ABSTRACT: The fate of crab-processed mangrove *Avicennia marina* leaf litter was compared with decomposition of whole leaf litter of *A. marina* and *Rhizophora stylosa*. Shredding by the grapsid crab *Parasesarma erythodactyla* resulted in reduction of mangrove leaf litter to fragments ~200 µm in the faecal material. The C/N ratio decreased linearly with time, attaining ~20 after 28 d. While the δ^{13}C signature remained constant, the δ^{15}N value steadily changed from ~4‰ to 1.8‰ over 4 wk. Decomposing whole leaf litter, however, showed only modest increases in %N content with time, attaining a C/N ratio of ~20 only after 24 wk, with no significant temporal trend in either δ^{13}C or δ^{15}N. δ^{15}N of decomposing crab faecal material was significantly negatively correlated with surface bacteria density. Demonstrating a sigmoid pattern of increase, surface bacteria density on crab-processed leaf fragments was ~70 times higher than that on whole leaf litter after 28 d of decomposition. Analysis of the C/N ratio, δ^{13}C and δ^{15}N signatures of micro-POM collected from 5 mangrove waterways in southern Moreton Bay, Australia, failed to detect significant differences in these parameters at 3 distances from mangrove forests, but location of the waterways had a strong effect. In laboratory feeding experiments involving the copepods *Temora turbinata* and *Oithona rigidis*, a diet of faecal material from crabs significantly improved copepod survivorship compared with a diet of the alga *Nannochloropsis* sp. or the no-food treatment for both copepods. Stable isotope analysis of the copepods suggest that the animals were able to utilise both crab faecal material and algal carbon, with greater assimilation of the former food source, in the mixed diet (*Nannochloropsis* + crab faeces) treatment. By acting as shredders, grapsid crabs process large quantities and effect rapid enrichment of mangrove organic production, potentially benefiting both benthic and pelagic estuarine consumers.

KEY WORDS: Mangroves · Grapsid crabs · Copepods · Outwelling · Decomposition · Micro-POM · Estuarine food chains

INTRODUCTION

Past studies of the trophic ecology mangroves have focussed on the degree of export of productivity of mangroves in support of nearshore consumers, probably a result of their apparent large standing crop, and often also high productivity (Alongi 1998). Much effort has been spent on measuring flux of organic matter and nutrients from mangroves (Twilley 1988), but mangrove–nearshore exchange patterns are still obscure. An emerging picture suggests that organic matter export from stands of mangroves is both limited in quantity and spatial extent (Lee 1995), and some systems may even import organic matter (Alongi 1998). Further, the trophic value of outwelled mangrove matter to nearshore consumers is debatable, as most studies employing chemical tracers such as stable isotopes of carbon, nitrogen and sulphur record little resemblance between consumers and mangrove signatures (Rodelli et al. 1984, Newell et al. 1995, Lee 2000).

Ground-dwelling grapsid crabs have been reported to be the main consumers of fresh mangrove litter, particularly in Indo-Pacific systems (Robertson et al. 1992, Lee 1998). Fresh mangrove litter has low nutrient (C/N
ratio > 50) but high feeding deterrent (e.g. soluble tannins) content, making it an unattractive staple food item (Wolcott & O’Connor 1992). Some grapsid crab species prefer leaf litter with specific C/N ratios (e.g. Lee 1993), while others feed indiscriminately (Micheli 1993). Grapsid crabs nevertheless can consume large percentages of fresh mangrove litter and help retain mangrove productivity within the forest. While it is still unclear how the crabs can benefit from such a poor diet, one of the little studied areas is the fate of the mangrove organic matter consumed and processed by the crabs. Acting as initial processors of mangrove leaf litter, grapsid crabs mediate significant physical, and potentially chemical and biological, changes to mangrove organic production. Ignorance of these changes makes a thorough evaluation of the trophic contribution of tropical mangroves difficult—the link to nearshore production may not be as direct as previously assumed (i.e. export of un consumed and only minimally microbially enriched macro-detritus).

In this study, our aim is to gauge the impact of leaf litter processing by grapsids on the quality of this litter as food for macrofauna and zooplankton in nearshore waters. Our approach consisted of a number of field and laboratory experiments, each testing part of the potential trophic link: (1) a litter decomposition experiment investigating the physical, chemical and biological changes associated with crab-processed and naturally senescent leaf litter; (2) a laboratory experiment comparing the survivorship of 2 common species of nearshore copepods fed diets of an unicellular alga or crab-processed mangrove leaf litter; and (3) an estimation of the contribution of crab-processed mangrove leaf litter to the micro-particulate organic matter (micro-POM) pool of mangrove estuaries in SE Queensland, Australia.

MATERIALS AND METHODS

All sampling were conducted in Moreton Bay, SE Queensland, where extensive mangrove forests dominated by Avicennia marina and, to a lesser extent, Rhizophora stylosa and Aegiceras corniculatum exist. Fig. 1 shows the sampling and experimental locations mentioned in the following subsections. Though not supporting grapsid crab densities comparable to tropical NE Queensland, significant populations of grapsid crabs still live in the local mangroves, with Parasesarma erythodactyla being one of the dominant species.

Transformation of mangrove litter without crab processing. Transformation of mangrove litter relevant to their trophic value (e.g. C/N ratio, surface bacteria density) was compared between pathways with and without grapsid crab involvement. Strictly speaking, this is not a direct comparison, as experimental conditions for the 2 pathways cannot be easily standardised. Freshly senescent leaf litter of Rhizophora stylosa and Avicennia marina was collected from Jabiru Island on the Gold Coast by lightly plucking the petiole of yellow leaves. Leaves that would not detach easily were not included in the sample. The wet weight of the leaves was then recorded to the nearest 0.01 g in the laboratory, before they were individually put into nylon bags of 2 mm mesh. A separate large sample of the leaves of each species was dried at 80°C for 48 h to obtain a regression relationship between wet and dry weight and in order to estimate the initial dry weight of the experimental leaves.

The litterbags were returned to Jabiru Island shortly after the measurements, and placed in the area (mid-intertidal, ~1.7 m above extreme low water spring) where they were first collected. The bags were laid flat on the sediment surface of the mangrove forest, and pneumatophores were cut at sediment level where necessary to ensure regular inundation. Twenty leaves of each species were retrieved at each of the 5 sampling dates, namely, 14, 28, 56, 112 and 168 d after the beginning of the experiment.

The leaf fragments were collected and briefly washed in de-ionised water in the laboratory and the dry weight recorded to indicate the rate of organic matter
loss over time. Five leaves from each combination of type and collection date were set aside for measuring transformations that may determine the trophic value of the material. Leaf squares of 1 cm² were randomly removed from the decomposing leaves in the laboratory and preserved in 3% glutaraldehyde for 2 h, followed by dehydration and the enumeration of bacteria by scanning electron microscopy. Counting of surface bacteria on each leaf disc was made on 20 randomly located spots at 5400× magnification and density was calculated as number per 0.225 mm². Trials using the acridine orange method in estimating bacteria density returned significantly lower densities compared with the SEM approach (authors’ unpubl. data), and the latter method was therefore adopted as an indication of the abundance of bacteria colonising the decomposing mangrove material.

The remaining material from each of the leaves was then dried at 60°C for 48 h and ground for analysis of their stable isotopic signatures for carbon and nitrogen (δ¹³C and δ¹⁵N) and carbon and nitrogen contents.

Transformation of mangrove litter with crab-processing. The transformations undergone by mangrove organic matter consumed by crabs were studied in the laboratory, because differences in the nature of the material (e.g. size of detritus fragments) made it difficult to conduct parallel comparisons in the field. Due to the limitation in available experimental space, only Avicennia marina, the local dominant species, was studied. Individuals of the grapsid crab Parasesarma erythodactyla were collected from the mangrove forest on Jabiru Island. The gut of the crabs was purged by starving them for 48 h and noting that the crabs had finished defaecating before the experiment began. Each crab was kept in a small container in a temperature-controlled room at 26°C and a light:dark cycle of 12:12 h. In order to minimise contamination, no sediment was provided in the containers but enough water (<1 cm) was provided to allow the crab to be kept moist for respiration and, occasionally, molting. The crabs were provided freshly senescent A. marina leaf litter ad libitum, typically at about 1 leaf every 48 h. Faecal pellets produced by the crabs were collected daily, and production by individual crabs was kept separate in smaller containers in a small (<5 ml) volume of seawater, also collected from Jabiru Island. In order to avoid contamination by external suspended particulate organic matter, water used in the experiment was filtered through Whatman No. 1 filter paper beforehand, which removed the particulates but retained the microbes present. The containers with the faecal material were kept on a shelf in a temperature-controlled room and covered with a 2 mm nylon mesh to allow ventilation while minimising contamination due to entry of foreign materials. Samples of the faecal material were collected every 3 d during the 3 wk incubation period. Similar to the whole leaf experiment, sub-samples from each container were collected for SEM analysis. The remaining samples were dried at 60°C and their stable isotope signatures and carbon and nitrogen contents were measured.

Size of leaf fragments in the faecal pellets was measured using light microscopy as described below. Three pellets produced by each of 20 crabs were collected fresh and the contents dispersed in a petri dish for measurement. Thirty fragments were randomly selected from the dispersed contents, and their maximum dimension measured under low power (40 to 100×) using a calibrated ocular micrometer.

SEM measurements were made on 5 separate fields (replicates) for each of the faecal samples, using a Hitachi electron microscope. Counting of bacteria was performed on photographic quadrats at 3000× magnification, each field covering an area of ~375 µm². The average diameter of leaf fragments in crab faeces was ~200 µm, providing enough area for independent bacteria counting on the fragments using the same protocol as in the whole leaf samples.

The stable isotope signature of the litter samples with and without crab processing was measured using a continuous-flow isotope-ratio mass spectrometer, with the appropriate standards. Standards were introduced at regular intervals to provide quality control as well as to give an indication of the level of precision, which was maintained at ± 1% for both C and N. The measurements also provide values of %C and %N for the samples, from which the C/N ratio is calculated.

The nature of suspended particulate organic matter in mangrove waterways. With the chemical characterisation information available, it was possible to assess the prevalence of crab-processed mangrove organic matter in waterways adjacent to mangrove forests. Particulate organic matter was collected at 3 stations: at the interface with the mangrove forest, the mouth of the mangrove creek, and the centre of the main channel along 5 mangrove waterways in southern Moreton Bay, where extensive mangrove stands dominated by Avicennia marina occur (see Fig. 1). These positions with different degrees of proximity to the mangroves would provide a gradient of the availability of mangrove organic matter, in fine particulate form. At each location, up to 10 replicate samples were collected by filtering about 50 l of water through pre-weighed and pre-combusted GF/C papers. Filters were wrapped in individual pre-combusted aluminium foils and dried at 60°C for 48 h before their stable isotope signatures were measured as described above.

The fate of crab-processed mangrove organic matter. Possible trophic linkages originating from crab-processed mangrove organic matter were studied by
performing laboratory-rearing experiments. Lee (1997) demonstrated that small invertebrates such as the amphipod *Parhydella* sp. could beneficially utilise crab faecal material and survived significantly longer periods of time in the laboratory than individuals maintained on a mangrove litter diet. In this experiment, the potential trophic value of resuspended crab-processed mangrove organic matter was evaluated using 2 species of copepods common in local waterways within 2 to 3 km from mangrove forests.

Feeding experiments were conducted on 2 species of common nearshore copepods, namely, *Temora turbinata* (Calanoida) and *Oithona rigida* (Cyclopoida). Since the 2 copepod species occur at different seasons, the 2 experiments were conducted at different times. Samples of *T. turbinata* and *O. rigida* were obtained from the Broadwater, southern Moreton Bay, using a 80 µm plankton net. The copepods were identified and sorted in the laboratory with the help of a stereomicroscope, and male and female animals were identified. Individuals were (1:2 F:M) transferred to 500 ml Schott bottles, which were maintained at 26°C in a constant temperature room with a 12:12 h light:dark regime. Gentle aeration was provided to provide oxygen as well as to keep the food particles in suspension in the containers. Locations of the bottles for the 4 treatments (details below) were randomised so as to minimise the impact of local environmental conditions on the behaviour of the copepods.

Four treatments of food were provided, with 13 replicates for *Temora turbinata* (each with 200 individuals) and 5 for *Oithona rigida* (each with 50 individuals) for each treatment. In one treatment, no food was provided. The 3 food treatments were, respectively, (1) crab faecal material conditioned at 26°C for >2 d (F treatment); (2) the unicellular algae *Nannochloropsis* sp. (A treatment); (3) a combination of crab faecal material and *Nannochloropsis* (M treatment). *Nannochloropsis* culture was obtained from the Bribie Island Aquaculture Research Station of the Department of Primary Industry and Fisheries.

Copepods are known to respond to N limitation. The total N content of the initial ration was therefore standardised with knowledge of the N concentration of the food items. New food was not provided unless there was a visible depletion of material in the containers.

Survival and population density of the copepods in the 4 treatments were monitored every 2 d. Both species of copepods are positively phototactic. The number of copepods in the containers was estimated by shining a light on one end of the container and counting of the number of individuals present. This enumeration method was necessary as other methods (e.g. homogenising the culture and sub-sampling) would inflict significant disturbance to the population. This method is also more accurate in determining the maximum survival period of the animals.

In order to ascertain the relationship between survivorship and the food provided, the δ13C and δ15N values of the food items and the copepods were determined. Pure samples of the 2 food items were measured using the procedure above, while copepods samples were collected from the experimental containers. Several independent samples were analysed by randomly pooling copepods from more than 1 container to provide enough samples for the stable isotope measurements. Copepod samples were mostly freshly dead specimens, usually with little gut contents.

**Data analysis.** Results of the field micro-POM stable isotope signatures were analysed by 2-way mixed-model ANOVAs with replication; location was a random factor and proximity to mangroves (forest, edge and channel) was a fixed factor. A planned contrast was performed on the effect of proximity to mangroves on the various chemical characteristics of the micro-POM.

Survival of the copepods in the feeding experiment was analysed by a repeated-measures ANOVA, with food (4 levels, fixed factor) as the between-subject factor and days of experiment as the repeated measures (within-subject) factor. Data collected on all sampling days except Day 0 (collected every 2 d for 14 and 24 d, respectively, for the *Temora* and *Oithona* experiments) were included in the analysis. This test assesses the difference in pattern of survivorship amongst the 4 food treatments over the experimental period. Where significant effect was detected in the ANOVA and there was no significant food–day interaction, comparison amongst the 4 food treatments was performed using Tukey’s test. All statistical analyses were conducted using SPSS.

**RESULTS**

Transformation of whole and crab-processed mangrove litter

Changes in ash-free dry weight (AFDW) of whole leaves of both mangrove species followed almost linear decrease over the 168 d period (Fig. 2A). Rate of AFDW loss was best described by the linear rather than negative exponential model, and was significantly faster in *Avicennia marina* than in *Rhizophora stylosa*, with the final AFDW remaining for 2 species at 16.8 ± 7.7% and 51.7 ± 13.4% (all values are mean ± 1 SD unless otherwise indicated), respectively. The senescent mangrove leaves were poor in N: the %N was 0.65 ± 0.09 for *A. marina* and 0.41 ± 0.08 for *R. stylosa*. The initial C/N ratio of litter from *A. marina* was 70.4 ± 8.9 and from *R.
stylosa, 115.6 ± 24.9. Change in the atomic C/N ratio (Fig. 2B) of decomposing litter was rapid in the period between 20 to 60 d for both species, with the final values after 168 d at 45.8 ± 5.1 for R. stylosa and 23.8 ± 2.8 for A. marina.

Change in the stable isotope signature of litterbag leaves did not follow distinct trends. δ13C varied widely over the decomposition period in both species, and there was no significant temporal trend (p > 0.05 for r in both cases, Fig. 2C,D). Similarly, there was no significant trend in δ15N for either species, with values fluctuating around +1 and +4‰ for Rhizophora stylosa and Avicennia marina, respectively.

Surface bacteria density of whole-leaf samples increased rapidly with time (Fig. 3A), linearly for both species in the first 56 d of experiment. The density of bacteria was significantly higher in Avicennia marina after 56 d. Bacterial density of decaying crab faecal material, however, conformed to a tight sigmoidal trajectory (r2 = 0.86, n = 15, p < 0.0001) (Fig. 3B), increasing by 8 times over the experimental period and peaking at Day 20.

Feeding by Parasesarma erythodactyla reduced whole mangrove leaf litter to fragments with a mean diameter of 219 ± 93 µm (mean ± 1 SD). Rate of decomposition was linear for the crab-processed mangrove leaf litter with a half-life of 90.1 d (linear regression: faecal material remaining = 90.1 – 0.45 d; r2 = 0.45, n = 15, p = 0.003). Nitrogen content of the faecal material increased slightly but was not correlated with bacteria colonization (r = 0.45, n = 15, p = 0.08) while carbon content decreased linearly (r2 = 0.09, n = 15, p = 0.25). Change in δ15N with time was significant (p = 0.0003) and gradually depleted from an initial average of +5 to +1.8‰ (Fig. 4A). This trend in δ15N with time was significantly negatively correlated (r = –0.73, n = 15, p = 0.003) to bacteria density. δ13C signature remained constant at around –28‰ over the same period. C/N ratio decreased significantly (from 30 to 21) indicating increasing nutritional value in the crab-processed mangrove material with time (Fig. 4C). Decrease in C/N ratio was strongly correlated with increase in bacteria density (r = –0.73, n = 15, p = 0.003).
There was generally no significant interaction between the factors of location and site (position in waterway) in micro-POM chemical composition, except for % organic matter (Table 1). All measured parameters of micro-POM composition showed a significant difference ($p < 0.001$) between sampling locations. Within each location the nature of micro-POM showed no significant differences for most of the parameters measured. $\delta^{13}$C signatures, however, varied significantly with proximity to mangrove forest (2-way mixed model ANOVA, $F = 12.3$, df = 2, 13.9, $p < 0.001$), with a significant contrast ($p = 0.031$) between the forest (F) and channel (C) positions. Micro-POM $\delta^{15}$N values from Jabiru Island and Tingalpa Creek were most depleted and had the closest values to the fresh crab-processed mangrove material $\delta^{13}$C value. $\delta^{15}$N differences to the $\delta^{15}$N faecal material standard for all locations were substantially greater than differences to $\delta^{13}$C and C/N ratio for all locations. Organic content of micro-POM showed

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Fig. 3. Change in bacteria density on (A) whole leaf litter of Rhizophora stylosa and Avicennia marina; and (B) Parasearma erythodactyla faecal material, with decomposition time. Data points are means of 5 replicates (whole leaf litter) or triplicates (faecal material).

Fig. 4. Parasearma erythodactyla. Change in (A) $\delta^{15}$N; (B) $\delta^{13}$C and (C) C/N ratio values of faecal material with decomposition time. Each data point is the average of 3 independent replicates ($\pm 1$ SD $n = 15$ for all parameters).
Table 1. Mean (±1 SD) values of the chemical measurements of micro-POM. Each mean is the average from different numbers of replicates. The results of univariate mixed-model ANOVAs including a planned contrast for difference between channel and forest sites are given. OM: organic matter; NS: not significant; F: forest; E: edge; C: channel

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a general trend of highest percentage within forest sites followed by edge and then channel sites for all locations except Tingalpa Creek, which showed the opposite trajectory (Table 1), which was probably responsible for the significant location × proximity interaction.

**Value of crab-processed mangrove organic matter to pelagic copepods**

Survivorship of *Temora turbinata* differed significantly between treatments over time (repeated-measures ANOVA, $F = 10.1$, df = 3, 15; $p = 0.001$; one of the no-food treatment flasks in the *Oithona* experiment has been excluded from the comparison because of an experimental error affecting copepod survivorship), but the length of experiment was substantially longer (25 d survivorship for *O. rigida* as opposed to 12 d for *T. turbinata*) (Fig. 5B). *O. rigida* reared on *Nannochloropsis* sp. had 80% mortality by Day 2, with 100% mortality by Day 20. Analysis of the change in survivorship with time for both experiments indicated a significant treatment by time interaction effect ($p < 0.001$) for both copepod species.

Comparison of *Temora turbinata* and *Oithona rigida* isotopic signatures and the signatures of their respective diets suggests best survival was attained by those copepods with crab-processed material in their diets. *T. turbinata* copepods in the M treatment had the closest δ¹³C signatures to their diet (difference in δ¹³C = 1.32‰) and similar differences (0.80‰) in δ¹⁵N signature to their diet compared to *T. turbinata* individuals in the faecal only treatment (0.58‰) (Fig. 6A). Difference in δ¹⁵N between copepod and diet in the *Nannochloropsis* sp. only treatment was substantially larger (4.56‰), but with closer δ¹³C values for *T. turbinata*. *O. rigida* copepods in the F treatment had a difference of
1‰ to the δ13C value of their food source and, 1.2‰ for those in the M treatment. Difference in δ13C value for *O. rigida* in the A treatment was much larger at 4.2‰ (Fig. 6B). Evaluation of the relative contribution of alga and faeces in the M treatment using mixing model calculations indicated contributions from crab faeces of 77% for *T. turbinata* and 51% for *O. rigida* for nitrogen whereas dependence on crab-processed carbon was 66% *T. turbinata* and 74% for *O. rigida* (Table 2).

### DISCUSSION

Shredded leaf litter—alternative form of mangrove export?

The results of this study suggest that leaf litter processed by crabs undergoes a significantly different trajectory compared with simple microbial decomposition. Crab consumption reduced whole-leaf litter into small particulates averaging ~200 µm in diameter after digestion, which then attracted significantly faster colonisation by bacteria and resulted in a rapid decline in the C/N ratio. The significant decline in δ15N of the aged crab faecal pellet material, approaching +1‰ after 3 wk further suggests that the decrease in C/N ratio had resulted from addition of external (atmospheric) N from microbial N-fixation (Goericke et al. 1994).

Past decomposition studies of mangrove and wetland vascular plant material suggest that the δ13C signature changes are negligible with decomposition (Zieman et al. 1984, Freudenthal et al. 2001). While the change in δ13C was insignificant and comparable to that in the whole-leaf litter bag experiment, crab-processed litter underwent significantly faster chemical changes in C/N ratio, 3 orders of magnitude higher bacterial density and significant depletion of δ15N compared with whole-leaf litter. Addition of external N through microbial colonisation is thought to be a significant step in the transformation of vascular plant litter (White & Howes 1994). The shredding action of grapsid crabs predisposes mangrove leaf litter to rapid microbial colonisation, effecting significant changes in litter quality favouring meio- and macrofaunal utilisa-

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Table 2. Two-member mixing model solutions to the relative contribution of *Nannochloropsis* sp. (Alga) and *Parasesarma erythodactyla* faeces (Faeces) to the signature in *Oithona rigida* and *Temora turbinata* (Copepods) in the mixed-food (M) treatment of the feeding experiment

<table>
<thead>
<tr>
<th></th>
<th><em>T. turbinata</em></th>
<th></th>
<th><em>O. rigida</em></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>δ13C</td>
<td>δ15N</td>
<td>δ13C</td>
<td>δ15N</td>
</tr>
<tr>
<td>Copepods Faeces</td>
<td></td>
<td></td>
<td>Copepods Faeces</td>
<td></td>
</tr>
<tr>
<td>Signature (%)</td>
<td>-26.2 ± 2.4</td>
<td>-27.4 ± 0.1</td>
<td>-29.5 ± 0.66</td>
<td>-27.4 ± 0.68</td>
</tr>
<tr>
<td>(SD)</td>
<td>4.76 ± 0.23</td>
<td>4.76 ± 0.23</td>
<td>4.76 ± 0.23</td>
<td>4.76 ± 0.23</td>
</tr>
<tr>
<td>Sample size</td>
<td>66 ± 3</td>
<td>77 ± 0.1</td>
<td>65 ± 3</td>
<td>51 ± 2</td>
</tr>
<tr>
<td>Contribution (%)</td>
<td>66 ± 0.3</td>
<td>77 ± 0.1</td>
<td>65 ± 3</td>
<td>51 ± 0.02</td>
</tr>
<tr>
<td>95% Confidence</td>
<td>66–67</td>
<td>77–80</td>
<td>65–83</td>
<td>51–50</td>
</tr>
<tr>
<td>limits (%)</td>
<td>34.4–35.1</td>
<td>22–35</td>
<td>17–35</td>
<td>50–49</td>
</tr>
</tbody>
</table>

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Fig. 5. Change in survivorship with time in cultures of (A) *Temora turbinata* and (B) *Oithona rigida*, fed the 4 diets in the feeding experiments. Each data point is the average of 13 replicates for *T. turbinata* and 5 replicates for *O. rigida*, ±1 SE
tion. While microbial processing of whole-leaf litter would also involve the same general changes, only a modest decline in C/N ratio was noted for the litter bag experiment even after >110 d for both *Avicennia marina* and *Rhizophora stylosa* leaf litter.

Significant microbial colonisation often results from addition of ‘seed’ bacteria from the gut of deposit-feeders or those present in the environment, or through regrowth of bacteria surviving through the digestive process (Plante et al. 1989, Plante & Wilde 2001). Loss of feeding deterrents, such as phenolics, and dissolved organic matter, by leaching, a predominantly physical process, is also expected to be increased. Grapsid crabs generally egest about 70% of ingested leaf litter as faecal material (Lee 1997). The amount of raw mangrove leaf litter processed and then returned to the forest ecosystem in a form conducive to rapid microbial enrichment is therefore highly significant. Grapsid crabs thus play a role in mangroves similar to that of shredders in freshwater streams (Cummins 1974), a role mainly filled by insects (Wallace & Webster 1996) but occasionally also by decapods where insects are rare (e.g. Usio 2000). Early views of stream ecosystem functioning asserted that shredder consumption of vascular plant litter only follows microbial enrichment (Webster & Benfield 1986) but recent reviews conclude that such enrichment is not usually necessary and many shredders can consume seemingly nutritionally poor leaf material (Wallace & Webster 1996). Shredders physically fragment large vascular plant material and return small particulate organic matter, either as a result of ‘sloppy feeding’ or faeces, and thus, promote leaf litter breakdown rates (Jonsson et al. 2001). Release of dissolved organic matter by leaching is also enhanced by shredder activity (Meyer & O’Hop 1983, Wallace & Webster 1996). Grapsid crabs actually fit most of the expectations of shredders in ecosystems driven by vascular plant detritus input, acting as initial processors of leaf litter of low nutritional quality, physically fragmenting the leaf litter and accelerating the enrichment process.

In freshwater systems, shredders provide the food for the ‘gatherer’ and ‘filter-feeder’ functional groups (Wallace & Webster 1996), which, because of the unidirectional flow, are located downstream to the shredders. In tidal mangroves, flow of suspended materials is bi-directional at least over short time scales, e.g. over the course of a tidal cycle. It can be assumed, therefore, that a considerable portion of the shredded material processed by the crabs will be deposited on the forest floor. Grapsid crabs often spend more time feeding on surface sediments than feeding on whole leaf litter (Kwok 1999, Skov & Hartnall 2002), and the enriched faecal material deposited on the sediment surface could be a valuable food resource to the crabs, possibly explaining how grapsid crabs can
obtain enough nutrition from an apparently poor diet (the sesarmine crab Sesarma messa only assimilates 50% of the organic matter in fresh Rhizophora stylosa leaf litter, Lee 1997).

Herbivorous crabs face a number of challenges in consuming vascular plant litter (Wolcott & O'Connor 1994), and mangrove grapsids encounter additional physiological difficulties in digesting leaf litter that is usually taken fresh, i.e. before microbial enrichment. The density of bacteria on decomposing mangrove leaf litter, as indicated in the litter bag experiment, is far too low to be beneficial to the crabs. Crab-processed litter, however, typically supports bacterial densities many times higher, with consequential changes in C/N ratio, N content and δ15N. Bacteria density on crab-processed leaf fragments was ~30 times higher after only 20 d compared with the peak density reached by whole leaf litter after about 60 d. Despite the low nutritive value of fresh vascular plant litter, wetland crabs tend to maintain some consumption of plant material even when alternative, more nutritive, food sources are available (Buck et al. 2003).

By consuming some whole leaf litter, the crabs start an enrichment process that may generate more nutritious food through feeding on the surface sediment. This may represent a feeding mode similar to the ‘gardening’ behaviour of many deposit-feeders, providing a renewable food supply through the feeding activities of the organisms themselves (Rice & Rhoads 1989, Levinton 1995). Coprophagy is probably an important aspect of the feeding strategy of grapsid crabs in tropical mangroves, while their shredding activities may also benefit other components of the estuary. The dependence of grapsid crabs on food resources other than fresh mangrove leaf litter is reflected by the commonly observed, more enriched δ13C (~ −24‰) signature relative to that of mangrove litter (−27‰) (Rodelli et al. 1984, Lee 2000, Bouillon et al. 2002).

**Prevalence of mangrove micro-POM export in estuarine waterways**

A few studies have attempted to determine the origin of estuarine POM using chemical tracers (e.g. Thornton & McManus 1994, Middelburg & Nieuwenhuize 1998, Maksymowska et al. 2000, Riera et al. 2000). Thornton & McManus (1994) used a combination of δ13C, δ15N and C/N indicators to trace the origin of organic matter along the Tay Estuary in Scotland. These authors found that only δ13C seemed to give a reliable indication, as the other two were likely to be modified by diagenetic processes.

Survey of the micro-POM at 5 mangrove-lined waterways in southern Moreton Bay suggested that the impact of mangrove export is often overridden by other including anthropogenic sources. Only the δ13C value of micro-POM was significantly influenced by proximity to mangrove forests. The δ13C of mangrove leaf litter changed little with crab or microbe-mediated decomposition (~27 to ~28‰). Values of all other including δ15C indicators varied significantly amongst waterways but not with proximity to mangrove forests. Even samples from the ‘forest’ locations (range −25.5 to −19.9‰), which were expected to have the highest concentration of mangrove-derived POM, bore little direct correspondence to the δ13C signatures characteristic of either senescent or crab-processed mangrove leaf litter. Usually gradients in δ13C exist along the land-estuary continuum, with locations influenced by terrestrial sources demonstrating more depleted values (Canuel et al. 1995), and plankton material dominates shallow water bodies (Kennedy et al. 2004). The highly enriched δ15N (≥+9‰) signature of micro-POM from all 5 waterways still fall within those expected of estuarine environments (Middelburg & Nieuwenhuize 1998) but suggests relatively minor direct input from either whole or crab-processed mangrove leaf litter, but significant contribution from phytoplankton, microalgal (δ13C range −18 to −22‰) or anthropogenic sources. Sewage N and animal excreta are known to have relatively enriched δ15N (~+10‰, Macko & Ostrom 1994) and anthropogenic dissolved inorganic nitrogen (DIN) resulting from wastewater input. The highly enriched δ15N recorded throughout the waterways in southern Moreton Bay is probably a reflection of contamination from sewage and agricultural sources, diminishing the significance of mangrove-derived organic matter in the populous suburban embayment.

The C/N ratio of the micro-POM (≤9) is also too low to be of mangrove leaf litter origin. Decomposition of crab-processed leaf litter in the laboratory improved the C/N ratio to <20 after 25 d but is still much higher than the observed value in the waterways. Low C/N ratios are in fact typical of micro-POM in estuarine waterways and estuarine sediments (e.g. Canuel et al. 1995, Middelburg & Nieuwenhuize 1998, Kennedy et al. 2004), reflecting the dominance of the seston by bacterioplankton, which have low C/N ratios (~5 to 7, Kirchman 2000).

**Utilisation of crab-processed mangrove leaf litter by copepods**

The connection between mangrove production and nearshore secondary production has often been assessed based on estimates of tidal export of unprocessed leaf litter. Lee (1997), investigating the value of
crab faecal material to a ‘detritivorous’ amphipod *Parhyalella* sp., suggested that crab-processed mangrove leaf litter could fuel a coprophagous food chain and contribute to nearshore secondary production in a different way.

In our present study, aged crab-processed mangrove material in the form of disintegrated crab faecal pellets supported higher survivorship in 1 common nearshore (*Temora turbinata*) and 1 common estuarine copepod species (*Oithona rigida*). Copepods are ubiquitous in mangrove waterways (Robertson & Blaber 1992) and often form the main food source for larval and juvenile fish. Stable isotope analysis suggested that the copepods were utilising the mangrove-derived carbon for food. Contrary to findings on a congener (*Oithona longicornis*, Klein Breteler et al. 2002), *T. turbinata* demonstrated depletion of δ¹³C in their body biomass compared with their food source, and the degree of fractionation seemed to be dependent on the food items. A δ¹³C of ~4‰ was recorded for the F treatment whereas it was less than ~2‰ in the M and A treatments. This pattern of C and N isotopic fractionation was also recorded for *O. rigida*, suggesting that isotopic fractionation patterns may be species- and food-dependent. This finding questions the application of simple and fixed fractionation values to analysing trophic relationships.

Reproduction in copepods is known to be dependent on food quality (e.g. Rey-Rassat et al. 2002). Although most of the treatments did not result in production of a second generation, it was observed that eggs and nauplii larvae were present in the treatments where crab faecal material was supplied. The high percentage of leaf litter that can be processed by grapsid crabs before tidal export in tropical mangroves (Lee 1998) implies that crab-processed mangrove leaf material can be a more important form of export supporting nearshore food chains.

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