INTRODUCTION

Deep-sea hydrothermal vents are island habitats harbouring high levels of faunal biomass in the otherwise resource-starved deep-sea environment. This rich vent fauna is supported by local microbial chemolithoautotrophic production, in particular through oxidation of reduced sulphur and iron from the hydrothermal fluid (Karl 1995). One component of the vent fauna directly exploits microbial primary production by hosting endosymbiotic bacteria. This is the case for the vestimentiferan tubeworms, clams and mussels that constitute a large part of the biomass at hydrothermal vents in the eastern Pacific Ocean (Fisher 1995, Sarrazin & Juniper 1999). However, the greater portion of the vent faunal diversity and organism abundance is constituted by non symbiont-containing (NSC) organisms such as polychaetes and gastropods (Sarrazin et al. 1997, 1999, Tunnicliffe et al. 1998, Tsurumi & Tunnicliffe 2003). These organisms are often assumed to directly rely on a diet of free-living autotrophic microorganisms. Although there remains little doubt that chemolithoautotrophic bacteria and archaea constitute the base of vent NSC food chains (Karl 1995), a direct trophic link between these microbes and NSC metazoa, while often presumed, has never been thoroughly tested. In fact, many of these species feed on particulate organic matter (POM) through suspension- and deposit-feeding, as
evidenced by behavioural and morphological observations (Desbruyères & Laubier 1986, Tunnicliffe et al. 1993, Grelon 2001, Morineaux et al. 2002), gut content examinations (McHugh 1987, Tunnicliffe et al. 1993, Chevaldonné 1996) and stable isotope analyses (Van Dover & Fry 1994, Colaço et al. 2002, C. Levesque, S. K. Juniper & H. Limén unpubl. data). In addition to autotrophic microbes, other particulate organic constituents are present in the vent environment, including heterotrophic bacteria and archaea, protists, micrometazoa and detrital organic matter of local and allochthonous origin (Brault 1984, Brault et al. 1984, Comita et al. 1984, Albéric 1986, Daumas et al. 1988, Karl 1995, Levesque & Juniper 2002). As feeding on POM is apparently an important trophic strategy at vents, a thorough description of vent-associated POM is essential to elucidate the trophic position of vent NSC species, and to expand our general understanding of the functioning of vent food webs. The aim of this study was to trace the origin, describe the composition and assess the nutritional quality of POM at hydrothermal vents on Axial Volcano, northeast Pacific.

MATERIALS AND METHODS

Study site. Axial Volcano is located on the central segment of the Juan de Fuca Ridge (NE Pacific) at 46°55’N, 130°00’W (Fig. 1). The volcano is capped by a 3 × 8 km horseshoe-shaped caldera (depth ~1500m) where extensive hydrothermal venting occurs at 3 known vent fields: ASHES (Axial Seamount Hydrothermal Emissions Study), the South Rift Zone (SRZ) and CASM (Chase et al. 1985, Embley et al. 1990).

Nineteen active hydrothermal vent sites (12 in the SRZ, 6 at ASHES vent field and 1 at CASM vent field) were visited at Axial Volcano during New Millennium Observatory (NeMO) project cruises in September 1998, June to July 1999, June to July 2000, and July 2001 (Table 1). Some sites were visited more than once, and altogether samples represented 30 site–year combinations. As a point for comparison, 6 non-vent sites with deposits of pelagic sediment (at least 200 m away from any known vent) were sampled.

All vent sites, except 3, were ~1 to 10 m wide areas on the basaltic seafloor where warm (mean temperature = 19.1°C, SE = 2.2°C, n = 29), diffuse hydrothermal fluid was issuing from the oceanic crust. The remaining 3 sites were hosted on sulphide edifices bathed in diffuse vent fluid. All sites supported endemic vent faunal assemblages composed mainly of alvinellid polychaetes, gastropods and vestimentiferan tube-worms (see Tsurumi et al. 1998, Tsurumi 2001 and Marcus 2003 for detailed descriptions of the faunal assemblages of these sites).

Among the 12 SRZ sites, 7 were located on a lava flow formed by a January–February 1998 eruption (Embley et al. 1999). The age of these nascent sites was therefore known with precision. The remaining 5 sites

![Fig. 1. Location of the ASHES (Axial Seamount Hydrothermal Emissions Study), CASM (Canadian American Seamount Expedition) and SRZ (South Rift Zone) vent fields on Axial Volcano, Juan de Fuca Ridge, NE Pacific. The extent of the January–February 1998 lava flow in the SRZ is outlined](image_url)

Table 1. Sampling sites and years at the SRZ, ASHES and CASM hydrothermal vent fields on Axial Volcano. SRZ sites located on the 1998 lava flow are labelled as SRZ (lava). See Fig. 1 for explanation of vent field names

<table>
<thead>
<tr>
<th>Site</th>
<th>Location</th>
<th>1998</th>
<th>1999</th>
<th>2000</th>
<th>2001</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cloud (N4)</td>
<td>SRZ (lava)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cloud (N6)</td>
<td>SRZ (lava)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Marker 33</td>
<td>SRZ (lava)</td>
<td>•</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Marker 108</td>
<td>SRZ (lava)</td>
<td>•</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Marker N41</td>
<td>SRZ (lava)</td>
<td>•</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nascent</td>
<td>SRZ (lava)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Snail (N8)</td>
<td>SRZ (lava)</td>
<td>•</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bag City</td>
<td>SRZ</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coquilles</td>
<td>SRZ</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Joystick</td>
<td>SRZ</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Marker 113</td>
<td>SRZ</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Old Worms</td>
<td>SRZ</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gollum</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Haido</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Medusa-base</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Medusa-top</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phoenix</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pork Chop</td>
<td>ASHES</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T&amp;S</td>
<td>CASM</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
in the SRZ were located 50 to 200 m beyond the limits of the lava flow. Vent assemblages at these latter SRZ sites, as well as in the ASHES and CASM vent fields, pre-dated those at the nascent sites, but their approximate age could not be determined.

**Sampling.** The remotely-operated vehicle ROPOS was used for all sampling. Temperature at vent openings was recorded with temperature probes on the SUAVE fluid sampler (Massoth et al. 1991), the Hot Fluid Sampler (D. Butterfield, University of Washington & Pacific Marine Environmental Laboratory, Seattle, WA, USA), or by a thermistor mounted on the inlet of the ROPOS suction sampling device. The maximum venting fluid temperature reached during the observations (~3 to 10 min) was recorded.

**Particulate matter:** Particulate matter (PM) was collected with a suction-sampling device that pumped water into 2 l acrylic jars. Two layers of 200 µm Nitex nylon mesh at the outflow of each jar allowed sufficient flow to provide suctioning capability, yet avoid forcing soft particulates through the filter mesh. Sample accumulation in the suction jars was monitored with the ROPOS colour video camera. Suction was maintained until the filter mesh was visibly clogged and the flow rate substantially diminished. Operated in this manner, and at low to moderate flow rates, the suction sampler acted as a broad spectrum concentrator of particles and attached bacteria, rather than as an in situ filtration system for >200 µm particles. Finer mesh filters clogged too quickly to permit collection of enough material for analyses. During sampling, the suction intake was held in the centre of tubeworm assemblages or, for the non symbiont-containing fauna, slowly swept over the substratum within the faunal assemblage. At non-vent sites, sediments were sampled using the same device. One jar and mesh set were used for each sample, and the collection hose was flushed with bottom seawater between samples to avoid cross-contamination. Samples remained at ambient bottom temperature (~2°C) in closed jars until brought to the surface at the end of the dive.

On board the ship, PM was left to settle in the jars for 30 to 60 min at 4°C. Duplicate aliquots of PM were immediately taken for ATP determinations (see procedure below), for fixation in cacodylate-buffered glutaraldehyde (3% final concentration) and for fixation in seawater-buffered formalin (7% final concentration). Remaining sample material was frozen (–80°C) in polypropylene tubes or collected on pre-combusted (500°C) GF/F or GM/F glass fibre filters before freeze-drying, sputter-coated with gold, and kept at 4°C until analysed for dissolved inorganic carbon (DIC) stable isotopic composition.

**Stable isotope analyses.** In order to trace the origin of particulate organic matter, the stable carbon and nitrogen isotopic compositions of POM and DIC were analysed. POM aliquots for stable carbon isotope determination were acidified with 0.1 N HCl to remove carbonates. Vent fluid samples were acidified with phosphoric acid to convert DIC to CO₂, which was analysed for its stable carbon isotopic composition. Stable isotopic compositions were analysed with a Micromass Isoprime isotope ratio mass spectrometer and are reported as follows:

\[
d\% = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 10^3
\]

where \(X\) is 13C or 15N, and \(R\) is 13C/12C or 15N/14N (Ehleringer & Rundel 1989). Standards used were Vienna Pee-Dee Belemnite (VPDB) for carbon and atmospheric N₂ for nitrogen.

**Biochemical analyses.** Nutritional quality of POM was assessed by measuring available protein and total lipid contents, 2 important nutritional components for metazoa (e.g. Mayer et al. 1986, Grémare et al. 1997), as well as the C:N ratio, which provides information on both the origin and lability of organic matter.
culate matter samples were analysed for organic carbon (following treatment with 0.1 N HCl) and nitrogen contents using a Carlo-Erba CN analyser. Available (hydrolysable) protein content was measured colorimetrically in triplicate following the reaction with Coomassie Blue dye using the procedure of Mayer et al. (1986), and is reported as bovine serum albumin equivalents. Lipid content was assessed colorimetrically in triplicate using the sulphophosphovanillin method of Barnes & Blackstock (1973) following lipid extraction as in Folch et al. (1957). The Supelco 37 fatty acid methyl ester mix (Supelco/Sigma-Aldrich) was used as a lipid standard.

**Biomass.** Bacterial counts and ATP measurements were used to provide estimates of bacterial biomass and total microbial biomass, respectively. Immediately upon recovery of the samples on board, ATP was extracted in duplicate in 6 ml of boiling phosphate buffer (60 mM, pH 7.4; Karl 1993) for 10 min, and the extracts were frozen (–20°C). At the land-based laboratory, firefly lantern extract was added to the thawed ATP extract and bioluminescence was measured with a Monolight 2010 luminometer (Analytical Luminescence Laboratory). For conversion of ATP to total microbial biomass carbon, the conversion factor of 250 g C g⁻¹ ATP was used. This ratio is typically applied to natural microbial communities and reflects the variability in the C:ATP ratio between unicellular microorganisms (~200 to 350 g C g⁻¹ ATP) and micrometazoa (50 to 150 g C g⁻¹ ATP; Karl 1980, 1993).

Formalin-preserved samples were DAPI-stained for cell counting by epifluorescence microscopy (Porter & Feig 1980). DAPI-stained bacteria and archaea cannot be discerned by this method and are both included here in the estimated ‘bacterial’ abundance. Cell abundance was converted to bacterial biomass carbon here in the estimated ‘bacterial’ abundance. Cell abundances were also frequent. Epifluorescence microscopic observations of DAPI-stained samples indicated that bacterial cells were dominated by coccoid forms (Fig. 1). Remains of diatoms and coccolithophorids were also frequent. Epifluorescence microscopic observations of DAPI-stained samples indicated that bacterial cells were dominated by coccoid forms (Fig. 2A,B). Amorphous, apparently organic detrital particles (µm to mm scale) were ubiquitous. Remains of diatoms and coccolithophorids were also frequent. Epifluorescence microscopic observations of DAPI-stained samples indicated that bacterial cells were dominated by coccoid forms (Fig. 2C). Filamentous bacteria (µm-scale diameter) and larger (10 to 20 µm) Beggiatoa-like filaments (Nelson et al. 1989) were also present. Amorphous, non DAPI-staining material was abundant in all samples.

**Stable isotopic composition**

Vent POM showed a wide range of δ¹³C values (–11.4 to –26.8‰; Table 2). The mean δ¹³C of vent POM (–20.2‰) tended to be enriched in ¹³C compared to sediments (mean = –25.0‰), but this difference was not statistically significant. The δ¹⁵N of vent POM (mean = 4.3‰) was less variable (2.1 to 6.7‰) than the stable carbon isotopic composition, and was significantly lower than the δ¹⁵N of sediments (mean = 6.7‰; Kruskal-Wallis p = 0.0034, n = 36).

The stable carbon isotopic composition of vent fluid DIC ranged between –0.7 and 8.1‰ and was positively correlated with the δ¹³C of POM (r² = 0.4861, p = 0.0117, n = 12; Fig. 3A) and with fluid temperature at the vent opening (r² = 0.4874, p = 0.0247, n = 10; Fig. 3B).

**Biochemical composition**

Vent particulate matter was mostly inorganic, with sulphur as the dominant constituent element (31 to
Levesque et al.: Particulate matter at deep-sea hydrothermal vents

70 Mol wt %) followed by iron (19 to 27 Mol wt %). The silicon content was significant in some samples (up to 18 Mol wt %). The organic fraction was small, with an average of 4.0 % organic carbon and 0.7 % nitrogen content. In comparison, sediments had a similar organic carbon content (mean = 4.2 %) and a lower nitrogen content (mean = 0.3 %, Kruskal-Wallis p = 0.0258, n = 36; Table 2).

Despite their similar organic carbon content, vent PM and sediments were clearly different in their biochemical composition. The mean C:N ratio measured in vent POM (6.3) was significantly lower than that of sediments (17.4; Kruskal-Wallis: p = 0.0002, n = 36; Table 2). Available protein content was highly variable and tended to be larger in vent POM (mean = 227.2 mg g−1 C) than in sediments (mean = 31.5 mg g−1 C). The average lipid content of vent POM was 39.3 mg g−1 C. Lipids could not be measured accurately in sediments, because clean lipid extracts were not obtained. The ATP content of vent POM (mean = 119.3 µg ATP g−1 C) was ~8 times higher than in sediments (mean = 15.7 µg ATP g−1 C; Kruskal-Wallis p = 0.0253, n = 8). Total microbial biomass carbon, estimated from ATP concentrations, represented on average 3.0 % of vent particulate organic carbon. The abundance of DAPI-staining

![Fig. 2](image_url) (A, B) Scanning electron and (C) epifluorescence micrographs of particulate matter from Axial Volcano diffuse vents, showing diatom remains, sulphur filaments and globules, amorphous material (A, B) and DAPI-stained coccoid bacteria (C)

Table 2. Biochemical composition of vent particulate matter and sediments from Axial Volcano. Mean, standard error (SE) of the mean, range and sample size (n) are given for each variable. Kruskal-Wallis p-values are from comparisons of means between vent particulate matter and sediments. % Corg: percent of organic carbon

<table>
<thead>
<tr>
<th></th>
<th>Vent</th>
<th>Sediment</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>% Corg</td>
<td>Mean 4.0</td>
<td>Mean 4.2</td>
<td>0.7989</td>
</tr>
<tr>
<td>% N</td>
<td>Mean 0.7</td>
<td>Mean 0.3</td>
<td>0.0258</td>
</tr>
<tr>
<td>C:N</td>
<td>Mean 6.3</td>
<td>Mean 17.4</td>
<td>0.0002</td>
</tr>
<tr>
<td>δ13C (‰)</td>
<td>Mean −20.2</td>
<td>Mean −25.0</td>
<td>0.0760</td>
</tr>
<tr>
<td>δ15N (‰)</td>
<td>Mean 4.3</td>
<td>Mean 6.7</td>
<td>0.0034</td>
</tr>
<tr>
<td>Proteins (mg g−1 C)</td>
<td>Mean 227.2</td>
<td>Mean 31.5</td>
<td>0.1123</td>
</tr>
<tr>
<td>Lipids (mg g−1 C)</td>
<td>Mean 39.3</td>
<td>Mean 31.5</td>
<td></td>
</tr>
<tr>
<td>ATP (µg g−1 C)</td>
<td>Mean 119.3</td>
<td>Mean 15.7</td>
<td>0.0253</td>
</tr>
<tr>
<td>Microbial C (% POC)</td>
<td>Mean 3.0</td>
<td>Mean 0.4</td>
<td></td>
</tr>
<tr>
<td>DAPI-staining cells</td>
<td>Mean 5.4</td>
<td>Mean 0.6</td>
<td>0.0606</td>
</tr>
<tr>
<td>(×1011 g−1 C)</td>
<td>Mean 0.5</td>
<td>Mean 0.06</td>
<td></td>
</tr>
<tr>
<td>Bacterial C (% POC)</td>
<td>Mean 0.2</td>
<td>Mean 0.06</td>
<td></td>
</tr>
</tbody>
</table>
cells in vent POM (mean = $5.4 \times 10^{11}$ cells g$^{-1}$ C) was 1 order of magnitude higher than in sediments (mean = $0.6 \times 10^{11}$ cells g$^{-1}$ C), although this difference was not statistically significant (Kruskal-Wallis p = 0.0606, n = 15). Estimated bacterial carbon represented a minor fraction (mean = 0.5%) of the vent particulate organic carbon.

Lipid content of vent POM was positively correlated with both the C:N ratio and protein content (Spearman’s rho = 0.7818 and 0.7619, respectively; p < 0.05; Table 3). Bacterial abundance was negatively correlated with the C:N ratio (Spearman’s ρ = –0.5798) and positively correlated with protein content (Spearman’s ρ = 0.4654; p < 0.05 in both cases).

Temporal variability in particulate matter composition

Temperature, organic carbon and nitrogen, the C:N ratio, δ$^{13}$C, δ$^{15}$N, protein, lipid, ATP contents and bacterial abundance did not differ significantly between sites of different age groups (0.7 to 3.5 yr old nascent sites and non-nascent sites; Kruskal-Wallis p > 0.05 in all cases). Nitrogen content tended to increase at nascent sites from 1998 (0.7 yr) to 2001 (3.5 yr), and the C:N ratio tended to decrease accordingly (data not shown). δ$^{13}$C also tended to decrease during the same period. However, variability in the measured parameters was not significantly greater between than within age groups; therefore, data from all age groups will be discussed together.

DISCUSSION

Origin of vent POM

The POM stable isotopic composition data distinguish particulate carbon and nitrogen sources at vents from those supplying nearby pelagic sediments (Table 2). The significant positive relationship between δ$^{13}$C POM and δ$^{13}$C DIC (Fig. 3) is a strong indicator that the bulk of vent particulate organic carbon is locally fixed from vent fluid DIC. Even the lowest δ$^{13}$C POM values (overlapping with δ$^{13}$C sediment) can be explained in terms of vent fluid DIC stable isotopic composition (Fig. 3), indicating that these lower values do not necessarily reflect contributions from pelagic sources or advected sediment.

The stable carbon isotopic composition of fluid DIC at Axial Volcano diffuse vents spans a large range (–0.7 to 8.1‰; Fig. 3) compared to vent sys-

![Graph A](image1.png)

![Graph B](image2.png)

Table 3. Spearman’s rank correlation coefficients between POM constituents describing its nutritional quality. Values in bold are statistically significant (p < 0.05). ATP was excluded due to low overlap with other measurements.
tems on the Endeavour segment of the same ridge (~1 to 1‰ C. Levesque unpubl. data). Elsewhere, values range from ca. –4.0 to 3.9‰ (Rau 1981, Fisher 1995), depending on location. To our knowledge, the wide range reported here has not been previously observed on a local scale (within a vent field). The wide range on Axial Volcano may originate from kinetic isotopic fractionation of the CO₂-rich fluid due to the relatively shallow depth (~1500 m) compared to most other deep-sea vent systems (e.g.: ~2200 m for Endeavour segment). Phase separation of high temperature hydrothermal fluids prior to venting produces vapour-rich and brine-rich phases at Axial (Massoth et al. 1989).

During this process, kinetic isotopic fractionation of DIC is likely to occur, where isotopically lighter DIC is enriched in the vapour phase, leaving a 13C-enriched DIC is likely to occur for epidermal protection and detoxification (Juniper et al. 1986). Mucus was reported to account for 25% of a 2 kg wet weight biomass sample from an Axial Volcano vent site (Tunnicliffe et al. 1985). *P. palmiformis* mucus has a low C:N ratio (average = 4.6; Juniper et al. 1986); thus, it could be a significant source of non-living, nitrogen-rich organic matter. Other alvinellids (*P. sullicola* and *P. pandorae*) produce mucus-based, nitrogen-rich sheathed tubes (Morineaux et al. 2002), and shredded remains of these structures were observed in our POM samples. Other potential sources of detrital organic matter include faunal mortality (in particular of the sessile, biomass-dominant tubeworms) caused by abrupt shifts in temperature and composition of the vent fluid, and bacterial exudates that sometimes produce biofilms covering mineral and biological structures.

Detrital organic matter has been shown to be quantitatively important at vents elsewhere. Based on analyses of lipid classes in particulate matter from 5 to 40°C vents of the East Pacific Rise (21°N), Comita et al. (1984) concluded that copepods, amphipods and macrofaunal invertebrates (and detritus derived from these organisms) constituted the major proportion of particulate organic matter. At 13°N on the East Pacific Rise, Albéric (1986) argued that the amino acid composition of POM reflected a large contribution of detritus derived from vestimentiferans and alvinellid biomass. Other studies (e.g. Brault 1984, Daumas et al. 1988, Juniper 1988) also suggest that detrital material may be a significant component of particulate matter at other vents in the Pacific. It is likely that detrital organic matter is particularly important in vent systems of the Eastern Pacific that are colonised by organisms shedding mucus or producing mucus-based tubes (such as alvinellid polychaetes), as well as by vestimentiferan tubeworms creating bushes where particulate detritus can accumulate. Detrital material may be quantitatively less important in systems lacking these 2 characteristics, such as vents in the Atlantic and some vents in the Pacific.

The small bacterial fraction observed here and in other studies may reflect a rapid turnover rate of bacterial biomass in the vent POM pool compared to other components such as detritus. Free-living primary productivity is assumed to be intense at vents but bacterial biomass may be kept at low levels by bacterial mortality and grazing by micro- and macro-consumers. Ultimately, the production and consumption of detritus by vent consumer species must be supported by a re-supply of new organic matter from primary producers. This detritus likely fuels heterotrophic microbial activity and may play a major role in organic matter cycling.
at vents. Despite the recognition of a diverse heterotrophic microbiota at vents, to our knowledge the existence and functioning of a heterotrophic microbial food web has never been specifically investigated.

Previous studies of diffuse (Levesque & Juniper 2002) and chimney-hosted vents (Sarrazin et al. 2002) suggested that organic detritus accumulates and POM becomes more heterotrophic as a vent site ages. In this study, the large inter-site variability in POM composition conceals any potential temporal variability. It is likely that site-specific differences in physico-chemical conditions and biotic assemblage composition constrain POM composition by affecting rates of primary production as well as detritus production and consumption.

Implications for vent consumers


The low organic content of vent PM implies that vent suspension- and deposit-feeders should be adapted to processing large volumes of organic-poor material, as are consumers in most sedimentary environments. However, the nutritional quality of our vent POM samples was superior to that of deep-sea sediments, as indicated by the lower C:N ratios.

POM protein and lipid contents were positively correlated, as might be expected for these 2 macromolecular indicators of the labile and usually more nutritional fraction of POM. However, lipids and proteins only accounted for a small fraction of the bulk particulate organic carbon and nitrogen. Lipids (average content = 39.3 mg g\(^{-1}\) C) accounted for less than 4% of organic carbon (even if lipids were assumed to be composed exclusively of carbon). Hydrolysable proteins (average = 227.2 mg g\(^{-1}\) C) represented an average of 12% of organic carbon and 24% of nitrogen, assuming that proteins are 53% C and 17% N (Sterner & Elser 2002). This indicates that a significant part of the particulate nitrogen may not be available to the fauna. Non-hydrolysable proteins and mucopolysaccharides, such as those found in the mucus of alvinellid polychaetes (Juniper et al. 1986), may account for a significant part of the remaining organic carbon and nitrogen. Non-characterizable organic matter may also represent a large part of the organic fraction, as is often the case in sediment.

Bacterial abundance (and hence availability as a food source) was higher in more labile and nutritionally-rich POM (based on protein content and the C:N ratio; Table 3) but remained low overall. Although standing stocks alone cannot establish the importance of nutritional sources for consumers, the large abundance of detrital material observed here and in other investigations suggests that organic detritus is potentially a quantitatively important food source for the non symbiont-containing vent fauna. Measurements of the productivity of autotrophic bacteria (i.e. turnover of living microbial carbon) would provide a better estimate of their availability as a potential direct food source for vent consumers, and as a source for the POM pool.

CONCLUSION

Axial Volcano diffuse vents host a complex and heterogeneous particulate organic matter pool. The bulk of POM is apparently derived from local microbial primary production, yet bacterial biomass constituted less than 1% of sampled POM and is likely insufficient to provide a pure bacterial diet to consumers, without invoking a very rapid turnover of microbial biomass. Only species selectively feeding on pure bacterial mats or on autotrophic bacteria and archaea in the complex POM matrix can operate as strict primary consumers. It is not likely that most species can achieve such a degree of selective feeding. As POM is a complex mixture of autotrophic and heterotrophic microbial cells and detritus, most suspension- and deposit-feeders may include a significant part of detrital organic matter in their diet. In this case, a substantial contribution of autotrophic microbial productivity to the detrital pool would be required in order for the ecosystem to operate sustainably. The assumption that non symbiont-containing vent fauna directly relies on a free-living autotrophic microbial diet may not represent the actual complexity of the trophic links between vent primary producers and consumers. The largely overlooked detrital compartment should be considered in future studies of the trophic ecology and organic matter cycling at hydrothermal vents.

Acknowledgements. We wish to thank chief scientist R.W. Embley and participants of the New Millennium Observatory (NeMO) project, as well as D. Grelon, R.J. Léveillé, M. Morineaux, A. Adamowicz, Z. Bourass, R. Mineau and D. Papineau for their help in sampling and analyses. G.J. Massoth and D.A. Butterfield provided vent fluid temperature data. We also thank the ROPOS team and the crews of the NOAA RV ‘Ronald H. Brown’ and of the University of Washington RV ‘Thomas G. Thompson’ for their support at sea.
acknowledge the comments of 4 anonymous reviewers which substantially improved an earlier version of this manuscript. This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC) and the Fonds québécois de la recherche sur la nature et les technologies (FQRNT). C.L. was supported by post-graduate scholarships from NSERC, FQRNT and Fisheries and Oceans Canada. H.L. was supported by a post-doctoral fellowship from CanRidge, a collaborative research opportunity funded by NSERC.

LITERATURE CITED


Submitted: December 22, 2003; Accepted: October 21, 2004
Proofs received from author(s): March 8, 2005