of Falmouth, Massachusetts, USA. Replicate cultures of 50 nauplii obtained from a single gravid *B. eburneus* were grown in cabinets at 20, 25 and 30°C. Hence, siblings (or half siblings) fed from the same algal cultures were compared in replicate at 3 different temperatures spanning the range within which nauplii larvae normally occur during development in the region of Cape Cod (Fish, 1925). Larvae were examined daily to determine survival. The surviving nauplii were transferred to clean culture dishes with fresh seawater and algae. Cultures were maintained at a 12 h light-dark cycle.

RESULTS

Survival

The effect of temperature on the successful completion of development to the cyprid stage is summarized

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in Table 1 and shown graphically in Fig. 1 where the percent survival of sibling (or half sibling) nauplii is given for 3 different temperatures, 20, 25 and 30°C and at 2 concentrations of algal food, 1×10^5 and 2×10^5 cells ml⁻¹. At the greater of the 2 algal concentrations (i.e. 2×10^5 cells ml⁻¹) there were no significant differences in percent mortality among the 3 different temperatures ($p > 0.50$); survival was uniformly high, ranging from 88 to 100%. At the lower algal concentration there was, however, a significant difference in survival correlated with temperature. At 25°C a mean survival of 66.0% (range 60 to 72%) was significantly less than 89.5% (range 84 to 94%) found at 30°C ($p = 0.01$). A comparison between cultures grown at the higher and lower algal concentrations show significant differences at 20 and 25°C ($p < 0.002$ in both instances). When reared at 1×10^5 cells ml⁻¹, survivorship of larvae in the experiments averaged 29% less at 20°C (range 22 to 34%) and 30.5% less at 25°C (range 28 to 36%) than when grown at 2×10^5 cells ml⁻¹. Such a difference disappeared at 30°C at which temperature mortality was never greater than 16% at either algal concentration.

Nauplii did not consistently reach the cyprid stage at algal concentrations of less than 1×10^5 cells ml⁻¹; at a food concentration of 1×10^4 , development was rarely completed. This result is in agreement with experiments on other species of barnacle nauplii larvae by Hirano (1962) who used *Skeletonema costatum* at 10^4 to 10^5 cells ml⁻¹; by Moyse (1963) who utilized *Phaeodactylum tricornutum* at 5×10^4 to 10^6 cells ml⁻¹; by Freiberger and Cologer (1966) who used 1 to 2×10^5 cells ml⁻¹ of *Cyclotella nana* (= *Thalassiosira pseudonana*); and finally by Tighe-Ford et al. (1970) who used *Skeletonema costatum* at 1 to 2×10^6 cells ml⁻¹. Only Hirano, who employed a running water

system, was able to obtain some cyprids at food concentrations as low as 1×10^4 cells ml⁻¹.

Length of Development

The median time interval required for 50% of the nauplii to reach the cyprid stage was computed to the nearest 0.1 d by linear interpolation from data in Table 2. Results of these computations are compiled in Table 3 and show that at 2×10^5 cells ml⁻¹ there is a statistically significant difference in the median time required for development to the cyprid between 20 and 25°C ($p \cong 0.001$) but not between 25 and 30°C ($p > 0.10$).

At the lower algal concentration of 1×10^5 cells ml⁻¹ there is a statistically significant difference in median time required for development between each temperature interval ($p \cong 0.05$ between 20 and 25°C; $p \cong 0.001$ between 25 and 30°C).

When comparing the effect of temperature between the 2 algal concentrations used there is a significant difference in median time required for development to the cyprid at both 20 and 25°C ($p \cong 0.01$ and 0.001 , respectively) but not at 30°C. These relationships can be seen in Fig. 2 which shows the difference in response of the nauplii larvae to increasing temperature in relation to the concentration of food present; a shorter time interval was required at both 20 and 25°C to reach the cyprid stage at 2×10^5 than at 1×10^5 cells ml⁻¹. At 30° there is no significant difference in median time to complete development between the two concentrations.

DISCUSSION

The temperature range within which the development of *Balanus eburneus* nauplii normally occurs in

Table 1. *Balanus eburneus*. Percent survival to cyprid stage by nauplii at 3 different temperatures using 2 concentrations of mixed unicellular algae, *Pavlova lutheri*, *Platymonas* sp. and *Thalassiosira pseudonana* at ratios of 1:1:1 and 1:1:2 as a source of food*

Algal species ratio**	2×10^5 cells ml ⁻¹			1×10^5 cells ml ⁻¹		
	20°C	25°C	30°C	20°C	25°C	30°C
1:1:1	88	96	98	58	60	92
1:1:1	92	90	94	62	62	84
1:1:2	96	100	98	62	72	88
1:1:2	94	100	98	72	70	94
Mean	92.5	96.5	97.0	63.5	66.0	89.5
s	3.4	4.7	2.0	6.0	5.9	4.4
$s/\sqrt{n-1}$	2.0	2.7	1.2	3.4	3.4	2.6

* Survival data derived from Table 2
 ** There is no statistically significant difference between survival values of the 2 different 'algal species ratios'. Consequently, these ratios have been ignored in computing the mean values for survival

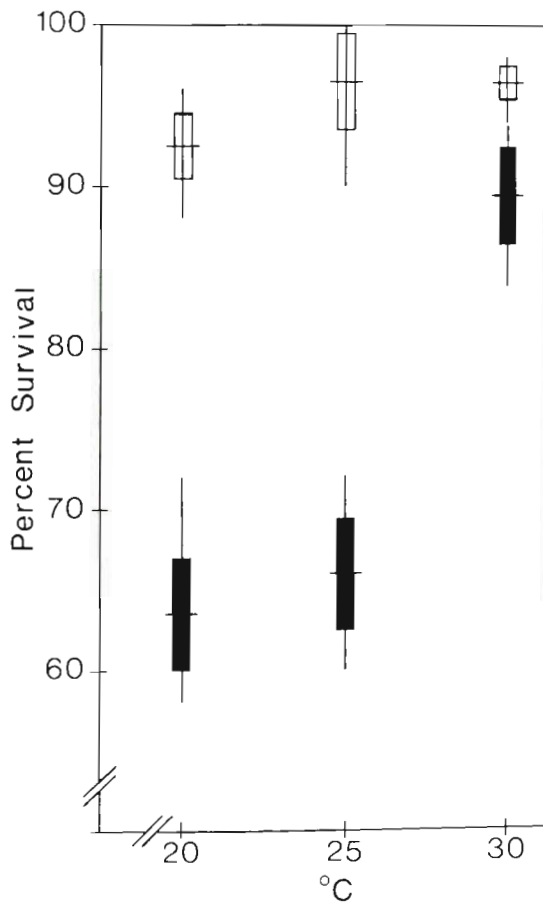


Fig. 1. *Balanus eburneus*. Percentage survival of nauplii larvae to the cyprid stage at 3 different temperatures. Vertical bars: results of 4 experiments with 50 larvae each. Filled bars: cultures of nauplii grown at an algal concentration of 1×10^5 cells ml^{-1} ; open bars, at an algal concentration of 2×10^5 cells ml^{-1} . Mean survivals indicated by short horizontal lines; length of bars shows 2 standard errors on each side of mean survival. Long vertical lines through the bars are extreme survival values. Data derived from Table 2 and compiled in Table 1

natural populations at Woods Hole, Massachusetts, USA lies between 20 and 30°C. Laboratory experiments show that an increase from 20 to 25° does not significantly enhance larval survival; however, doubling the food concentration from 1×10^5 to 2×10^5 cells ml^{-1} at either 20 or 25°C increases mean survival by 30%. Further increase in temperature from 25 to 30° again does not significantly affect larval survival at the higher cell concentration tested ($2 \times 10^5 \text{ ml}^{-1}$), but increases survival by more than 20% at the lower cell concentration of $1 \times 10^5 \text{ ml}^{-1}$ ($p = 0.001$). At 30°C there is no statistically significant difference in survival of nauplii between the two food concentrations tested (Table 1, Fig. 1).

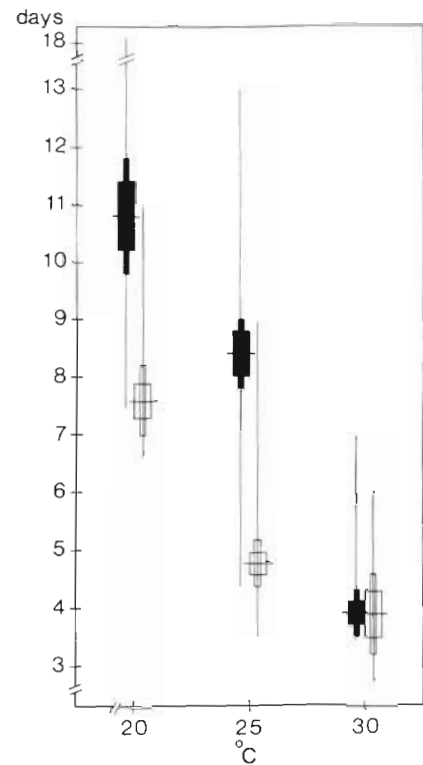


Fig. 2. *Balanus eburneus*. Mean of the median number of days required by nauplii to reach the cyprid stage at 3 different temperatures. Vertical bars: 4 experiments with 50 larvae each. Filled bars: cultures of nauplii grown at an algal concentration of 1×10^5 cells ml^{-1} ; open bars, at an algal concentration of 2×10^5 cells ml^{-1} . Means of the median values for the 4 experiments indicated by short horizontal lines; one standard error and one standard deviation are shown on each side of the mean median value by the wide and narrow portions of the vertical bar, respectively. Vertical line through the bar shows shortest and longest time required to reach cyprid stages in all experiments represented by the bar (i.e. not the extremes of the median values). Data derived from Tables 2 and 3

Over the interval from 20 to 30°C, the time to complete development decreased 49% at 2×10^5 cells ml^{-1} , and 64% at the lower food level of 1×10^5 cells ml^{-1} . A significant part of the decrease in time required to the cyprid stage at the higher algal concentration occurred between 20 and 25°C. In contrast, at the lower food concentration the decrease was more uniform over the temperature interval between 20 and 30°C (Table 3, Fig. 2).

Some changes in survival and rate of development of nauplii that are related to temperature appear to be paradoxical. For example, why should larvae survive significantly better at 30 than at 25°C when grown at low cell levels of $1 \times 10^5 \text{ ml}^{-1}$, whereas the same

Table 2. *Balanus eburneus*. Number of nauplius larvae attaining the cyprid stage in cultures of 50 individuals as recorded at 1-d intervals from the time of their release; larvae grown at 3 different temperatures using 2 concentrations of mixed unicellular algae *Pavlova lutheri*, *Platymonas* sp. and *Thalassiosira pseudonana* as a source of food

Algae at 2×10^5 cells ml ⁻¹ ; ratio 1:1:1							Algae at 2×10^5 cells ml ⁻¹ ; ratio 1:1:2						
Days since release	20°C		25°C		30°C		Days since release	20°C		25°C		30°C	
	Replicate A	Replicate B	Replicate A	Replicate B	Replicate A	Replicate B		Replicate A	Replicate B	Replicate A	Replicate B	Replicate A	Replicate B
1	-	-	-	-	-	-	1	-	-	-	-	-	-
2	-	-	-	-	-	-	2	-	-	-	-	-	-
3	-	-	-	-	-	-	3	-	-	-	-	6	16
4	-	-	-	-	-	4	-	-	7	11	44	33	
5	-	-	23	19	47	44	5	-	-	37	32	-	-
6	-	-	17	18	1	-	6	-	-	4	6	-	-
7	4	1	7	7	-	-	7	8	31	2	1	-	-
8	18	24	1	-	-	-	8	30	12	-	-	-	-
9	13	9	-	1	-	-	9	7	4	-	-	-	-
10	•	•	-	-	-	-	10	•	-	-	-	-	-
11	9	10	-	-	-	-	11	3	-	-	-	-	-
12	-	-	-	-	-	-	12	-	-	-	-	-	-
13	-	2	-	-	-	-	13	-	-	-	-	-	-
Total to cyprid**	44	46	48	45	48	48	Total to cyprid**	48	47	50	50	50	49

Algae at 1×10^5 cells ml ⁻¹ ; ratio 1:1:1							Algae at 1×10^5 cells ml ⁻¹ ; ratio 1:1:2						
Days since release	20°C		25°C		30°C		Days since release	20°C		25°C		30°C	
	Replicate A	Replicate B	Replicate A	Replicate B	Replicate A	Replicate B		Replicate A	Replicate B	Replicate A	Replicate B	Replicate A	Replicate B
1	-	-	-	-	-	-	1	-	-	-	-	-	-
2	-	-	-	-	-	-	2	-	-	-	-	-	-
3	-	-	-	-	-	-	3	-	-	-	-	-	-
4	-	-	-	-	20	17	4	-	-	-	-	41	45
5	-	-	-	-	18	21	5	-	-	1	1	2	2
6	-	-	1	4	6	4	6	-	-	2	2	1	-
7	-	-	4	5	2	-	7	-	-	7	5	-	-
8	-	-	4	5	-	-	8	3	-	10	8	-	-
9	3	-	4	4	-	-	9	6	6	9	6	-	-
10	•	•	•	•	-	-	10	•	•	•	•	-	-
11	9	9	16	9	-	-	11	15	23	7	12	-	-
12	•	•	1	4	-	-	12	•	•	-	-	-	-
13	14	14	-	-	-	-	13	7	7	-	1	-	-
14	-	•	-	-	-	-	14	-	-	-	-	-	-
15	1	7	-	-	-	-	15	-	-	-	-	-	-
16	-	-	-	-	-	-	16	-	-	-	-	-	-
17	-	-	-	-	-	-	17	-	-	-	-	-	-
18	-	1	-	-	-	-	18	-	-	-	-	-	-
Total to cyprid**	27	31	30	31	46	42	Total to cyprid**	31	36	36	35	44	47

• Cultures not examined this day
 ** Total of the original 50 nauplii to reach cyprid stage; difference from original 50 at start represents mortality

temperature increase has no apparent effect on mortality of larvae at high algal concentrations? Why are there no differences in the survival or rate of development at 30°C regardless of the concentration of algal cells?

It is difficult to imagine how the availability of food alone could account for decreased survival of the lar-

vae in any of the experiments. Even the lowest concentration of unicellular algae used in the laboratory experiments exceeded that available to most natural populations of barnacle larvae. For instance, the total mean standing crop of phytoplankton in lower Vineyard Sound, Massachusetts has been measured at 2.3×10^{-1} cells ml⁻¹ and only in certain small New

Table 3. *Balanus eburneus*. Median number of days required to reach the cyprid stage by nauplii at 3 different temperatures using 2 concentrations of mixed unicellular algae, *Pavlova lutheri*, *Platymonas* sp. and *Thalassiosira pseudonana* at ratios of 1:1:1 and 1:1:2 as a source of food*

Algal species ratio**	2×10^5 cells ml ⁻¹			1×10^5 cells ml ⁻¹		
	20°C	25°C	30°C	20°C	25°C	30°C
1:1:1	8.0	5.0	4.5	11.2	9.3	4.2
1:1:1	7.9	5.2	4.5	11.9	8.4	4.2
1:1:2	7.5	4.5	3.4	9.9	7.8	3.5
1:1:2	6.8	4.4	3.3	10.0	8.2	3.5
Mean of median values	7.6	4.8	3.9	10.8	8.4	3.9
<i>s</i>	0.6	0.4	0.7	1.0	0.6	0.4
$s/\sqrt{n-1}$	0.3	0.2	0.4	0.6	0.4	0.2

* Median time to complete development to cyprid derived from data in Table 2
 ** No statistically significant relationship can be demonstrated between median days for development and algal species ratios, therefore algal species ratios have been ignored in computing the mean of the median values

England estuaries or ponds have values as high as 2.2×10^5 cells ml⁻¹ been measured (Smayda, 1957; Hulburt, 1963). Yet the way larval development responds to temperature is apparently modified by the amount of food present.

One explanation of this paradox may lie in the ability of nauplii to harvest algal cells, i.e. the feeding efficiency of the larvae. At 30°C the concentration of food cells is seemingly irrelevant; larval survival and length of development were nearly the same at both 1×10^5 and 2×10^5 cells ml⁻¹. At 20°C, however, increasing the food concentration not only increased survival but also decreased the length of time for larval development to the cyprid stage. These results suggest that at 30°C a maximum feeding efficiency (intake per unit time) has already been reached and possibly surpassed at 1×10^5 cells ml⁻¹, and that at 2×10^5 cells ml⁻¹ there is a surplus of food which cannot be used. Apparently at lower temperatures increasing the food concentration can compensate for decreased feeding efficiency and result in an enhanced survival and a shorter time to complete development to the cyprid stage (Fig. 2). Barnes and Barnes (1958) anticipated such an hypothesis when they concluded that the availability of food may be 'fundamentally important in the development of planktonic larvae . . . [and] may interact with and compensate for temperature effects'.

The total length of planktonic life is determined not only by the 6 nauplius stages but also by the final non-feeding cyprid which is only indirectly dependent upon the factors which control growth and survival of earlier instars. Lucas et al. (1979) have shown that the maximum time that a cyprid of *Balanus balanoides* can remain competent to settle - up to 5 wk - is related to the amount of stored lipid energy-reserves. The

amount of this reserve found in a cyprid will depend on conditions under which the naupliar stages grow. Tighe-Ford et al. (1970, p. 399) found that larvae of *Elminius modestus*, cultured under otherwise identical conditions, are smaller (shorter overall length) at the 6th nauplius stage when grown at 20° than at 10°C. Can it be that at high temperatures the respiration and other catabolic activities increase at the expense of anabolic processes or 'scope for growth', and that increase in biomass is not closely coupled to morphological differentiation of nauplii larvae? Is it not possible that nauplii larvae grown at high temperatures will have less stored energy reserves?

Even though the maximum potential length of the planktonic cyprid stage is a function of food reserves, its actual length will be determined by the availability of a surface for attachment, and in *Balanus balanoides*, *Elminius modestus* and several other species of barnacles settlement is enhanced by an aggregation or intraspecific response of larvae to resident juveniles or adult members of its own species (Knight-Jones and Stephenson, 1950; Knight-Jones, 1953; Crisp, 1963). Accordingly, the length of the cyprid stage in *B. balanoides* will vary from a few days up to a maximum of 5 wk, depending both on an encounter with a site suitable for settlement and upon the amount of energy reserves.

To summarize, we propose the hypothesis that both temperature and food concentration affect feeding efficiency and can modify the rate of development of the nauplius and the amount of food reserves found in the cyprid. To test this hypothesis will require research on the precise relationship of cell concentration and temperature to the feeding efficiency. The effect of high algal and larval concentrations presently used in

laboratory experiments are unknown and the question of scaling may present some difficult future technical problems.

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LITERATURE CITED

- Barnes, H., Barnes, M. (1958). The rate of development of *Balanus balanoides* L. larvae. *Limnol. Oceanogr.* 3: 29-32
- Costlow, J. D., Bookhout, C. G. (1957). Larval development of *Balanus eburneus* in the laboratory. *Biol. Bull. mar. biol. Lab., Woods Hole* 112: 313-324
- Crisp, D. J. (1963). Adsorbed layers: the stimulus to settlement in barnacles. *Proc. R. Soc. Lond. B.* 158: 364-387
- Crisp, D. J., Ritz, D. A. (1967). Changes in temperature tolerance of *Balanus balanoides* during its life-cycle. *Helgoländer wiss. Meeresunters.* 15: 98-115
- Davis, H. C., Guillard, R. R. (1958). Relative value of ten genera of micro-organisms as food for oyster and clam larvae. *Fish. Bull. U. S.* 58: 293-304
- Fish, C. J. (1925). Seasonal distribution of the plankton of the Woods Hole region. *Bull. Bur. Fish, Wash.* 41: 91-179
- Freiberger, A., Cologer, C. P. (1966). Rearing acorn barnacle cyprids in the laboratory for marine fouling studies. *J. Am. Soc. nav. Engrs.* 78: 881-890
- Henry, D. P., McLaughlin, P. A. (1975). The barnacles of the *Balanus amphitrite* complex (Cirripedia, Thoracica). *Zool. Verh., Leiden* 141: 3-254
- Hirano, R. (1962). Mass rearing of barnacle larvae. *Bull. mar. Biol. Station Asamuchi, Tohoku Univ.* 10: 77-80
- Hulburt, E. M. (1963). The diversity of phytoplankton populations in oceanic, coastal, and estuarine regions. *J. mar. Res.* 21: 81-93
- Knight-Jones, E. W. (1953). Laboratory experiments on gregariousness during settling in *Balanus balanoides* and other barnacles. *J. exp. Biol.* 30: 584-598
- Knight-Jones, E. W., Stephenson, J. P. (1950). Gregariousness during settlement in the barnacle *Elminius modestus* Darwin. *J. mar. biol. Ass. U. K.* 29: 281-297
- Landau, M., D'Agostino, A. (1977). Enhancement of laboratory cultures of the barnacle *Balanus eburneus* Gould using antibiotics. *Crustaceana* 33: 223-224
- Landau, M., D'Agostino, A. (1978). Culture of the barnacle *Balanus eburneus* in artificial seawaters. *Crustaceana* 34: 315-318
- Lang, W. H. (1977). The barnacle larvae of North Inlet, South Carolina (Cirripedia: Thoracica). Ph. D. dissertation, University of South Carolina
- Lucas, M. I., Walker, G., Holland, D. L., Crisp, D. J. (1979). An energy budget for the free-swimming and metamorphosing larvae of *Balanus balanoides* (Crustacea: Cirripedia). *Mar. Biol.* 55: 221-229
- Moyse, J. (1963). A comparison of the value of various flagellates and diatoms as food for barnacle larvae. *J. Cons. perm. int. Explor. Mer* 28: 175-187
- Scheltema, R. S. (1967). The relationship of temperature to the larval development of *Nassarius obsoletus* (Gastropoda). *Biol. Bull. mar. biol. Lab., Woods Hole* 132: 253-265
- Smayda, T. H. (1957). Phytoplankton studies in lower Narragansett Bay. *Limnol. Oceanogr.* 2: 342-359
- Tighe-Ford, D. J., Power, M. J. D., Vaile, D. C. (1970). Laboratory rearing of barnacle larvae for antifouling research. *Helgoländer wiss. Meeresunters.* 20: 393-405

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