Simultaneous measurement of phytoplanktonic primary production, nutrient and light availability along a turbid, eutrophic UK east coast estuary (the Colne Estuary)

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ABSTRACT: Primary production, algal biomass, light conditions and dissolved inorganic nutrient concentrations were monitored simultaneously along the Colne Estuary, UK for a period of 15 mo. Phytoplanktonic primary production and chlorophyll a (chl a) concentration ranged from 0.64 × 10⁻³ to 17 µgC l⁻¹ h⁻¹ and 0.5 to 37.5 µg chl a l⁻¹, respectively. Nutrient concentrations decreased along a downstream gradient, with high concentrations of nitrate (564 µM), ammonium (291 µM) and phosphate (52 µM) present at the head of the estuary. Despite high light-attenuation coefficients (mean $K = 3.2$ m⁻¹), the greatest phytoplankton biomass and primary production were measured at the head of the estuary, where nutrient concentrations were high, and phytoplankton biomass and primary production decreased down the estuarine gradient. Despite year-round high nutrient concentrations, chlorophyll a concentrations and rates of primary production remained below levels reported for bloom formation in similar systems. Comparison of field data with measurements derived from phytoplanktonic $P$ vs $E$ curves showed that simulated in situ rates of surface-water biomass-normalised production ($P_B$) were usually significantly lower than the $P_B^{\text{max}}$ values of the phytoplankton. Therefore phytoplankton primary production in the Colne Estuary was light-limited for most of the year, and increased algal biomass and production towards the head of the estuary (despite high attenuation coefficients) was due to a shallow, well-mixed water column (low $Z_{\text{mix}}:Z_{\text{eu}}$ [critical mixing ratio]).

KEY WORDS: Phytoplankton · Estuary · Primary production · Chlorophyll a · Light-limitation

INTRODUCTION

Estuaries are characterised by strong gradients in river flow, circulation, tidal mixing, nutrient inputs, sediment loading and temperature. The interplay of these properties produces spatially and temporally heterogenous conditions for phytoplankton (Lawrence & Harding 1994). These heterogenous conditions cause seasonal and spatial variation in the estuarine environment, and in biological processes such as phytoplankton growth and primary production and in the factors controlling these biological processes (Pennock & Sharp 1994). Estuaries are among the most productive environments and their importance in terms of carbon fixation, fisheries habitat, nutrient assimilation, water storage and sediment stabilisation has long been recognised (Baban 1997). A few studies of estuaries provide a simultaneous examination of temporal and spatial variability in phytoplankton production and factors causing this variability (Pennock & Sharp 1986, Cloern 1996, Malone et al. 1996, Kinney & Roman 1998). However, relatively little information is available on the spatial and temporal variability of estuarine phytoplankton, especially in the turbid European estuaries (Cadee & Hegeman 1974, Kromkamp & Peene 1995).
Estuarine phytoplankton production is often considered to be regulated by either light or nutrients, although the two may interact (MacIsaac & Dugdale 1972, Underwood & Kromkamp 1999). Light is the major regulator of primary production in turbid estuaries (Cole & Cloern 1984, 1987, Pennock 1985, Soetart et al. 1994, Kromkamp & Peene 1995). Although nitrogen is usually considered to be the nutrient limiting primary production in marine and coastal marine systems (D’Elia et al. 1986, Hecky & Kilham 1988), the evidence for this is not as strong as the evidence that freshwaters tend to be P-limited. The situation regarding N and P is even more uncertain for estuaries, as the interplay of heterogenous conditions causes spatial and seasonal variation in the variables limiting primary production (Malone et al. 1996, Underwood & Kromkamp 1999). The general seasonal pattern is of light-limitation during winter and N-limitation over summer, while a spatial trend is identified with N-limitation at the seaward end and P-limitation at the freshwater end (D’Elia et al. 1986, Fisher et al. 1992, Pennock & Sharp 1994, Doering et al. 1995).

This paper presents the results of a study of phytoplankton activity in a hypernutrified estuary, the Colne Estuary, UK, which exhibits a strong gradient of nutrient concentrations from the freshwater to the marine end (Ogilvie et al. 1997). The seasonal and spatial variation in primary production, chlorophyll a (chl a), dissolved inorganic nutrients (ammonium, nitrate, phosphate and silicate) and light characteristics (as depth of euphotic zone, $Z_{mix}$,$Z_{eu}$ ratios, depth-integrated average irradiance) were examined, and the data are discussed with regard to light-limitation.

**MATERIALS AND METHODS**

**Sampling area.** The Colne is a shallow, well-mixed (Sage 1995, Ogilvie et al. 1997), macro-tidal (tidal range 3.5 to 4.0 m) estuary in SE England, entering the North Sea at Brightlingsea (Fig. 1). The estuary is comparatively short (10 km long), but branching (90 km shore-line), with 5 tidal arms running into the main river channel (Fig. 1 and Elliot et al. 1993). The depth of the water column at high tide varies between 14 m at Brightlingsea, 10 m at Aldboro Point, 5 m at Wivenhoe and 3 m at Hythe. Salinity varies between 0 and 33 along the estuary on high tide. The biological and chemical quality of the water is affected by riverine discharge of nutrients and by discharges from domestic sewage treatment works (STW), resulting in a strong gradient of increasing concentrations of ammonium, nitrate, silicate and phosphate towards the head of the estuary (Ogilvie et al. 1997). STW effluent is the main source of ammonium and phosphate while riverine discharge accounts for most of the total oxidized nitrogen, TON (= nitrate + nitrite), and silicate (Elliot et al. 1993).

Four sites (Hythe, Wivenhoe, Aldboro Point and Brightlingsea; Fig. 1) were selected to cover the estuarine salinity gradient. Quadruplicate samples of surface water were collected at each site in mid-channel at high tide approximately every month from September 1994 (nutrients from June) to November 1995. Samples were collected over 2 successive days to enable primary production incubations to be commenced within 1 h of sample collection. On the first day, Brightlingsea and Aldboro Point were sampled from midchannel using an inflatable boat. The narrow nature of the 2 remaining sites (Wivenhoe and Hythe) permitted sampling off the edge of a tidal storm-surge barrier and a quayside, using a rope and bucket. Algal biomass (as chl a concentration) dissolved inorganic nutrients (ammonium, nitrate, nitrite, phosphate and silicate), the vertical light-attenuation coefficient ($K$, m$^{-1}$) and phytoplanktonic primary production were measured.
**Physical measurements.** Temperature was measured by a mercury thermometer. An optical handheld refractometer (Model 10419, Leica) was used to measure salinity. The light-attenuation coefficients were calculated by measuring the light intensity at the water’s surface and at 0.3 m depth by a Skye PAR (photosynthetically active radiation) sensor. The attenuation coefficient $K$ (m$^{-1}$) was then calculated as:

$$K = \left( \frac{\ln I_o - \ln I_z}{z} \right)$$

where $I_o$ and $I_z$ are the light intensities at the surface and 0.3 m, respectively (Nielsen & Bresta 1984). The depth of the euphotic zone ($Z_{eu}$) was calculated as the depth at which irradiance was 1% of the surface value, obtained from the attenuation coefficient $K$ ($Z_{eu} = 4.6/K$).

**Determination of $Z_{mix}$ and $Z_{eu}$ ratios.** The depth of the deepest part of each sector (sampling sites) of the estuary at Brightlingsea, Aldboro Point, Wivenhoe and Hythe at high tide was calculated from UK Admiralty Chart No. 3741. The water column in the Colne Estuary is completely mixed (Ogilvie et al. 1997) and therefore the depth of the water column at each sampling site at high-tide was regarded as the depth of the mixed zone ($Z_{mix}$). Then $Z_{max}$ was divided by $Z_{eu}$ to obtain $Z_{mix}/Z_{eu}$ ratios.

**Biomass.** A cold methanol extraction technique was used for pigment analysis (Jensen 1978). A suitable volume of pigment was filtered through Whatman GF/F filters and placed in methanol at 4°C in darkness for 24 h before measurement at 665 and 750 nm.

**Taxonomic composition of phytoplankton.** Qualitative observations on the taxonomic composition of the phytoplankton community were made using light microscopy. To facilitate identification of diatoms, samples were fixed with glutaraldehyde (50% v/v) to 1% final concentration and acid-cleaned; permanent slides were prepared in Naphrax solution (Northern Biological Supplies).

**Dissolved inorganic nutrients.** Water samples collected for dissolved inorganic nutrient analyses were filtered through Whatman GF/F filters and frozen in liquid nitrogen before storing at –20°C until analysed. Nitrate, nitrite, phosphate and silicate were analysed according to Parsons et al. (1984). Ammonium concentration was measured using a modification of the indophenol blue method (Harwood & Kuhn 1970) with di-isochlorocyanurate replacing bleach as the chlorine donor (Krom 1980).

**Primary production.** The simulated in situ rate of primary production was measured by $^{14}$C-uptake (Nielsen & Bresta 1984) with samples continuously mixed using a rolling incubator. The rolling incubation (75 rpm) prevented settlement of phytoplankton during the incubation period and simulated light conditions in the surface of the water column. This provided a better estimation of the simulated in situ rate of production at the surface, and in preliminary experiments yielded values of primary production ~2.5 fold higher than in a static incubation (data not shown). For each site, 4 light and 4 dark bottles (250 ml, borosilicate glass) were filled with water from quadruplicate containers. Immediately before incubation, we added 100 µl 5 µCi (185 kBq) ml$^{-1}$ NaH$^{14}$CO$_3$ solution (commercially prepared ampules, specific activity 1.85 GBq mmol$^{-1}$, pH 9.5: Dupont Nen) to each bottle, which was then placed on a rolling incubator in a temperature-controlled water bath situated outside under natural illumination. The temperature in the water bath was adjusted to the in situ temperature of the site by a thermo-circulator (Betta-Tech controls, Milton Keynes, UK). Incubations lasted 2 h over midday (11:00 to 13:00 h) under natural daylight. At the end of the incubation, biological activity was stopped by adding glutaraldehyde solution (50% v/v) to the samples (to a final concentration of 1% v/v). Samples were filtered through Whatman GF/F. After filtration, filters were washed with filtered site water, drained, and then exposed to fumes of concentrated hydrochloric acid in a desiccator to remove residual inorganic $^{14}$C.

To measure assimilated $^{14}$C present in the dissolved organic carbon (DOC) leaked from cells, the filtrate from each sample was acidified to pH <2 and purged with air to remove residual inorganic $^{14}$C. Subsamples (1 ml) of filtrate and filter papers with algal cells were placed in 20 ml plastic vials which were then filled with scintillation liquid (Optiphase Safe, LKB Wallac). Radioactivity was counted on a liquid scintillation counter (1217 Rack Beta, LKB Wallac). Any quenching was corrected by using an external standard.

**Total carbon dioxide.** Carbonate alkalinity and all forms of carbon dioxide were determined according to Parsons et al. (1984) on the day of sampling.

**PE curve experiments.** Photosynthesis vs irradiance (PE) curves were determined on samples from the Hythe on a number of occasions over the annual primary production cycle. This site was chosen as the sampling site on the basis of its having the highest chl $a$ concentration throughout the study. For the PE incubations, a range of light intensities (1425, 838, 540, 271 and 0 µmol m$^{-2}$ s$^{-1}$) were used (tungsten lamps). TriPLICATE light and dark bottles were incubated for 2 h and processed as for simulated in situ primary production measurements.

**Determination of light-saturated zone as a percentage of mixed depth.** Surface irradiance measurements at 30 min intervals on each day between September 1994 and November 1995 were obtained (Essex University data). The average solar noon irradiances (Sm, solar midday) for each day of each month were...
obtained from these data, and the total daily insolation (mol d⁻¹) was calculated as:

$$\text{Daily total (mol d}^{-1} \text{)} = \left[\frac{2n}{\pi} \right] S_m$$

where \(n\) is the average daylight length in seconds for the particular month in question, and \(S_m\) is the monthly averaged irradiance (µmol m⁻² s⁻¹) (Platt et al. 1990). Total daily insolation was converted into mean daily irradiance (µmol m⁻² s⁻¹) by dividing by average monthly daylength. Average daily surface irradiance was integrated for 0.1 m depth intervals down the water column using the \(K\) value for each month to obtain the average watercolumn irradiance for each site. The depth at which light was saturating (i.e., where irradiance was >\(I_m\)) was regarded as the bottom of the light-saturated zone. The saturating light intensities (\(I_m\)) were obtained from the \(PE\) curve experiments (see above).

**Statistical analysis.** Environmental variables were analysed using 2-way ANOVAs (site and time as factors) to determine significant spatial and temporal patterns. The percentage of the total sum of squares (SS) was used to determine the contribution of spatial and temporal patterns of variation in the data. Pearson’s correlation coefficients determined correlations between nutrient concentrations and algal biomass, while Spearman’s rank correlation (non-parametric) was used to determine significant correlations between water-column light climate (\(K, Z_{\text{mix}}:Z_{\text{eu}}, I, \text{ etc.}\)) and phytoplankton (chl a, primary production, \(P_{B}\)) variables.

**RESULTS**

**Physical measurements**

The average winter (November to March) and summer (April to October) temperatures were 7.6 and 16°C, respectively. There was no significant difference in water temperature between sites along the estuary. Salinity varied between 0 and 33 along the estuary at high tide. Vertical light-attenuation coefficients (\(K\) m⁻¹) ranged from 0.39 to 5.89, with a general decrease down the estuary (mean annual values of 3.2, 2.4, 1.8 and 1.4 at the 4 sites, respectively). Annual variation in \(K\) was greatest at Brightlingsea (coefficient of variation, CV, of annual mean = 55%), with an order of magnitude decrease in \(K\) values between December/January and June/July of almost 1. At the Wivenhoe and Hythe sites, seasonal variation in \(K\) was much less (CV 31 to 33%) (Fig. 2a). The depth of the euphotic zone (depth of 1% light) increased down the estuary from an annual mean \(Z_{\text{eu}}\) of 1.5 m at Hythe to 4.5 m at Brightlingsea.

\(Z_{\text{mix}}:Z_{\text{eu}}\) ratios varied between 8.5 in November 1994 at Brightlingsea and 0.85 in October 1995 at Aldboro Point. In general, at the 4 sites, \(Z_{\text{mix}}:Z_{\text{eu}}\) ratios were higher during winter than summer; and were higher in the lower estuary, where the water column was on average deeper than in the upper estuary (Fig. 2b). \(Z_{\text{mix}}:Z_{\text{eu}}\) ratios were significantly correlated (\(r_s = 0.32, p < 0.05\)) with \(K\) values.

**Dissolved inorganic nutrients**

The concentrations of all the major dissolved inorganic nutrients (ammonium, nitrate, phosphate and silicate (Fig. 3) showed significant spatial and temporal variation (ANOVA, \(p < 0.001\)) along the Colne Estuary (Table 1). Common to each nutrient was a concentration gradient decreasing down the estuary as salinity increased, with concentrations of nutrients sometimes 2 orders of magnitude higher at the head of the estuary compared to the mouth (Fig. 3).

Differences between sites (Fig. 3a) were responsible for the greatest amount of variation in ammonium concentrations (67%, \(p < 0.001\)) while seasonal changes accounted for ~17% of the variance in ammonium concentration (Table 1). The maximum ammonium concentration measured was 291 µM at the Hythe in July 1994 (Fig. 3a), with values as low as 2 µM at Brightlingsea on the same sampling day.

Spatial variation between sites accounted for 41% of the variance in nitrate concentrations (Fig. 3b). Nitrate concentrations showed a much stronger seasonal pattern (49% of variance: Table 1) than ammonium, with
higher concentrations present during the winter and spring months. Nitrate concentrations varied between 5.75 µM at Brightlingsea in October 1995 and 564 µM at the Hythe in December 1994. Nitrate was by far the most abundant form of inorganic nitrogen in the estuary, and was also present in much higher molar concentrations than other dissolved nutrients (Fig. 3).

Phosphate concentrations varied between 0.1 µM (in July 1994 at Brightlingsea) to 52 µM (in September 1995 at Hythe). Variation in phosphate concentration was largely due to spatial differences (55% of variation), with higher concentrations at the head of the estuary. There was a significant decrease in phosphate concentrations in the estuary during the period April to June, particularly at the 3 lower estuary stations, with concentrations falling below 1 µM at Brightlingsea and Aldboro Point (Fig. 3c).

Seasonal differences accounted for 50% of the variance in silicate concentrations (Table 1), with peak silicate concentrations in early spring and in mid-autumn at Hythe (88 and 168 µM, respectively: Fig. 3d). High concentrations of silica in early spring (late February and March) decreased sharply between April and June (concentrations <0.1 µM at Brightlingsea and Aldboro Point) before increasing in the later summer period. With the single exception of October 1995 at Brightlingsea and Aldboro Point, N:Si ratios along the estuary throughout the sampling period were above the Redfield N:Si ratio of 1, attaining a maximum of >1400 in May 1995 at Brightlingsea (Table 2).

**Phytoplankton biomass and taxonomic composition**

Chl a concentrations varied between 0.5 µg chl a l⁻¹ (in March at Brightlingsea) and 37.5 µg chl a l⁻¹ (in July 1995 at Hythe). Water-column chl a exhibited a clear seasonal pattern, with lowest values measured at all sites between January and March 1995, after which there was an increase in chl a to peak values in July (Fig. 4a). This seasonal pattern of variation accounted for 75% of the variance in chl a concentration data (Table 1), but there were also significant site and site × time effects, with higher concentrations of chl a at the
head of the estuary than at the seaward end in all
months except December to February (Fig. 4a). Maxi-
mum chl $a$ concentrations were measured at Hythe in
July 1995 in a phytoplankton community largely con-
sisting of flagellated euglenophytes (12 to 47 µm in
length). Flagellates were the major components of the
phytoplankton in all seasons, although their abun-
dance decreased in winter. Qualitative observation
also revealed evidence of some resuspension of ben-
thic epipelagic diatoms (e.g. *Cylindrotheca closterium*,
*Nitzschia* spp. and *Navicula* spp.) in the water column
throughout the year along the estuary, particularly
at the 2 upper sampling sites. The relative abundance
of benthic diatoms was highest during spring and
decreased in the summer, but generally diatoms were
relatively rare (compared to flagellates) within the
phytoplankton. There were significant positive corre-
lations ($p < 0.01$) between chl $a$ concentrations and
ammonium and phosphate concentrations and temper-

cature ($r^2 = 0.29$, 0.19 and 0.55, respectively, $n = 240$)
over the 15 mo study period. Water-column chl
$a$ concentrations were positively correlated with daily
and water-column irradiance (Table 3).

**Phytoplanktonic primary production**

The magnitude of simulated surface phytoplank-
tonic primary production (measured at *in situ*

temperatures under natural illumination) varied during the
study between $0.64 \times 10^{-3} \mu g C l^{-1} h^{-1}$ (in February at
Brightlingsea) and ~17 µgC l–1 h–1 (in August at Hythe)
(Fig. 4b). Seasonal changes accounted for 92% of the
variation in primary production, with a minimum in
primary production during February, followed by a
period of almost exponential increase to peak values in
August (Fig. 4b). Spatial differences along the estuary
explained only 1% of the variation in the primary pro-
duction data over the 15 mo period, but during the
period of rapid increase in primary production there
were significantly higher ($p < 0.01$) rates of primary
production towards the head of the estuary (Fig. 4b).
Primary production rates were significantly correlated
with chl $a$ ($r_s = 0.63$, $p < 0.001$), and with incident and
integrated water column irradiance values (Table 3).

**Biomass-specific primary production**

*(P*$_B$, µgC µg–1 chl $a$ h–1) showed a seasonal trend similar to that of primary
production (Fig. 5a). *P*$_B$ values ranged between
$4 \times 10^{-4}$ µgC µg–1 chl $a$ h–1 (in February at Bright-
lingsea) and 3 µgC µg–1 chl $a$ h–1 (in August at Wiven-
hoe). During the initial autumn period, *P*$_B$ was signifi-
cantly higher at the downstream sites (Aldboro and

Table 1. Two-way ANOVA of variables measured in the
The percentage of the total sum of squares (SS) explained by
each factor within the ANOVA is given. All $F$-values are
significant at $p < 0.001$

<table>
<thead>
<tr>
<th>Variable</th>
<th>Factor</th>
<th>df</th>
<th>% of total SS</th>
<th>$F$</th>
</tr>
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<td>NH$_4$$^+$</td>
<td>Time</td>
<td>14</td>
<td>16.6</td>
<td>451</td>
</tr>
<tr>
<td></td>
<td>Site</td>
<td>3</td>
<td>67</td>
<td>8533</td>
</tr>
<tr>
<td></td>
<td>T × S</td>
<td>42</td>
<td>16</td>
<td>145</td>
</tr>
<tr>
<td>NO$_3$$^-$</td>
<td>Time</td>
<td>14</td>
<td>49</td>
<td>1093</td>
</tr>
<tr>
<td></td>
<td>Site</td>
<td>3</td>
<td>42</td>
<td>4297</td>
</tr>
<tr>
<td></td>
<td>T × S</td>
<td>42</td>
<td>9</td>
<td>64</td>
</tr>
<tr>
<td>PO$_4$$^-$</td>
<td>Time</td>
<td>14</td>
<td>34</td>
<td>1590</td>
</tr>
<tr>
<td></td>
<td>Site</td>
<td>3</td>
<td>51</td>
<td>1100</td>
</tr>
<tr>
<td></td>
<td>T × S</td>
<td>42</td>
<td>15</td>
<td>226</td>
</tr>
<tr>
<td>SiO$_3$$^-$</td>
<td>Time</td>
<td>14</td>
<td>66</td>
<td>3989</td>
</tr>
<tr>
<td></td>
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<td>3</td>
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<td>7553</td>
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<tr>
<td></td>
<td>T × S</td>
<td>42</td>
<td>8</td>
<td>156</td>
</tr>
<tr>
<td>Chl $a$</td>
<td>Time</td>
<td>14</td>
<td>76</td>
<td>1382</td>
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<td></td>
<td>Site</td>
<td>3</td>
<td>14</td>
<td>1169</td>
</tr>
<tr>
<td></td>
<td>T × S</td>
<td>42</td>
<td>10</td>
<td>62</td>
</tr>
</tbody>
</table>
| Primary produc-
| tion       | Site   | 3  | 1             | 189   |
|               | T × S  | 42 | 7             | 87    |

Table 2. N:Si ratios at 4 sampling sites between June 1994
and November 1995 in the Colne Estuary

<table>
<thead>
<tr>
<th>Date</th>
<th>Brightlingsea</th>
<th>Aldboro</th>
<th>Wivenhoe</th>
<th>Hythe</th>
</tr>
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<tbody>
<tr>
<td>1994</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jun</td>
<td>27.48</td>
<td>22.30</td>
<td>22.74</td>
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<td>Jul</td>
<td>1.44</td>
<td>1.30</td>
<td>1.64</td>
<td>6.15</td>
</tr>
<tr>
<td>Aug</td>
<td>1.34</td>
<td>3.42</td>
<td>2.06</td>
<td>4.67</td>
</tr>
<tr>
<td>Sep</td>
<td>5.32</td>
<td>11.63</td>
<td>8.48</td>
<td>5.40</td>
</tr>
<tr>
<td>Oct</td>
<td>5.16</td>
<td>4.79</td>
<td>4.12</td>
<td>8.48</td>
</tr>
<tr>
<td>Nov</td>
<td>9.80</td>
<td>11.84</td>
<td>19.05</td>
<td>11.16</td>
</tr>
<tr>
<td>Dec</td>
<td>6.09</td>
<td>10.47</td>
<td>19.99</td>
<td>20.16</td>
</tr>
<tr>
<td>1995</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jan</td>
<td>8.97</td>
<td>12.69</td>
<td>7.81</td>
<td>11.48</td>
</tr>
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<td>7.99</td>
<td>11.64</td>
<td>7.34</td>
<td>10.00</td>
</tr>
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<td>Mar</td>
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<td>6.32</td>
<td>5.96</td>
<td>8.94</td>
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<td>23.96</td>
<td>22.96</td>
<td>14.07</td>
<td>14.09</td>
</tr>
<tr>
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<td>1480.9</td>
<td>677.4</td>
<td>489.08</td>
<td>106.64</td>
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<tr>
<td>Jun</td>
<td>530.6</td>
<td>43.18</td>
<td>23.06</td>
<td>21.20</td>
</tr>
<tr>
<td>Jul</td>
<td>34.76</td>
<td>29.27</td>
<td>8.20</td>
<td>12.47</td>
</tr>
<tr>
<td>Aug</td>
<td>11.55</td>
<td>7.73</td>
<td>3.07</td>
<td>65.33</td>
</tr>
<tr>
<td>Sep</td>
<td>2.31</td>
<td>3.13</td>
<td>2.56</td>
<td>8.73</td>
</tr>
<tr>
<td>Oct</td>
<td>0.49</td>
<td>0.81</td>
<td>1.09</td>
<td>1.02</td>
</tr>
<tr>
<td>Nov</td>
<td>2.61</td>
<td>2.28</td>
<td>2.88</td>
<td>4.48</td>
</tr>
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</table>
Brightlingsea), although by December this pattern had disappeared. There was a similar exponential increase in $P_B$ from April to August (Fig. 5a). $P_B$ was correlated with both primary production and chl $a$ concentrations ($r_s = 0.83, 0.36, p < 0.001, 0.05$, respectively), but also showed a significant negative correlation with light-attenuation, $K$ (Table 3).

Light-saturated rates of primary production ($P_{B \text{max}}$) obtained from $P_E$ curves were generally greater than $P_B$ measured under natural illumination conditions (Fig. 5a), except during August and October 1995. $I_m$ values were always higher than the calculated average integrated light intensity in the water column at each site (Fig. 5b), and in late summer $I_m$ and the average daily surface irradiance were similar. The percentage of the mixed depth ($Z_{\text{mix}}$) at which irradiance was $>I_m$ ranged between 2.7 and 21.6 throughout the period of study. There was a significant increase up the estuary in the percentage of the water column in which $I > I_m$ ($F = 9.5, p < 0.001$), with the lowest values at Brightlingsea (Fig. 6).

**DISCUSSION**

The rates of primary production, biomass, and dissolved inorganic nutrient concentrations showed significant seasonal and spatial variation in the Colne Estuary (Table 1). The seasonal changes in production and biomass were very similar, with higher values during summer. This reflected the fact that, in general, seasonally driven changes such as light conditions in the water column regulated both primary production and phytoplankton biomass. The peaks in primary production and biomass occurred in August and July, respectively. The occurrence of a spring (usually diatom-dominated) and summer peak in both phytoplankton biomass and production is common in temperate estuaries (e.g. Malone et al. 1996, Marker & Collett 1997), but there was no distinct spring bloom in the Colne Estuary (Fig. 4). Rather, production and algal biomass began to increase during March and April and remained at their highest levels during the summer, a scenario indicative of light-limited situations (van Spaendonk et al. 1993). Decreases in production and biomass after the spring bloom are usually attributed to nutrient limitation, increased grazing or parasitism. The Colne Estuary shows clear seasonal patterns in nutrient concentrations, primarily due to seasonal changes in the inputs of nutrients from the river and sewage treatment works (Dong et al. 2000). In the upper reaches of the estuary, nutrient concentrations far exceed those usually considered limiting to algal growth. Kromkamp & Peene (1995) suggested that although primary production in estuaries can be high in comparison to nearby coastal areas due to relatively high nutrient concentrations, this may not always be

![Fig. 4. Seasonal and spatial changes (mean ± SD, n = 4) in (a) chl $a$ concentrations and (b) primary production at 4 sampling sites along the Colne Estuary between September 1994 and November 1995. Symbols as in Fig. 3](image)

Table 3. Spearman’s rank-correlation coefficients between water-column light climate and phytoplankton variables (n = 60; ns: not significant, *p < 0.05, **p < 0.01, ***p < 0.001). $P_B$: biomass-normalised production

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mean daily irradiance</th>
<th>Mean water-column irradiance</th>
<th>Light attenuation ($K \text{m}^{-1}$)</th>
<th>$Z_{\text{mix}} - Z_{\text{eu}}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chl $a$ ($\mu$g l $^{-1}$)</td>
<td>0.49***</td>
<td>0.48***</td>
<td>0.13 ns</td>
<td>−0.1 ns</td>
</tr>
<tr>
<td>Primary production ($\mu$g C l $^{-1}$ h $^{-1}$)</td>
<td>0.31*</td>
<td>0.32*</td>
<td>−0.20 ns</td>
<td>−0.12 ns</td>
</tr>
<tr>
<td>$P_B$ ($\mu$g C $\mu$g chl $a$ l $^{-1}$ h $^{-1}$)</td>
<td>0.17 ns</td>
<td>0.19 ns</td>
<td>−0.36**</td>
<td>−0.17 ns</td>
</tr>
</tbody>
</table>
the case since estuaries, especially the well-mixed coastal plain systems, are often very turbid, imposing light limitation on production. Similarly in the Delaware Estuary, despite high anthropogenic nutrient inputs, changes such as shifts in phytoplankton species composition, high chl a concentrations and depressed oxygen concentrations have not been observed, because of light-limitation of algal production (Pennock 1985). In the Colne Estuary, chl a concentrations did not reach levels typical of nuisance algal blooms. Chl a concentrations were always <50 µg l⁻¹, with an annual mean of ~5 µg l⁻¹, which is at the lower end of the range for estuarine habitats (Heip et al. 1995). This was much lower than in other nearby estuaries (e.g. Great Ouse Estuary: Rendell et al. 1997), despite high nutrient concentrations, especially in the upper reaches, all year round.

There were some seasonal changes in the broad composition of the algae in the estuary. Spring phytoplankton displayed an increased abundance of diatoms compared to the rest of the year, and there was a clear decrease in silicate concentrations during this period of increasing chl a and primary production (Figs. 3 & 4). Diatoms disappeared from the summer phytoplankton community, possibly due to silicate limitation, and the species composition shifted to an assemblage rich in flagellates. Silicate concentrations in the Colne at the end of winter period were comparable with concentrations in the Chesapeake Estuary, USA, which showed a similar pattern of spring decline of silicate coincident with diatom growth (Conley & Malone 1992). Similar species shifts during the growing season have been observed in other estuaries (Bakker et al. 1994). A change from diatom-dominated to flagellate-dominated phytoplankton communities is also a feature of coastal eutrophication and the development of nuisance algal blooms (Underwood & Kromkamp 1999). Even though large standing stocks of phytoplankton did not develop, the conditions in the Colne (high nutrient concentrations, low light availability) appear to favour the dominance of flagellates.

The N:Si ratios in the Colne were always >1, indicating a potential stoichiometric limitation by silicate. Such potential limitation of biomass would occur in an algal community only if silicate dropped to limiting concentrations, when diatoms would be replaced by flagellates, with possible consequences to the food chains in the estuary. Franzs (1986) reported that increased nutrient discharge from the Rhine, Meuse and Scheldt rivers had multiplied the summer flagellate blooms in Dutch coastal waters by a factor of 2 to 4 since 1930, and had caused a shift in dominance from diatoms to other algae. Similarly, in the northern Adri-
atic Sea and the northern Gulf of Mexico, the changes in N:P and Si:P ratios over a 30 yr period resulted in these systems becoming potentially less limiting in terms of N and P but more Si-limited (Justic et al. 1995). Justic et al. (1995) hypothesised that silicon deficiency might be exacerbating eutrophication by reducing the role of diatoms in coastal food webs. More recent evidence comes from the River Danube, where Humborg et al. (1997) reported a decrease in silicate concentration and increase in N and P inputs due to damming of the river. This has resulted in dramatic shifts in the phytoplankton composition from diatoms (siliceous) to coccolithophores and flagellates (non-siliceous).

Another possible reason for the general lack of diatoms in the phytoplankton community in the Colne Estuary during the summer could be decreased resuspension of bottom sediments due to reduced wind energy. Resuspension of benthic microalgae by tidal and wind-induced currents has been shown to contribute up to 25% of the suspended chl a in the Ems Estuary (de Jonge & van Beusekom 1995), while in the turbid Gironde Estuary the strong correlation between chl a concentration and suspended particulate material suggested an important influence of resuspension on the distribution of chl a in the turbidity maximum zone (Irigoien & Castel 1997).

The seasonal trend in $P_B^*$ was similar to that in primary production, with higher values during the summer; but while over the whole period of study primary production was significantly correlated with irradiance, $P_B^*$ showed no significant correlation with $I$. So although biomass and primary production began to increase in March/April (correlated with increases in average irradiance), major changes in $P_B^*$ did not occur until June. By July and August, $P_B^*$ values were close to the $P_B^*_{\text{max}}$ values reflected in the PE curves (Fig. 5a), indicating that at this time algal photosynthesis was not light-limited at the surface of the water column. During the rest of the year, $P_B^*$ was lower than $P_B^*_{\text{max}}$, indicating that phytoplankton were not photosynthesising at their physiological maximum potential rate for most of the year. The values of $P_B^*_{\text{max}}$ measured in the Colne Estuary were lower than in other comparable studies (Kromkamp & Peene 1995). $P_B^*_{\text{max}}$ values were not directly related to temperature, peaking some time after the highest water temperatures in July (see Kocum et al. 2002, this issue, for further discussion).

Comparison of the $I_m$ and $K$ values and the mean daily incident and depth-integrated water-column irradiance showed that, on average, <14% of the mixed zone depth was light-saturated throughout the sampling period in the estuary. This is an average based on mean daily irradiance, so at times during the daily insolation cycle, saturating light intensities would be present further down the water column. Light attenuation coefficients in the Colne Estuary were high, especially in the upper reaches, resulting in a shallow euphotic zone ($Z_{\text{eu}}$). This might indicate the possibility of increased light limitation up the estuary. However, $Z_{\text{max}}:Z_{\text{eu}}$ ratios were lower in the upper estuary as a result of a shallow water-column depth (Fig. 7). The $Z_{\text{max}}:Z_{\text{eu}}$ ratio relates the depth of the mixed layer to the euphotic zone, and thus gives some indication of the average time spent by the algae in the dark and in the light. With larger $Z_{\text{max}}:Z_{\text{eu}}$ ratios, the average time spent by algae in the light decreases, and thus net photosynthesis decreases. Grobbelaar (1990) suggested that the $Z_{\text{mix}}:Z_{\text{eu}}$ ratio would be the most important factor affecting phytoplankton production in turbid ecosystems, which usually have large $Z_{\text{mix}}:Z_{\text{eu}}$ ratios. Furthermore, Cloern (1996) suggested that blooms often develop when the effective mixing depth of the water column is reduced by vertical stratification, thereby decreasing the ratio. Values of $Z_{\text{mix}}:Z_{\text{eu}} < 5$ to 20 have been proposed as necessary for net primary production to occur in an ecosystem (Wofsy 1983, Cloern 1987, 1996, Grobbelaar 1990). In the Colne Estuary, the $Z_{\text{mix}}:Z_{\text{eu}}$ ratios varied only between 0.85 and 8.5, and increased down the estuary as the water depth increased (Fig. 2). The only site which under these criteria could be considered to approach light limitation (i.e. no net primary production; ratio > 5 according to Wofsy 1983) was Brightlingsea during the winter months (Fig. 2). However, while calculations of critical mixing ratios may indicate the potential for net community growth, they take no account of absolute light levels in the water column. Comparisons of $P_B^*$ values with $P_B^*_{\text{max}}$ and averaged water-column irradiance with $I_m$ values show that on average, light would
have been a major limiting factor for photosynthesis in the water column for the whole period studied. Yet, during particular periods during the summer months, there would have been times when photosynthesis was light-saturated during the daily insolation curve. Interestingly, highest chl a, primary production and P\textsuperscript{B} values were measured in the turbid upper parts of the estuary at the Hythe (mean annual Z\textsubscript{eu} = 1.5 m). Thus, at the Hythe, in the absence of nutrient limitation, the low Z\textsubscript{max}:Z\textsubscript{eu} ratio due to the shallowness of the water column permitted high production rates despite the high turbidity (Fig. 7). Since the Z\textsubscript{max}:Z\textsubscript{eu} ratio was much higher at the seaward end of the Colne Estuary at Brightlingsea, phytoplankton were on average subject to greater light-limitation in the clearer lower estuary than in the more turbid upper estuary, especially during winter. This apparent paradox was due to the fact that the water column in the Colne is completely mixed and stratification does not occur (Ogilvie et al. 1997). Thus, in contrast to what high K values intuitively indicate, there was less possibility of light-limitation in the shallow upper estuary than in the deeper lower estuary (Fig. 7). Although they did not measure primary production, Fichez et al. (1992) also showed that in the Great Ouse Estuary, UK, there was evidence of development of primary production in the shallow water despite a shallow euphotic zone and a totally mixed water column, and rationalised this in terms of the Z\textsubscript{max}:Z\textsubscript{eu} ratio. Van Spaendonk et al. (1993) found that in the Westerschelde Estuary, the Netherlands, high turbidity resulted in light-limited growth of phytoplankton, and calculated a high annual primary production (200 to 300 gC m\textsuperscript{-2}) despite the fact that the vertical mixing zone was 5 times larger than the euphotic zone. They concluded that their result corroborated Grobbelaar’s (1990) hypothesis that the value of the Z\textsubscript{max}:Z\textsubscript{eu} at which phytoplankton can still be effective producers can be seriously underestimated, and that phytoplankton can still cope effectively with turbid conditions when Z\textsubscript{max}:Z\textsubscript{eu} is >20.

An estimated annual rate of primary production was calculated for each of the 4 sites in the Colne Estuary using the formulae for chl a-normalised primary production (P\textsuperscript{B}, g C gchl \textsuperscript{a} \textsuperscript{-1} h\textsuperscript{-1}: Cullen 1990), the mean chl a concentrations, water-column depth and light-intensity through the water column. This gave annual rates of 5.5, 4.2 and 4.3 g C m\textsuperscript{-2} yr\textsuperscript{-1} for Brightlingsea, Aldboro Point and Wivenhoe, respectively, and 160.4 g C m\textsuperscript{-2} yr\textsuperscript{-1} for Hythe. The mean annual production rates for each sector were multiplied by the area of water surface at mid-tide in the relevant sector of the estuary (Robinson 1996), all sectors were totalled, and then the total estuarine production was divided by the total area of estuary to derive an overall average production for the whole estuary. This was equivalent to 8.9 g C m\textsuperscript{-2} yr\textsuperscript{-1}, placing the Colne Estuary at the lower end of the range of the estuarine annual production values given in Underwood & Kromkamp (1999). The estuary is ‘oligotrophic’ in relation to the ranges of annual estuarine production suggested by Nixon (1995) despite the very high nutrient concentrations present. The outer Colne Estuary values were similar to those measured in the Bristol Channel by Joint & Pomeroy (1981), where light is also limiting due to high turbidity.

In conclusion, our study showed that throughout the estuary algal photosynthesis was almost always light-limited, even at the surface, with P\textsuperscript{B} < P\textsubscript{max} except during late summer. In late summer at the water surface, factors other than light would regulate photosynthesis. However, in only 6 to 13 % of the upper water column did average daily light intensity exceed I\textsubscript{max}, and even in late summer the algae throughout most of the water column would have been light-limited.

Despite higher turbidity, the critical mixing ratio ensured that phytoplankton cells were maintained more effectively in the higher light environments in the upper estuary than in the lower estuary. Even so, the average light intensity throughout the water column was far below I\textsubscript{max} so that algal photosynthesis remained light-limited even where Z\textsubscript{max}:Z\textsubscript{eu} was low. However, the more beneficial light regime in the upper estuary, combined with excess of nutrients, maintained high algal biomass in the upper estuary despite the relatively rapid flushing time.

Acknowledgements. We would like to thank Mr. R. Littlewood and Mr. J. Green for their help with sample collection. E.K. was supported by a studentship from the Turkish Higher Education Council to ‘TC Canakkale Onsekiz Mart University.’

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Submitted: January 11, 1999, Accepted: September 20, 2001
Proofs received from author(s): March 18, 2002