Effect of temperature and salinity on otolith element incorporation in juvenile gray snapper 
*Lutjanus griseus*

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**ABSTRACT:** Otolith chemistry provides one approach for identifying the relative contribution of juveniles from different nursery habitats to adult populations. The goal of this study was to validate otolith element incorporation by quantifying the relation between otolith and water element concentrations (Sr/Ca, Mg/Ca, Mn/Ca, and Ba/Ca) as a function of differences in water temperature and salinity using juvenile gray snapper *Lutjanus griseus*, a reef fish that inhabits estuarine and near-shore habitats as juveniles. We investigated the effects of 20 different temperature (18, 23, 28, 33°C) and salinity (5, 15, 25, 35, 45) combinations on otolith element incorporation (partition coefficient $D$) in *L. griseus*. Temperature and salinity had significant effects on $D_{Sr}$ but no significant effect on $D_{Mg}$ or $D_{Mn}$; however, salinity had a significant effect on $D_{Ba}$. The broad range of temperatures and salinities used in the present study encompasses those occupied by juveniles in the wild and therefore provides a realistic test for using otolith chemistry to infer environmental history of individual gray snapper. Element incorporation and the effects of temperature and salinity on element incorporation differ among fish species, limiting development of generalized models aimed at predicting water chemistry from otolith chemistry. Thus, the data presented here underscore the necessity of validation experiments to translate species-specific elemental signatures in otoliths.

**KEY WORDS:** Otolith chemistry · Salinity · Temperature · Strontium · Barium · Manganese · Magnesium · *Lutjanus griseus*
Rooker et al. 2001). Elemental signatures in fish otoliths can also be used to infer transport or migration pathways for individual fish (Secor 1992). Key assumptions underlying these applications are that fish incorporate elements from their environment and these elements are permanently deposited in their otoliths (Campana 1999).

While some studies have tested the effect of water temperature and salinity on element incorporation in otoliths (Farrell & Campana 1996, Chesney et al. 1998, Bath et al. 2000, Milton & Chenery 2001, Elsdon & Gillanders 2002, 2004, Martin et al. 2004, Martin & Thorrold 2005), the specific behavior of individual elements remains unclear. Elemental signatures in otoliths have been used successfully as natural tags in some species; however, important assumptions that influence the unambiguous use of these signatures have not been validated. For example, species-specific differences in elemental incorporation have not been evaluated in the laboratory under controlled conditions, and for most species the potential effect of differences in salinity or water temperature is unknown. Although understanding the details of elemental incorporation is not necessary for all applications of otolith elemental signatures, physical and biological factors can influence concentrations of these elements in the otolith, rendering the interpretation of elemental signatures more complex than assumed.

Gray snapper *Lutjanus griseus*, a reef fish whose juveniles use estuarine and near-shore habitats, can tolerate and grow over a wide range of temperatures and salinities (Wuenschel et al. 2004), making the species a good model for examining the effects of temperature and salinity on otolith element incorporation. Although previous experiments on gray snapper have demonstrated strong effects of temperature and salinity on feeding, growth, and metabolism (Wuenschel et al. 2004, 2005), the ultimate ecological consequences of inhabiting different nursery habitats and environments remains unclear. Elemental signatures in otoliths of gray snapper may provide a tool to evaluate the relative contribution of juveniles from different nursery habitats (sensu Gillanders & Kingsford 1996). The goals of this study were to validate element incorporation in gray snapper *L. griseus* otoliths by quantifying the relation between otolith and water element concentrations (Sr/Ca, Mg/Ca, Mn/Ca, and Ba/Ca) as a function of differences in water temperature and salinity.

**MATERIALS AND METHODS**

Gray snapper *Lutjanus griseus* were collected on a rising tide from the channel spanned by Pivers Island Bridge, near Beaufort, North Carolina (USA), approximately 2 km inside of Beaufort Inlet. Larvae were collected using a 1 × 2 m neuston net (1 mm mesh) with a floating live box attached to the cod end, fished at the surface (Hettler 1979). Sampling was conducted during nighttime maximum flood tides and concentrated around the days preceding full and new moons (Tzeng et al. 2003). Snapper were gently dipped from the live box and transferred to 100 l holding tanks with flow-through seawater for temperature acclimation (1°C d−1).

A fully crossed 2-way factorial design with equal replication was used to investigate the effects of 20 different temperature and salinity combinations on otolith element incorporation in juvenile gray snapper. A total of 5 tanks at each temperature and salinity combination were used (total n = 100, 5 replicates × 5 salinities × 4 temperatures). Temperature (18, 23, 28, and 33°C) and salinity (5, 15, 25, 35, and 45) levels were chosen to represent the range of environmental conditions experienced by gray snapper during the juvenile stage. Experiments were carried out in 2 trials due to limitations in laboratory space and availability of specimens. Owing to this constraint, a design was chosen that included all salinity levels within each trial. The first trial included the 28 and 33°C treatment temperatures and fish collected 5 to 6 d preceding the September new moon (1 to 5 September 2002). The second trial included the 18 and 23°C temperature treatments and the fish collected in the 5 d preceding the October new moon (30 September to 5 October 2002).

Individual gray snapper were stocked in experimental tanks (8 l) once the desired temperature treatment levels were attained. The 4 salinity levels were randomly assigned to the experimental tanks, and salinity in the experimental tanks was adjusted to 5 d−1 until the desired salinity treatment levels were reached. Salinity was maintained by mixing filtered seawater with either conditioned well water or Instant Ocean® synthetic sea salt. When all fish were at the desired treatment levels, they were acclimated for 1 wk before the start of the 55 d experiment.

Snapper were fed several times daily, with diet changing from recently hatched *Artemia nauplii* to a prepared gel diet and live larval fish (*Eucinostomus* sp. or guppies) as fish grew throughout the experimental period. The same diets were offered to all fish within a given trial each day to reduce the effects (if any) of diet on otolith element incorporation. A one-third volume water change was performed daily to prevent the buildup of metabolic wastes and to maintain the desired salinity levels. Tank water temperatures and salinities were measured daily throughout the acclimation and experimental periods.
Water samples. Water samples (n = 5 per tank) were taken weekly for analysis of elemental water composition. Samples were collected using acid-washed 10 ml polypropylene syringes and filtered through 0.2 µm polypropylene syringe filters (Whatman) into 7 ml acid-washed polypropylene vials. Each sample was acidified to pH 2 with ultrapure HClconc and stored frozen until subsequent analyses.

Otolith analyses. At the conclusion of each experimental trial, all surviving fish were measured and their sagittal otolith pairs removed for otolith analyses. Otoliths were scraped clean with acid-washed glass probes in a class-100 clean room. Otoliths were ultrasonically cleaned in Milli-Q water for 7 min and triple rinsed with ultrapure H2O2 (Ultrex II, purchased from J. T. Baker) followed by 3 sequential rinses of Milli-Q water. Otoliths were then placed on acid-washed glass slides to dry for 36 h under a class-100 laminar-flow hood. After drying, otoliths were stored in acid-washed 1.5 ml high-density polyethylene vials. The left otolith of each pair was mounted on a petrographic slide with superglue™ and polished along the sagittal plane. After polishing, otoliths were soaked in Milli-Q water, cleaned, and dried as described above. Finally otoliths were mounted on petrographic slides (21 per slide) for high-resolution, inductively coupled plasma mass spectrometer (ICP-MS) analyses.

Because the fish for the 2 trials were live-captured at different times, they may have experienced different temperatures and salinities during the larval period. For that reason, laser ablation was used rather than whole otolith dissolution. The portion of the otolith was sampled that corresponded in time to the period during which fish were exposed to the experimental conditions (>30 d old). Curvilinear paths (800 µm) along growth increments during the experimental period were plotted with the laser software and ablated on each otolith (Fig. 1) using a New Wave UP-213 laser with a 40 µm beam width coupled to a Thermo Finnigan Element ICP-MS equipped with a self-aspirating (50 µl min−1) PFA nebulizer (Elemental Scientific, Inc. [ESI]) and a dual-inlet quartz spray chamber. Due to the presence of significant interferences on most of the Ca isotopes, 44Ca, 88Sr 25Mg, 55Mn, 137Ba, and 115In were measured in medium resolution (nominal R = 4500). A total of 3 of the 5 weekly samples from each tank were analyzed and the mean values were then used in all subsequent analyses. To estimate precision of the water measurements, Ca, Sr, Mg, Mn, and Ba values in a seawater Certified Reference Material (CRM), (High Purity Standards, Inc.) were determined. The estimates of precision for element concentrations in the seawater CRM for Trial 1 were 2.1% RSD for Ca, 4.2% RSD for Sr, 1.7% RSD for Mg, 12.2% RSD for Mn, and 4.5% RSD for Ba (n = 3) and Trial 2 were 1.4% RSD for Ca, 1.3% RSD for Sr, 1.3% RSD for Mg, 6.8% RSD
for Mn, and 3.9% RSD for Ba (n = 3). Partition coefficients (DMe) were calculated by dividing the metal/calcium (Me/Ca) ratio measured in an otolith by the mean Me/Ca ratio measured in the treatment tank water (Morse & Bender 1990).

Statistical analyses. Two-way analysis of variance (ANOVA) was used to test for significant differences in [Me/Ca]water, [Me/Ca]otolith, and DMe among temperature and salinity treatments. Because different cohorts of fish and temperatures were used in the 2 trials, a separate ANOVA was performed for each trial (Bonferroni corrected α = 0.0125). Salinity and temperature were treated as independent categorical variables, and [Me/Ca]water, [Me/Ca]otolith, and DMe as dependent variables in the analyses. [Me/Ca]otolith data were log-transformed to reduce heteroscedasticity, lessening the magnitude of differences among treatment groups. The data transformations did not make all of the variances homogeneous, but considering the F statistic is robust despite differences in sample variances (Lindman 1974), we feel confident in the subsequent ANOVAs. Because DMe are proportions, data were arcsin-transformed (Zar 1999).

RESULTS

The means and ranges of [Me/Ca]otolith, and DMe from 90 (Trial 1: n = 45, Trial 2: n = 45) juvenile Lutjanus griseus and the tank [Me/Ca]water, (n = 90) are reported in Table 1. Survival of reared L. griseus was high (90%) over the range of experimental treatment combinations, and the few mortalities (10 out of 100) were not related to temperature or salinity over this range, as in other studies (Wuenschel et al. 2004, 2005).

Water chemistry

[Me/Ca]water values were variable for each element analyzed (Table 1). As expected, elemental concentrations in the tank water were significantly affected by salinity treatments for both trials, but not temperature (Table 2).

Otolith [Me/Ca]

[Me/Ca]otolith values were also variable for each element analyzed (Table 1). Temperature (Trial 2) and salinity (Trial 1) had significant effects on [Sr/Ca]otolith for Lutjanus griseus without a significant interaction effect.
effect on [Mg/Ca]_{otolith} or [Mn/Ca]_{otolith} (Table 3, Fig. 3). There was a significant salinity effect (Trial 1) but no significant temperature effect on [Ba/Ca]_{otolith} (Table 3, Fig. 3), and the interaction between the 2 factors was not significant.

**Me/Ca partition coefficients**

Temperature (Trial 2) and salinity (Trial 1) had significant effects on $D_{Sr}$, with no interaction between the 2 factors (Table 4, Fig. 4). The effect of temperature was significant at the higher temperature treatments (18 and 23°C), and the effect of salinity was significant only at the higher temperature treatments (28 and 33°C). The effects of temperature, salinity, and the temperature–salinity interaction on $D_{Mg}$ and $D_{Mn}$ were not significant (Table 4, Fig. 4). Salinity had a significant effect on $D_{Ba}$ in both trials, with a non-significant temperature effect and a non-significant interaction between the 2 factors (Table 4, Fig. 4).

**DISCUSSION**

For a given species, otolith element incorporation is not solely a function of water elemental composition because it is uniquely influenced by temperature and salinity for each element investigated. Application of otolith elemental signatures to determine nursery habitats assumes that differences observed in otolith chemistry reflect differences in the water chemistry of various nursery habitats. The realization that otolith element incorporation can be influenced by abiotic (temperature and salinity) and biotic (growth rate and diet) factors indicates the need to quantify the relative importance of these effects for individual species and elements. The relative magnitude of these effects (which may be statistically significant) may not result in misclassification of individuals; however, it may prove useful in explaining variation in specific elements and determine the precision of inferring environmental histories. The main objective of this experiment was to determine the effect of temperature and salinity on element incorporation in the otoliths of juvenile gray snapper to assess the usefulness of otolith element signatures for inferring life history information such as nursery habitat use for this species. The broad range of temperatures and salinities used in the present study encompass those occupied by juveniles *Lutjanus griseus* in the wild (Starck & Schroeder 1970, Chester & Thayer 1990, Nagelkerken et al. 2000) and therefore provide a realistic test for using otolith chemistry to infer environmental history of individuals.

Few laboratory-based validation experiments have been used to address otolith element incorporation for different species. One of the only other laboratory experiments to address the effect of temperature and salinity on Sr/Ca, Mg/Ca, Mn/Ca, and Ba/Ca incorporation in otoliths (Elsdon & Gillanders 2002) used both single-factor and 2-factor designs. They found significant temperature and salinity interaction effects on all 4 Me/Ca ratios in juvenile black bream *Acanthopagurus butcheri*. If the assumption that otolith element incorporation is proportional to the water elemental composition is correct, then the most appropriate way to discern interspecies differences would be to compare partition coefficients. Elsdon & Gillanders (2002) did not calculate partition coefficients, but best estimates of the ranges (for 12 to 24°C and 5 to 30 salinity) made by inspection of their graphs and water data table are reported in Table 5 for $D_{Sr}$, $D_{Mg}$, $D_{Mn}$, and $D_{Ba}$. Using the results from this study for *Lutjanus griseus* (for 18 to 33°C and 5 to 45 salinity) and that of Martin et al. (2004) and Martin & Thorrold (2005) for *Leiostomus xanthurus*
(for 17 to 26°C and 15 to 25 salinity), the $D_{Sr}$ values for $A. butcheri$ were higher than the values for both $L. xanthurus$ and $L. griseus$. Whereas the $D_{Mn}$ values for $A. butcheri$ were similar to $L. xanthurus$ and only slightly higher than $L. griseus$, $D_{Mn}$ values were similar between $A. butcheri$ and $L. xanthurus$ but were 1 order of magnitude greater than those calculated for $L. griseus$. Similarly, $D_{Mg}$ ranges were the same for $A. butcheri$ and $L. xanthurus$ but higher than $D_{Mg}$ in $L. griseus$. Differences in ontogeny do not explain the varied results of temperature and salinity effects on otolith element incorporation because the experiments were conducted with fish at the same life stage (late larval-early juvenile). Diets fed to fish for each experiment were different, which may explain some of the variability in relative $\text{[Me/Ca]}_{\text{otolith}}$ values, although experimental studies have found little evidence for an effect of diet on otolith element incorporation (Hoff & Fuiman 1995, Milton & Chenery 2001, Walther & Thorrold 2006, but see Buckel et al. 2004). These discrepancies highlight the potential for species-specific element incorporation. Mechanisms controlling the uptake of individual elements into the otolith matrices differentially influence the partition coefficients.

Species may be different with regard to otolith chemistry because they have different environmental tolerances and employ different methods to maintain hydraulic balance. The present study investigated a wider range in salinity than previous studies (see Elsdon & Gillanders 2003 for review), which were both below and above the average isosmotic salinity for marine teleost fishes (~10 to 15; Karnaky 1998). Euryhaline fishes employ different extrarenal epithelial tissues (including the gills, integument, gut, and bladder) to

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**Fig. 2.** Calculated metal/calcium ratios in the tank water for both trials: (a) Sr/Ca, (b) Mg/Ca, (c) Mn/Ca, and (d) Ba/Ca concentrations ±1 SE by salinity treatment. Each of the 4 temperature treatments is represented by a different symbol: 18 (●), 23 (○), 28 (▼), and 33 (▽) °C. Data markers are offset so that all error bars and symbols can be seen clearly.
achieve osmoregulation depending on whether they are above or below the isosmotic salinity. Ionic balance may be achieved through a combination of passive and active transport. Therefore the pathways and potential barriers between the external environment and the fish are not constant over this range in salinity, which may confound the relationship between element concentrations in the external environment and otolith. Thus, in addition to being species and element specific, $D_{Me}$ relationships for euryhaline fishes are likely to be different in hyposmotic versus hyperosmotic environments.

Somatic growth rate and otolith precipitation rate may also influence element incorporation, as demonstrated for [Mg/Ca]$_{otolith}$ in *Lutjanus xanthurus* (Martin & Thorrold 2005). It is well documented that slower growing fish have proportionately heavier otoliths (Reznick et al. 1989, Secor & Dean 1989, Pawson 1990, Worthington et al. 1995); therefore, growth rate can influence both the amount and relative proportion of elements incorporated in the matrix. Thus, somatic growth rate may act to modify the relative importance of water chemistry, diet, temperature and salinity, and internal physiology on otolith element incorporation. The effect of somatic growth rate on element incorporation may be particularly important for larval and juvenile fishes that exhibit higher (and more variable) growth rates compared to later life stages. The ad libitum feeding condition and wide range of temperatures in the present study resulted in a wide range of growth rates, largely determined by temperature, thus precluding a test of growth rate effects. To determine the effect of growth rate on element incorporation, an experimental design with various feeding (growth) rates within each temperature and salinity combina-

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**Fig. 3. Lutjanus griseus.** Mean (a) Sr/Ca, (b) Mg/Ca, (c) Mn/Ca, and (d) Ba/Ca ±1 SE in otoliths of *L. griseus* for both trials as a function of salinity at tank temperatures 18 (●), 23 (○), 28 (▼), and 33 (▼)°C. Data markers are offset so that all error bars and symbols can be seen clearly.
tion would be necessary. The influence of growth rate on otolith element incorporation may be obscured in gray snapper, which have been shown to alter their energetics (feeding rate, gross conversion efficiency, and metabolism; Wuenschel et al. 2004, 2005) in relation to salinity, yet salinity had little effect on growth. Subtle changes in the energetics of gray snapper in response to salinity affect the proportion of consumption that is translated to somatic growth. It remains uncertain how changes in gross conversion efficiency affect the influence of diet on otolith element incorporation. Likewise, changes in metabolism may require more (or less) water to come in contact with gills for gas exchange and thereby affect the influence of water chemistry on otolith element incorporation.

Species-specific differences in element incorporation in otoliths make it difficult to generalize results among taxa, but perhaps of greater importance to studies using these techniques are the marked differences in element incorporation attributable to changes in temperature and salinity. For example, studies on eels (Tzeng et al. 1997, Jessop et al. 2002, Daverat et al. 2006), American shad (Limburg 2001) and striped bass (Secor 1992, Secor et al. 1995) have used [Sr/Ca]otolith as proxies for salinity changes in the water. These studies interpreted fish migration paths based primarily on salinity differences from Sr/Ca signatures in the otoliths. However, conclusions about fish migrations based on the [Sr/Ca]otolith–salinity relationship are potentially confounded by the effect of temperature on [Sr/Ca]otolith incorporation. The magnitude of change in [Sr/Ca]otolith observed in the present study, due to temperature and salinity (1.72 to 2.88) is much less than the range in values generally used to infer migration in previous studies, which was 0 to 10 for eels (Daverat et al. 2006), 0 to 3 in American shad.

Fig. 4. *Lutjanus griseus*. Mean partition coefficients (a) Sr/Ca (D<sub>Sr</sub>), (b) Mg/Ca (D<sub>Mg</sub>), (c) Mn/Ca (D<sub>Mn</sub>), and (d) Ba/Ca (D<sub>Ba</sub>) for otoliths of *L. griseus* for both trials as a function of salinity at tank temperatures 18 (●), 23 (○), 28 (◇), and 33(■)°C. Data markers are offset so that all error bars and symbols can be seen clearly.
Bath Martin & Wuenschel: Otolith element incorporation in gray snapper

(2001), 0 to 3 in striped bass (Secor et al. 2001) and 0 to 5 in sea-run brown trout (Limburg et al. 2001). Although the range of \([\text{Sr/Ca}]_{\text{otolith}}\) varies among species, it can be reliably used to infer habitat residence among systems that have marked differences in water chemistry (underlying geology) (Munro et al. 2005). The same \([\text{Sr/Ca}]_{\text{otolith}}\) could result from cool, salty water or warmer, fresher water, although the relative magnitude of these effects within a given system (estuary) are likely to be less than the differences among systems (estuaries). The interaction between temperature and salinity, however, does complicate the prospect of retrospectively identifying individual fish transport pathways across heterogeneous water chemistries (i.e. estuaries) as a function of their otolith elemental signatures through time. Although \([\text{Sr/Ca}]_{\text{otolith}}\) alone may not be a reliable predictor of water temperature and salinity, when used in combination with other metals (i.e. \([\text{Ba/Ca}]_{\text{otolith}}\), it may be possible to use more precise predictive models to retrospectively identify individual fish transport pathways as a function of their otolith elemental signatures through time.

Field studies have been able to use a suite of elements to assign individual fish to unique nursery habitats (Gillanders & Kingsford 1996, Thorrold et al. 1998, 2001). Detected differences are used to assign fish to their respective nursery areas, even though the mechanisms of incorporation or the influence of temperature and salinity are poorly understood. Given the complexity of the relationship between the water and the otolith, it would be difficult to discern temperature or salinity experienced by individual fish. Nevertheless, these results do not invalidate the approach of retrospectively classifying experienced environments; if different otolith signatures are found, then different environments were inhabited. The differences in water chemistry in the present study, which used filtered seawater from a single system mixed with well water and synthetic sea salts, are likely to be less than the differences in water chemistry experienced by fishes occupying different estuaries. Although individual species incorporate trace metals in their otoliths in different quantities (in absolute terms, e.g. Table 5), otolith chemistry is a function of water chemistry and as such can be used to infer previous environmental (habitat) history among environments (habitats) that differ in water chemistry, particularly when multiple elements are analyzed to differentiate otolith signatures. In cases where validation is impractical (endangered species) or impossible (historic samples/forensic applications), otolith chemistry can still provide useful information.

Table 4. *Lutjanus griseus*. Results of 4 separate 2-way ANOVA for both trials testing the effect of temperature (T) and salinity (S) on 4 partition coefficients \(D\) (arcsin-transformed) in the otoliths of juvenile gray snapper *Significant at \(\alpha = 0.0125\). Trial 1: 28–33°C, Trial 2: 18–23°C

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<td></td>
<td>S</td>
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Table 5. Comparison of partition coefficient \(D\) ranges for Sr, Mg, Mn, and Ba, for tested temperature and salinity ranges, as reported in this study for *Lutjanus griseus*, as estimated from Elsdon & Gillanders (2002) for *Acanthopagurus butcheri*, and from Martin et al. (2004) and Martin & Thorrold (2005) for *Leiostomus xanthurus*

<table>
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<th>Salinity range</th>
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<th>(D_{\text{Mg}})</th>
<th>(D_{\text{Mn}})</th>
<th>(D_{\text{Ba}})</th>
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<td>5–45</td>
<td>0.21–0.38</td>
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<td>5–30</td>
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</tbody>
</table>
The present study demonstrates the complex and poorly understood mechanisms driving the [Me/Ca]_{otolith} to [Me/Ca]_{water} relationship. Element incorporation and the effects of temperature and salinity on element incorporation differ among fish species, limiting the development of generalized models to predict water chemistry from otolith chemistry. Therefore, the data presented here underscore the necessity of validation experiments to translate species-specific elemental signatures in otoliths.

Acknowledgements. This research was supported by grants from the NOAA Sea Grant to S. Thorrold, J. Hare, S. Sponaugle, and L. Barbieri, the NOAA South Florida Ecosystem Research and Monitoring Program funded jointly by NMFS SEFSC and NOS NCCOS, a grant in aid from the International Women’s Fishing Association, and a PEO International Women’s Organization Scholar Award to G.B.M. We thank Z. Chen (LITER, Old Dominion University) for assisting with the water samples, A. Jugovich (NOAA, National Ocean Service, CCFHR, Beaufort, North Carolina) for snapper rearing and tank maintenance, and S. Thorrold and J. Fitzgerald (Woods Hole Oceanographic Institution, Woods Hole, Massachusetts) for assisting with sample preparation and running the laser and ICP-MS. We also thank J. Hare, D. Ahrenholz, L. Settle, P. Marraro, B. Wells, C. Grimes, and 2 anonymous reviewers for comments that helped to significantly improve the manuscript. The views expressed herein are those of the authors and do not necessarily reflect the views of NOAA or any of its subagencies.

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Editorial responsibility: Kenneth Sherman (Contributing Editor), Narragansett, Rhode Island, USA

Submitted: August 6, 2005; Accepted: March 28, 2006
Proofs received from author(s): October 2, 2006