

Experimental evidence of fatty acid limited growth and survival in Pacific cod larvae

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ABSTRACT: Changing environmental conditions in the North Pacific are altering the lipid/fatty acid (FA) composition of zooplankton assemblages, but the consequences to resident fish larvae are unknown. In the laboratory, we reared Pacific cod *Gadus macrocephalus* larvae over 4 wk on prey enriched with varying levels of 2 essential FAs (docosahexaenoic acid, DHA, 22:6 ω -3, and eicosapentaenoic acid, EPA, 20:5 ω -3) to determine how this species responded to such changes in prey quality. Ratios of DHA:EPA were chosen to represent the natural variation observed in zooplankton of the North Pacific. We tested the hypotheses whether (1) energetically similar diets comprised of varying levels of DHA and EPA affect growth and survival in Pacific cod larvae, and (2) the highest levels of DHA:EPA (2:1) are optimal for Pacific cod larvae, as it has been shown for Atlantic species. Pacific cod larvae grew fastest with diets containing high levels of ω -3 polyunsaturated fatty acids (PUFA > 22%). Diets with the same total lipid content but different DHA:EPA ratios (<0.1:1 to 2:1) also mediated growth and lipid composition of the larvae. Unlike Atlantic cod, Pacific cod larvae did not show as high a requirement for DHA relative to EPA but rather achieved largest size-at-age with intermediate DHA:EPA ratios (0.8:1 to 1.1:1). This range most closely resembled DHA:EPA ratios reported from North Pacific copepods, suggesting anomalous years with an over- or under-abundance of DHA-rich dinoflagellates or EPA-rich diatoms may be detrimental to survival and growth of Pacific cod larvae in the field.

KEY WORDS: *Gadus macrocephalus* · Essential fatty acids · DHA · EPA · Prey quality

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INTRODUCTION

Prey quality is an important but poorly understood factor regulating growth and survival in larval fish (Cushing 1990, Munk 1997, Beaugrand et al. 2003). Changing climate and emerging ocean acidification have the potential to affect fish communities through large-scale changes in planktonic communities. Changes in dietary quality for larval fish can be manifested both through changes in available zooplankton populations or through compositional change in specific nutritional components (i.e. essential fatty acids) within a given zooplankton prey species such as copepods. In the North Pacific, zooplankton species composition changes dramatically between warm and cold years (Batten & Welch 2004), with cold-years typified

by large boreal copepods and warm years characterized by higher numbers of small, southern species (Mackas et al. 2007). Further, biochemical changes in a major North Pacific copepod species, *Neocalanus plumchrus*, have been correlated with changes in phytoplankton species abundance and have resulted in shifts in zooplankton fatty acid (FA) composition (El-Sabaawi et al. 2009a). However, while qualitative changes in the planktonic community such as these are hypothesized to impact growth and survival of marine fish larvae, they have rarely been examined explicitly. This is largely due to the difficulty in determining *a priori* which qualitative components of prey are limiting (e.g. prey size, energy, proximate composition, etc.) and the difficulty in manipulating those conditions experimentally for marine fish larvae (Rainuzzo et al. 1997).

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From aquaculture studies, lipids and FAs are considered to be a limiting factor in determining prey quality for cold-water marine fish, as they play a vital role both as a source of energy and as important structural components of cell membranes (Sargent et al. 1989, Arts et al. 2001). In particular, polyunsaturated fatty acids (PUFAs) have been shown to be a critical component of larval fish nutrition as they affect growth, survival, metamorphosis and pigmentation in many species (Wantanabe 1993, Sargent et al. 1999, Copeman et al. 2002).

Two PUFAs, docosahexaenoic acid (DHA, 22:6 ω -3) and eicosapentaenoic acid (EPA, 20:5 ω -3), are abundant in organisms found in cold-water marine ecosystems, but are considered essential fatty acids (EFAs) to marine fish as they cannot be synthesized in adequate amounts from short chain precursors. Marine larval fish must therefore rely on dietary input of DHA and EPA stemming from primary production (Sargent 1995, Arts et al. 2001, Budge et al. 2001, Copeman & Parrish 2003). Levels of EPA in plankton have been correlated with diatom production whereas DHA is found at higher proportions in dinoflagellates (Dunstan et al. 1994, Parrish et al. 2000, Stevens et al. 2004). Although both FAs have been found to be essential to marine fish larvae, many species have shown membrane specificity to be higher for DHA than EPA (Rodriguez et al. 1997, Copeman et al. 2002). Given that DHA is naturally found at high levels in neural tissue, it is thought to play a specialized role in neural cell membrane structure and function (Bell & Dick 1991). Therefore, higher dietary EPA in comparison to DHA is postulated to have a negative effect on larval neural function and, consequently, on growth and survival (Bell et al. 1995, Rodriguez et al. 1997).

Together with arachidonic acid (AA, 20:4 ω -6), EPA is also an important substrate for the formation of biologically active localized hormones such as prostaglandins (Sargent et al. 1999). Localized hormones have been indicated to be important in a wide range of fish physiological processes such as stress responses, ionic regulation and pigmentation development (Sargent 1995). EPA and AA are both substrates for the formation of eicosanoids, with AA being the preferred one and producing eicosanoids of high biological activity (Bell et al. 1994). EPA produces eicosanoids of lower biological activity and hence modulates the efficiency of AA. Therefore, it is often important to consider the ratio of DHA:EPA:AA as has been indicated in recent nutritional larval fish studies on Atlantic species (Copeman et al. 2002, Garcia et al. 2008a).

Generally, a DHA:EPA ratio of 2:1 in larval fish diets is cited as being optimal for growth and survival (Sargent 1995), largely based on egg composition of Atlantic species. However, Saito & Kotani (2000) found FA profiles of wax esters from 4 North Pacific copepod

species that had low DHA:EPA ratios (0.2:1 to 0.4:1). Recently, El-Sabaawi et al. (2009a) showed that DHA:EPA ratios from the North Pacific were lower than those from the Atlantic, but varied considerably depending on the relative abundance of EPA-rich diatoms and DHA-rich dinoflagellates (0.3:1 to 1.1:1). Interestingly, Laurel et al. (2010) found that DHA:EPA ratios in Pacific cod eggs were 1.4:1, i.e. lower than the 2:1 reported from their Atlantic congeners. The degree to which low DHA:EPA ratios affect Pacific marine fish larvae remains uncertain, largely because the majority of cold-water larval fish nutrition is based on Atlantic aquaculture species (Sargent et al. 1999) with little comparison to wild zooplankton assemblages (St. John et al. 2001).

Here we designed a laboratory experiment to examine how changes in DHA and EPA affected the growth and survival in the larvae of a Pacific cold-water fish, Pacific cod *Gadus macrocephalus*. This species was chosen because it (1) is highly abundant and plays an important role in predator-prey dynamics in the North Pacific (Hunt et al. 2002) and (2) makes an interesting comparison with its well-studied Atlantic congener, *Gadus morhua*. We tested the hypotheses whether (1) energetically similar diets comprising varying levels of EPA and DHA influence size-at-age and survival in Pacific cod larvae, and (2) high levels of DHA:EPA (e.g. 2:1) are optimal for marine fish larvae in the Pacific as it has been shown for Atlantic species. We discuss our results in relation to natural variation in the lipid and FA composition of larval prey in the North Pacific.

MATERIALS AND METHODS

Experimental design. Four rotifer enrichments were formulated with varying levels of the essential fatty acids DHA and EPA, and thus variable DHA:EPA ratios (Dr. Moti Harel, Advanced BioNutrition). Enriched rotifers were harvest twice daily throughout the experiment and fed to cod larvae for 4 wk. During this time we measured changes in lipid composition of the larvae while simultaneously monitoring size-at-age and survival.

Rotifer emulsions. Of the 4 experimental emulsions, Diet 1 was high in monounsaturated fatty acids (MUFAs) and served as a control, while the experimental Diets 2, 3 & 4 were high in PUFAs (Table 1). The target ratios of DHA:EPA in rotifers ranged from < 0.1:1 (Diet 1) to 2:1 (Diet 4). The 3 PUFA emulsions were formulated by blending different ratios of algae oil (DHASco-S™) and cod liver oil. The DHA-rich algal oil was extracted from the heterotrophically grown *Schizochitrium* sp. (Martek BioSci, Harel et al. 2002).

The FA composition of DHAsco-S was 27 % DHA and no EPA while cod liver oil contained ~20 % DHA and 30 % EPA (information provided by manufacturer). The control emulsion was prepared using only olive oil, which was low in PUFA. A mixture of 5 % lecithin, 1 % vitamin E, 1 % ascorbic acid and 1 % Tween-80 (w:oil weight) was added to the oils. Oil mixtures were emulsified with equal amounts of distilled water by (1) homogenizing at low speed (Ultra-turrax T8, IKA Labortechnik) for 15 s and (2) sonicating for an additional 15 s at one third of the maximum sonication energy level (Sonifier 450, Branson Sonic Power). Emulsions were stored under nitrogen at 4°C for daily use.

Rotifer culture and enrichment. Rotifers were reared in a continuous high-density rotifer culture system (150 l) and maintained on *Nannochloropsis* Premium 3600 (Suantika et al. 2000) obtained from Aquatic Ecosystems, Inc). At 08:00 h and 16:00 h daily, rotifers were harvested and placed in smaller enrichment vessels in order to produce 2 batches of enriched rotifers for daily larval fish feedings. Rotifer enrichments were carried out at a density of 500 000 ind. l⁻¹ under gentle aeration with an air stone placed in the bottom of the conical 50 l vessel. Rotifer batches were enriched for 8 or 16 h (08:00 to 16:00 or 16:00 to 08:00 h) each 24 h period by adding 0.1 g of oil per liter of rotifer culture at the beginning of each enrichment (Dhert et al. 2001, Copeman et al. 2002). Emulsion oils were blended for ~30 s in 2 l of distilled water and added to enrichment vessels. Enriched rotifers were sampled from each ves-

sel in triplicate for lipid analysis 2 times each for the 8 and 16 h experiment.

Larviculture. Fish larvae for feeding experiments were reared in the laboratory from eggs collected from spawning adults. In April 2008, 2 female and 3 male Pacific cod were caught by commercial jigging gear from spawning grounds in Chiniak Bay, Kodiak Island, Alaska. The gametes were mixed and placed into 4 l incubation trays at 4°C. At 24 h post-fertilization, fertilized eggs were shipped in insulated containers filled with 4°C chilled seawater to Alaska Fisheries Science Center (AFSC) laboratory facilities in Newport, Oregon. Eggs were transferred to 4 l plastic flow-through trays and incubated at 4°C until hatching, as described by Laurel et al. (2008). Hatching occurred 19 to 22 d post-fertilization. Hatched larvae were transferred into 100 l cylindrical upwelling tanks, which were stocked with larvae from multiple hatching trays in order to avoid differential effects due to egg-incubation environment. To each of the 4 dietary treatments, 3 larval tanks were assigned for a total of 12 larval first-feeding culture tanks.

The feeding experiment was carried out in 12 conical upwelling 100 l fiberglass tanks with dark green interiors. Larvae were randomly sorted into 3 replicate tanks assigned to each of 4 dietary treatments. Tanks were kept at 12:12 h photoperiod to approximate day length conditions experienced by cod larvae in the Gulf of Alaska in March-April. Overhead fluorescent bulbs provided a photon level of 6.7 $\mu\text{mol m}^{-2} \text{s}^{-1}$ at the

Table 1. Ingredients of the 4 different oil emulsions, and lipid class composition of the 4 rotifer diets enriched for 8 or 16 h using these oil emulsions (mean \pm SEM, n = 4). In addition, all emulsions contained 5 % lecithin, 1 % vitamin E, 1 % ascorbic acid and 1 % Tween-80 (w/oil weight). Enriched rotifer diets contained <2.5 % hydrocarbons, ethyl & methyl esters, ethyl & methyl ketones, alcohols, and diacylglycerols (not specified in the table)

	Diet 1	Diet 2	Diet 3	Diet 4
Ingredients of oil emulsions (g)				
DHAsco-S	0	3.5	9.5	21.8
Cod liver oil	0	37.7	25.5	25.5
Olive oil	45	3.8	10	10
Water	50	50	50	50
Total ω -3	0	8.7	8.7	8.7
Lipids of enriched rotifers				
Total lipids ($\mu\text{g mg}^{-1}$)	132.8 \pm 42.8	133.7 \pm 65.1	80.1 \pm 38.9	107.2 \pm 7.3
Lipid classes (% total lipid)				
Steryl/wax esters	2.6 \pm 0.2	1.8 \pm 0.4	1.6 \pm 1.0	2.2 \pm 0.6
Triacylglycerols	35.0 \pm 4.3	44.2 \pm 3.5	35.4 \pm 5.8	38.0 \pm 3.8
Free fatty acids	7.7 \pm 1.1	6.0 \pm 1.5	6.7 \pm 1.2	7.4 \pm 0.9
Alcohols	2.7 \pm 1.3	5.6 \pm 0.5	6.7 \pm 2.5	3.7 \pm 2.7
Sterols	3.7 \pm 0.5	3.3 \pm 0.5	8.8 \pm 4.9	6.2 \pm 1.0
Acetone mobile polar lipids	6.1 \pm 0.7	8.7 \pm 1.3	16.2 \pm 6.5	11.0 \pm 2.1
Phospholipids	38.5 \pm 6.0	27.7 \pm 0.8	22.0 \pm 7.5	29.4 \pm 2.6

water surface. Water was supplied at a rate of 250 ml min⁻¹ through central-bottom intake to minimize disturbance to the larvae. Gentle aeration by an air-stone provided additional circulation in the tanks. From Day 2 until the end of the experiment, differentially enriched rotifers were added to tanks twice per day at a density of 4000 ind. l⁻¹ as considered optimal, saturating food conditions for cod larvae (Brown et al. 2003). Tanks were 'greened' by adding *Nannochloropsis* Premium 3600 (Aquatic Ecosystems, Inc) twice daily at a density of 1.06 \times 10⁹ cells l⁻¹. Greening larval tanks with microalgae provides larviculture benefits that are not fully understood, but are related to beneficial effects on foraging activity and increased nutritional condition in larvae through direct ingestion of microalgae (van der Meeren et al. 2007).

Size-at-age and survival. Fish larvae were sacrificed from experimental tanks at Weeks 1, 2, 3 & 4 for morpho-

metric measurements. From each of the 3 replicate tanks per treatment, 10 larvae tank⁻¹ wk⁻¹ were taken, i.e. 30 larvae treatment⁻¹, for measuring standard length (length in mm from the tip of the snout to the end of the notochord, SL), and body depth (width in mm of the larvae just posterior to the anus not including the fin fold, BD), using an image analysis system connected to a dissecting microscope. For dry weight (DW) determination, the 10 larvae were rinsed collectively in 3% ammonium formate solution to rid excess salt and inorganic material, placed on 1.5 cm² pre-weighed aluminum foils and dried in an oven at 68°C for 48 h. DW of pooled larvae was determined with a microbalance (Sartorius R16OP) to the nearest µg resulting in 1 measure tank⁻¹ and 3 measures treatment⁻¹ wk⁻¹. Average individual DW was calculated by subtracting the foil weight and dividing by the number of individuals on the foil. Foils were then stored in a desiccator and reweighed within 1 h. Survival was determined at the end of the study by counting all remaining larvae left in experimental tanks.

Lipid analysis. Total lipids and lipid classes were measured in both rotifers and larval fish to determine differences in dietary quality and their effects on larval condition. The major lipid classes in fish are triacylglycerols (TAG), sterols (ST) and phospholipids (PL). TAG is generally considered as the major storage lipid class in larval fish while PL and ST are important components of cellular membranes. However, recent studies have shown that PL is also important as an energy source in eggs and larval fish as well as in low-lipid juveniles (Evans et al. 1998, Copeman et al. 2008, Laurel et al. 2010). Relative improvements in larval condition in other species, such as herring or Atlantic cod have been attributed to elevated total lipid, TAG per DW and TAG:ST ratios (Fraser 1989, Lochman et al. 1995).

Lipid samples of larvae were collected at the beginning of the experiment (time zero) and at the end of Week 2 and 4. Not enough larvae survived Diet 1 to sample at Week 4 so only larvae from the 3 high PUFA diets were sampled for lipids at this end point. Fifty larvae per tank, and 3 samples per diet, were collected at Weeks 2 & 4. Individuals were pooled in order to obtain sufficient material for lipid class and FA analysis. Lipids were extracted in chloroform:methanol according to Parrish (1987) using a modified Folch procedure (Folch et al. 1957). Lipid classes were determined by thin layer chromatography with flame ionisation detection (TLC-FID) using a MARK V Iatroscan (Iatron Laboratories) as described by Parrish (1987). Extracts were spotted on silica gel coated Chromarods and lipid classes were separated in a 3 stage system. The separation was developed in (1) hexane:diethyl ether:formic acid (98.95:1.00:0.05) for 20 min, (2), hexane:diethyl ether:formic acid (79:20:1) for 40 min,

(3), 100% acetone (15 min) followed by 10 min chloroform:methanol:water (5:4:1). After each separation, the rods were scanned and the 3 chromatograms were combined using T-data scan software (RSS, Bemis, Tennessee, USA). The signal detected (mV) was quantified using lipid standards (Sigma, St. Louis, Missouri, USA). Lipid classes were expressed both in relative (mg g⁻¹ wet weight) and absolute amounts (µg ind.⁻¹).

Total lipid was analysed for FA composition. Fatty acid methyl esters (FAME) were prepared by transesterification with 14% BF₃ in methanol at 85°C for 90 min (Morrison & Smith 1964, Budge 1999). The FAMEs were analyzed on a HP 6890 GC FID equipped with a 7683 autosampler and a ZB wax GC column (30 cm long, internal diameter 0.25 µm; Phenomenex, USA). The column temperature began at 65°C for 0.5 min, ramped to 195°C at a rate of 40°C min⁻¹, held for 15 min, then ramped to a final temperature of 220°C at a rate of 2°C min⁻¹. This final temperature was held for 3.25 min. The carrier gas was hydrogen at a flow rate of 2 ml min⁻¹. The injector temperature started at 150°C and ramped to a final temperature of 250°C at a rate of 200°C min⁻¹. The detector temperature stayed constant at 260°C. Peaks were identified using retention times from standards (37 component FAME, BAME (bacterial fatty acid methyl ester), PUFA 1, PUFA 3; Supelco). Chromatograms were integrated using the HP ChemStation Chromatography Software (Version B00.00).

Data analysis. Differences in size-at-age and lipid profiles were performed on tank means. Differences between treatments in size-at-age measurements were analyzed using a repeated measures ANOVA with the explanatory variables being 'dietary treatment', week ('wk'), and the interaction between 'diet' and 'wk', (Statistix 7, Hicks 1982). There was a significant interaction between the effect of 'wk' and 'diet' on larval size-at-age and therefore, we examined weekly effects of diet on morphometrics and lipid composition using one-way ANOVAs with Tukey's multiple comparison tests. Data were examined for normality, homogeneity and independence to satisfy the assumption of the ANOVA. Significance for all tests was set at $\alpha = 0.05$. FA percentage data were arcsine-square root transformed to meet the assumptions of the model. Differences in the lipid classes and FAs of enriched rotifers were compared across diets using a one-way ANOVA with Tukey's multiple comparison tests.

Principal component analysis (PCA) was used to simplify multivariate FA and lipid class data by transforming correlated variables into a set of uncorrelated principal components (Minitab, version 15; Meglen 1992). This technique was employed using 9 highly discriminatory lipid variables from first-feeding Pacific cod larvae, and larvae analyzed at Weeks 2 & 4 from all

4 diets. The first 2 principal components (PC1, PC2) accounted for 78% of the variance among samples, which allowed a display of the major trends within the data set without significant loss of the total original variation. PCA lipid loading coefficients are defined as the correlation coefficients between the original lipid variables and the PCA axis. PCA scores are defined as the position of the original variables along the new PCA axis (Meglen 1992). Lipid variables were chosen based on biological significance and the degree of variance explained by a given lipid class or FA. A correlation matrix was used and coefficients and scores were grouped by cluster analysis using single linkage.

RESULTS

Enriched rotifers

Following lipid enrichment there was no significant difference in the total lipid or the proportion of different lipid classes among the 4 batches of rotifers ($F_{3,12} = 1.35$, $p > 0.5$, Table 1). On average, DW of all groups comprised 11.3% lipid, 38% TAG, and 29% PL.

Feeding different emulsions to rotifers resulted in 4 diets for cod larvae with significantly different FA profiles (Table 2, $p < 0.05$). Diets 2, 3 & 4 (high PUFA) did not vary significantly in their levels of total saturated fatty acids ($\Sigma\text{SFA} = 24\%$), total monounsaturated fatty acids ($\Sigma\text{MUFA} = 33\%$), and total polyunsaturated fatty acids ($\Sigma\text{PUFA} = 42\%$). In contrast, control Diet 1 had significantly lower levels of ΣSFA (22%), higher ΣMUFA (51%), and lower ΣPUFA (27%) than the other 3 diets. Elevated levels of 18:1 ω -9 in Diet 1 reflected the utilized olive oil while varying levels of DHA, EPA, AA, and ω -6 DPA were found in the other diets. Diet 2 had a significantly lower DHA:EPA ratio (0.8:1) than Diet 4 (2:1), while Diet 3 was intermediate (1:1). Levels of total ω -3 differed significantly, ranging from 9% in Diet 1 to 27% in Diet 2, while total ω -6 PUFA ranged from 19% in Diet 4 to 14% in Diet 2 (Table 2).

Growth and survival

There was a significant effect of sampling week on the 3 size-at-age parameters (SL, BD & DW; $F_{3,23}$,

Table 2. Fatty acid composition (% of total FA) of the 4 different rotifer diets enriched for 8 or 16 h using 4 different oil emulsions (mean \pm SEM, $n = 4$). ^{a,b,c}; different letters represent a significant difference among groups; $p < 0.05$, $F_{3,12}$, 1-way ANOVA with Tukey's multiple comparison test

Fatty acid (FA)	Diet 1	Diet 2	Diet 3	Diet 4
14:0	1.1 \pm 0.1 ^a	3.1 \pm 0.2 ^b	2.8 \pm 0.2 ^{bc}	2.3 \pm 0.2 ^c
16:0	16.7 \pm 0.1 ^a	16.4 \pm 0.4 ^a	16.9 \pm 0.4 ^{ab}	19.0 \pm 0.9 ^b
18:0	3.0 \pm 0.2	3.0 \pm 0.2	2.8 \pm 0.2	2.8 \pm 0.2
ΣSFA ¹	21.7 \pm 0.4 ^a	23.5 \pm 0.4 ^{ab}	23.7 \pm 0.4 ^{ab}	25.4 \pm 1.2 ^b
16:1 ω -7	4.4 \pm 0.5 ^a	6.4 \pm 0.6 ^b	5.8 \pm 0.5 ^{ab}	4.0 \pm 0.3 ^a
18:1 ω -9	41.2 \pm 2.2 ^a	18.0 \pm 0.5 ^b	18.9 \pm 0.3 ^b	23.1 \pm 0.9 ^b
18:1 ω -7	2.5 \pm 0.1 ^a	2.8 \pm 0.1 ^a	2.5 \pm 0.1 ^a	2.0 \pm 0.1 ^b
20:1 ω -9	0.9 \pm 0.1 ^a	2.0 \pm 0.2 ^b	1.8 \pm 0.2 ^{ab}	0.6 \pm 0.2 ^a
ΣMUFA ²	51.2 \pm 1.6 ^a	33.0 \pm 0.5 ^b	33.0 \pm 0.5 ^b	33.0 \pm 0.4 ^b
18:2 ω -6	15.0 \pm 0.7 ^a	10.2 \pm 0.8 ^b	10.4 \pm 0.7 ^b	10.9 \pm 0.7 ^b
18:3 ω -3	0.7 \pm 0.1 ^a	1.4 \pm 0.2 ^b	1.1 \pm 0.2 ^{ab}	0.6 \pm 0.1 ^a
20:4 ω -6 (AA)	0.9 \pm 0.3 ^a	1.6 \pm 0.0 ^{ab}	1.8 \pm 0.1 ^b	2.1 \pm 0.2 ^b
20:5 ω -3 (EPA)	5.2 \pm 0.8 ^a	11.1 \pm 0.6 ^b	9.3 \pm 0.7 ^b	6.0 \pm 0.5 ^a
22:5 ω -6 (ω 6DPA)	<0.01 ^a	1.1 \pm 0.2 ^{ab}	2.4 \pm 0.3 ^b	4.7 \pm 0.7 ^c
22:5 ω -3	2.5 \pm 0.2 ^a	3.8 \pm 0.1 ^b	3.6 \pm 0.1 ^{abc}	3.0 \pm 0.2 ^{ac}
22:6 ω -3 DHA	0.2 \pm 0.1 ^a	8.3 \pm 0.7 ^b	9.8 \pm 0.5 ^{bc}	11.4 \pm 0.8 ^c
ΣPUFA ³	26.6 \pm 1.7 ^a	42.6 \pm 0.9 ^b	42.7 \pm 0.9 ^b	41.2 \pm 1.4 ^b
$\Sigma\omega$ -3	9.1 \pm 1.1 ^a	26.8 \pm 0.5 ^b	25.6 \pm 0.5 ^b	21.7 \pm 0.7 ^c
$\Sigma\omega$ -6	16.9 \pm 0.9 ^a	14.0 \pm 0.6 ^b	15.9 \pm 0.3 ^{ab}	18.8 \pm 0.6 ^a
DHA:EPA	<0.1 ^a	0.8 \pm 0.1 ^b	1.1 \pm 0.1 ^b	2.0 \pm 0.3 ^c
EPA:AA	4.7 \pm 0.4 ^a	6.9 \pm 0.2 ^b	5.1 \pm 0.5 ^a	3.0 \pm 0.4 ^c

¹Includes minor amounts of *i*-15:0, *ai*-15:0, 15:0, *i*-16:0, *ai*-16:0, *i*-17:0, *ai*-17:0, 17:0, 20:0, 22:0, 24:0; ²Includes minor amounts of 14:1, 15:1, 16:1 ω -11, 16:1 ω -9, 16:1 ω -5, 17:1, 18:1 ω -11, 18:1 ω -5, 20:1 ω -11, 20:1 ω -7, 22:1 ω -11(13), 22:1 ω -9, 22:1 ω -7, 24:1; ³Includes minor amounts of 16:2 ω -4, 16:3 ω -4, 16:4 ω -3, 16:4 ω -1, 18:2 ω -4, 18:3 ω -6, 18:3 ω -4, 18:4 ω -3, 18:4 ω -1, 20:2 ω -6, 20:3 ω -6, 20:3 ω -3, 20:4 ω -3, 22:4 ω -6, 22:4 ω -3

$F > 75$, $p < 0.001$). Repeated measures also indicated a significant interaction between dietary treatment and the sampling week on all 3 morphometric parameters ($F_{9,23} > 4.11$, $p < 0.003$, Fig. 1a). Weekly differences in size-at-age were examined using 1-way ANOVA with Tukey's pairwise comparisons. At Week 4, larvae from Diet 1 were significantly shorter than larvae from the other treatments.

Further, BD of larvae in Diet 1 were significantly different from all other treatments at the end of Week 4 ($F_{3,7} = 8.3$, $p < 0.011$, Fig. 1b).

DW of larvae was also significantly affected by diet as larvae in Diet 1 weighed significantly less than all of the PUFA treatments. Further, larvae in Diets 2 & 3 weighed more than larvae from Diet 4 ($F_{3,10} = 42.3$, $p < 0.001$, Fig. 1c).

Larvae which were removed for lipid analysis and morphometric measurements were not considered in the calculation of survival. At the end of Week 4, the lowest survival (average \pm SD = 0.2 \pm 0.2%) was found for Diet 1, and not enough larvae were left to collect lipid data. Survival in the high PUFA Diets 2, 3 & 4 was similar (6 \pm 3%, 6 \pm 3%, and 7 \pm 3%, respectively).

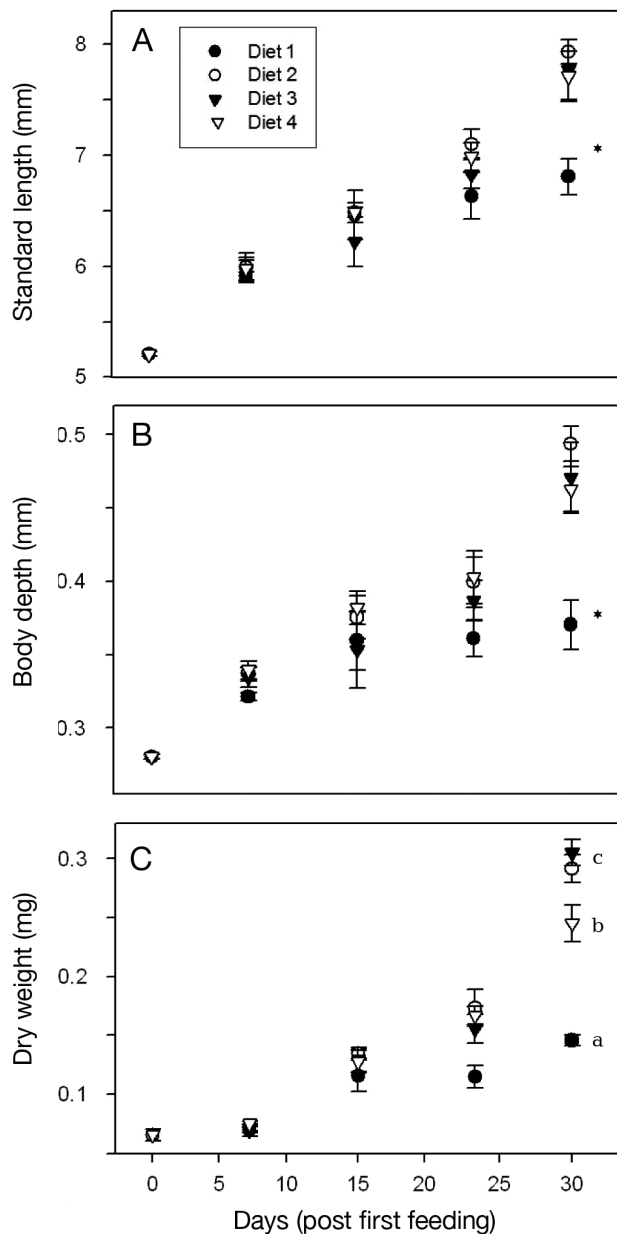


Fig. 1. *Gadus macrocephalus*. (A) Standard length, (B) body depth and (C) dry weight of Pacific cod larvae reared on 4 differently enriched rotifer diets for the first 4 wk post-hatch. Data are mean \pm SEM. Each symbol represents 30 individuals: ^{a,b,c}: different letters represent significant differences among dietary groups (ANOVA, Tukey's multiple comparison). *: Diet 1 was significantly different from all other treatments

Lipid class composition of larvae

On a DW basis there was no significant increase in total lipid over the course of the entire feeding trial ($F_{2,19} = 0.07$, $p = 0.933$, Fig. 2). Larvae had an average of $110 \mu\text{g mg}^{-1}$ of lipid per DW per individual throughout all treatments during the 3 sampling periods. There

was a trend towards higher levels of lipid per DW in Diet 2 compared to the other diets, but this was not significant (Week 2, $F_{2,5} = 2.4$, $p = 0.08$). Absolute amount of lipid per larva (average \pm SD) did increase from $15.6 \pm 5.1 \mu\text{g}$ in all dietary treatments at Week 2 to $31.9 \pm 10.0 \mu\text{g}$ at the end of the experiment (data not shown).

The effect of dietary treatment on the proportion of TAG present in the larvae was not significant at Week 2 or 4. However, by pooling all dietary treatments there was a significant effect of sampling week on the proportion of TAG in the larvae ($F_{2,19} = 22.9$, $p < 0.001$). Larvae at the beginning of the experiment had significantly higher levels of TAG than larvae at Week 2, while larvae at Week 4 had the highest levels of TAG (Fig. 2b).

The TAG:ST ratio followed the same trend as TAG, with an initial decrease to Week 2 and an increase to Week 4. However, at Week 4 this ratio showed a significant difference among dietary treatments ($F_{2,5} = 6.5$, $p = 0.04$). At Week 4, larvae in Diet 2 had a significantly higher TAG:ST ratio than larvae in Diet 4 (Fig. 2c). There was no significant effect of diet on the proportion of PL in the larvae at either Week 2 or 4 (Fig. 2d).

Fatty acid composition of larvae

Total FA per DW varied from $59 \mu\text{g mg}^{-1}$ in Diet 1 at Week 2 to $87 \mu\text{g mg}^{-1}$ in Diet 2 at the end of the experiment. When both weeks were pooled, larvae from Diet 2 had significantly more FA per DW than those from Diet 4 ($F_{3,15} = 5.23$, $p = 0.01$).

After just 2 wk of feeding, larvae showed significant differences in the levels of individual FAs (Table 3). For Diet 1, larvae had higher levels of 18:1 ω -9 (~17%) and lower levels of many of the longer chain PUFAs than for the other diets. Survival was not assessed at Week 2, however, larvae from Diet 1 showed higher mortality at Week 2 than those in the other PUFA diets. At Week 2, the larval DHA:EPA ratio varied from 1.4:1 in Diet 1 to 4.3:1 in Diet 4.

After 4 wk of feeding, larvae in the 3 high PUFA diets showed variable levels of individual PUFA but no significant differences ($p > 0.05$) in the ΣSFA , ΣMUFA or ΣPUFA . DHA levels reached 24% in Diet 4, while EPA was highest in Diet 2 (12%). The larval DHA:EPA ratio was significantly higher for Diet 4 (4.2:1) than for either Diet 2 or 3 (~2:1). Among larvae groups, 22:5 ω -6 varied significantly with the highest levels found in Diet 4 (6.5%) and significantly lower levels in Diets 1 & 2 (1.7% & 3.6%, respectively).

Dietary FA levels affected PUFA retention in larval tissue after only 2 wk of feeding on enriched rotifers (Fig. 3a). Larvae from the control had much higher lev-

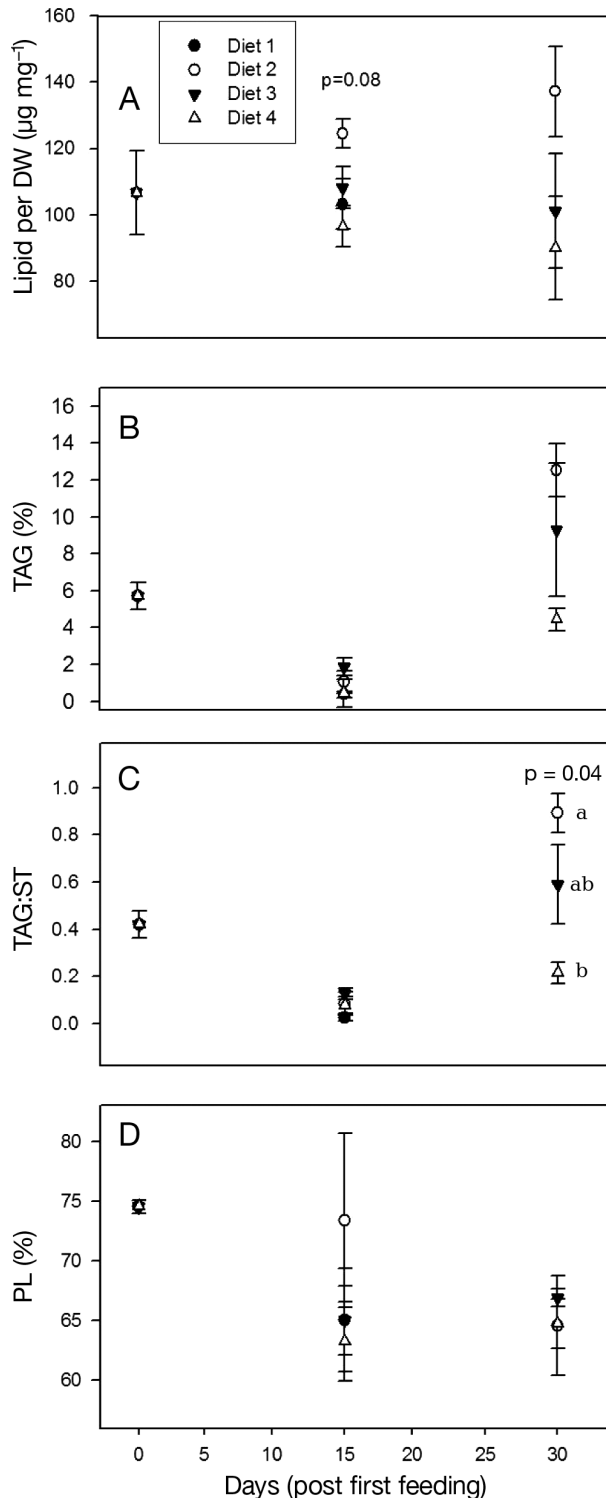


Fig. 2. *Gadus macrocephalus*. Lipids in Pacific cod reared on 4 differently enriched rotifer diets for the first 4 wk post-hatch. (A) Lipid per dry weight (DW), (B) triacylglycerols (TAG) in %, (C) ratio triacylglycerols:sterols (TAG:ST), (D) phospholipids (PL) in %. Data are means \pm SEM. ^{a,b,c}: different letters represent significant differences among dietary groups (ANOVA, Tukey's multiple comparison)

els of all PUFA in their tissues compared to dietary levels ($1.7 \times$ more Σ PUFA, $60 \times$ more DHA, Fig. 3a) and higher levels of 18:0, while levels Σ MUFA and 18:1 ω -9 were lower in all larvae than in their diets.

At Week 4, enough larvae to perform lipid analysis remained only in the PUFA enriched Diets 2, 3 & 4. Larvae from these diets showed conservation of DHA, 22:5 ω -3, 22:5 ω -6 (ω 6DPA), 20:4 ω -6 (AA), and 18:0 at a rate of 1.5 to $2 \times$ more than in the rotifers. EPA was present at levels approximately equal to that found in the rotifers while MUFA and the short chain PUFA 18:2 ω -6 were present at $\sim 50\%$ the proportions found in the diet (Fig. 3b).

PCA of 9 FA and lipid class variables simplified the lipid composition of larvae in terms of weekly and dietary effects (Fig. 4). Fig. 4a shows the first 2 principal components and separates larvae in terms of dietary differences. PC1 explained 44% of the variance, and shows a separation of the PUFA with high levels of DHA and AA on the positive side of the axis and higher levels of total lipids, and EPA on the negative side of the axis. Examination of the lipid loading coefficients for PC 2 (33%, Fig. 4a) indicated that this axis represented an unsaturation axis, with MUFA loaded on the positive side and high levels of PUFA and increased condition (TAG:ST) on the negative side of the axis.

Examination of sample scores showed that larvae in Diet 4 from Week 2 & 4 were associated with high levels of DHA, AA, and ω 6DPA. These larvae clustered together along the positive side of the axis. On the negative side of the axis, first-feeding larvae clustered together with the 4-week old larvae from Diet 2, indicating that these larvae had the most similar lipid composition, typified by higher levels of EPA relative to DHA. Larvae from Diet 1 had the lowest condition and low levels of PUFA, with the highest levels of MUFA. They clustered separately at the top of PC2 with an outlying larva from Diet 2. Larvae from Diet 3 showed an intermediate lipid composition.

Fig. 5 shows the relative proportions of EFAs, DHA, EPA and the ratio of DHA:EPA in differently enriched rotifers compared to that reported from wild copepods. We calculated the average and standard error of 6 yr of data on *Neocalanus plumchrus* from the Straits of Georgia (El-Sabaawi et al. 2009a). DHA, EPA and DHA:EPA ranged from 3.9 to 8.6%, 6.6 to 17%, and 0.3 to 1.1%, respectively (El-Sabaawi et al. 2009a). Proportions of DHA in the rotifers ranged from 0.2% in Diet 1 to 11.4% in Diet 4, while EPA was also lowest in Diet 1 (5.2%) and highest in Diet 2 (11.1%). DHA:EPA ratios ranged from $<0.1:1$ in Diet 1 to $2:1$ in Diet 4. A comparison of *N. plumchrus* with our rotifer diets indicated that Diet 2 was the most well-matched diet compared to wild copepods in terms of all 3 PUFA measures (Fig. 5).

Table 3. Fatty acid composition (% of total FA) of Pacific cod larvae fed differentially enriched rotifers for 4 wk post-hatch (mean ± SEM, n = 3). ^{a,b,c}: different letters represent a significant difference among groups; p < 0.05, $F_{3,7}$ at Week 2, $F_{2,5}$ at Week 4, 1-way ANOVA with Tukey's multiple comparison test

Fatty acids	First feeding	Week 2				Week 4		
		Diet 1	Diet 2	Diet 3	Diet 4	Diet 2	Diet 3	Diet 4
Total ($\mu\text{g mg}^{-1}$ DW)	86.7 ± 6.0	58.8 ± 4.9	75.5 ± 2.1	65.5 ± 6.4	54.4 ± 4.7	86.7 ± 9.0	61.5 ± 11.7	50.6 ± 10.3
16:0	20.9 ± 0.3	19.9 ± 3.0	16.9 ± 0.7	19.1 ± 1.8	16.6 ± 1.0	13.8 ± 0.5	14.7 ± 0.6	16.8 ± 1.2
18:0	4.8 ± 0.1	7.4 ± 0.7	6.6 ± 0.3	7.7 ± 1.0	6.4 ± 0.3	5.0 ± 0.1	5.3 ± 0.3	6.1 ± 0.3
ΣSFA^1	28.3 ± 0.3	30.9 ± 4.7	26.7 ± 1.0	29.9 ± 2.8	25.0 ± 1.5	21.5 ± 0.5	22.6 ± 1.0	24.9 ± 1.6
16:1 ω -7	2.7 ± 0.1	2.4 ± 0.1	4.0 ± 0.4	3.3 ± 0.3	2.4 ± 0.1	4.2 ± 0.1	3.7 ± 0.1	2.7 ± 0.2
18:1 ω -9	8.5 ± 0.2	16.6 ± 1.2	11.9 ± 1.0	12.3 ± 0.7	13.6 ± 0.2	11.7 ± 0.2	12.9 ± 0.1	14.6 ± 1.0
18:1 ω -7	5.0 ± 0.1	3.2 ± 0.1	3.9 ± 0.1	3.6 ± 0.3	3.1 ± 0.1	3.2 ± 0.0	3.1 ± 0.1	2.8 ± 0.2
ΣMUFA^2	20.2 ± 0.3	26.3 ± 1.5	23.8 ± 1.8	22.9 ± 1.0	22.2 ± 0.3	22.6 ± 0.3	22.7 ± 0.4	22.7 ± 1.42
18:2 ω -6	0.5 ± 0.0	7.4 ± 0.8	5.4 ± 0.6	4.9 ± 0.5	5.2 ± 0.3	6.5 ± 0.2	6.4 ± 0.2	5.8 ± 0.3
20:4 ω -6	2.9 ± 0.0	3.4 ± 0.3 ^a	3.1 ± 0.2 ^a	3.2 ± 0.1 ^a	4.3 ± 0.1 ^b	3.3 ± 0.0 ^a	3.8 ± 0.1 ^a	4.5 ± 0.2 ^b
20:5 ω -3	15.2 ± 0.0	9.1 ± 0.8 ^a	9.6 ± 0.3 ^a	7.8 ± 0.3 ^{ab}	5.9 ± 0.4 ^b	12.3 ± 0.2 ^a	9.9 ± 0.4 ^b	5.7 ± 0.1 ^c
22:5 ω -6	0.3 ± 0.0	0.3 ± 0.0 ^a	1.4 ± 0.1 ^b	2.8 ± 0.1 ^c	5.5 ± 0.2 ^d	1.7 ± 0.0 ^a	3.6 ± 0.1 ^b	6.5 ± 0.6 ^c
22:5 ω -3	1.6 ± 0.0	5.9 ± 0.6 ^a	4.9 ± 0.1 ^{ab}	3.7 ± 0.2 ^{bc}	3.1 ± 0.1 ^c	6.2 ± 0.1 ^a	4.9 ± 0.2 ^b	3.2 ± 0.1 ^c
22:6 ω -3	27.8 ± 0.1	12.5 ± 1.0 ^a	20.2 ± 2.6 ^{ab}	20.4 ± 1.6 ^{ab}	25.1 ± 0.7 ^b	20.4 ± 0.3	21.9 ± 0.5	24.1 ± 2.3
ΣPUFA^3	51.3 ± 0.1	42.8 ± 3.4	49.5 ± 2.2	47.2 ± 2.2	52.8 ± 1.8	55.7 ± 0.2	54.7 ± 0.7	52.4 ± 3.0
DHA:EPA	1.8 ± 0.0	1.4 ± 0.1 ^a	2.1 ± 0.3 ^{ab}	2.6 ± 0.1 ^b	4.3 ± 0.2 ^c	1.7 ± 0.1 ^a	2.2 ± 0.1 ^a	4.2 ± 0.3 ^b

¹Includes minor amounts of 14:0, *i*-15:0, *ai*-15:0, 15:0, *i*-16:0, *ai*-16:0, *i*-17:0, *ai*-17:0, 17:0, 20:0, 22:0, and 24:0; ²Includes minor amounts of 17:1, 18:1 ω -11, 18:1 ω -5, 18:1 ω -6, 20:1 ω -11, 20:1 ω -7, and 24:1; ³Includes minor amounts of 16:2 ω -4, 16:3 ω -4, 16:4 ω -3, 18:2 ω -4, 18:3 ω -6, 18:3 ω -4, 18:3 ω -3, 18:4 ω -3, 18:4 ω -1, 20:2 ω -6, 20:3 ω -6, 20:4 ω -3, and 22:4 ω -6

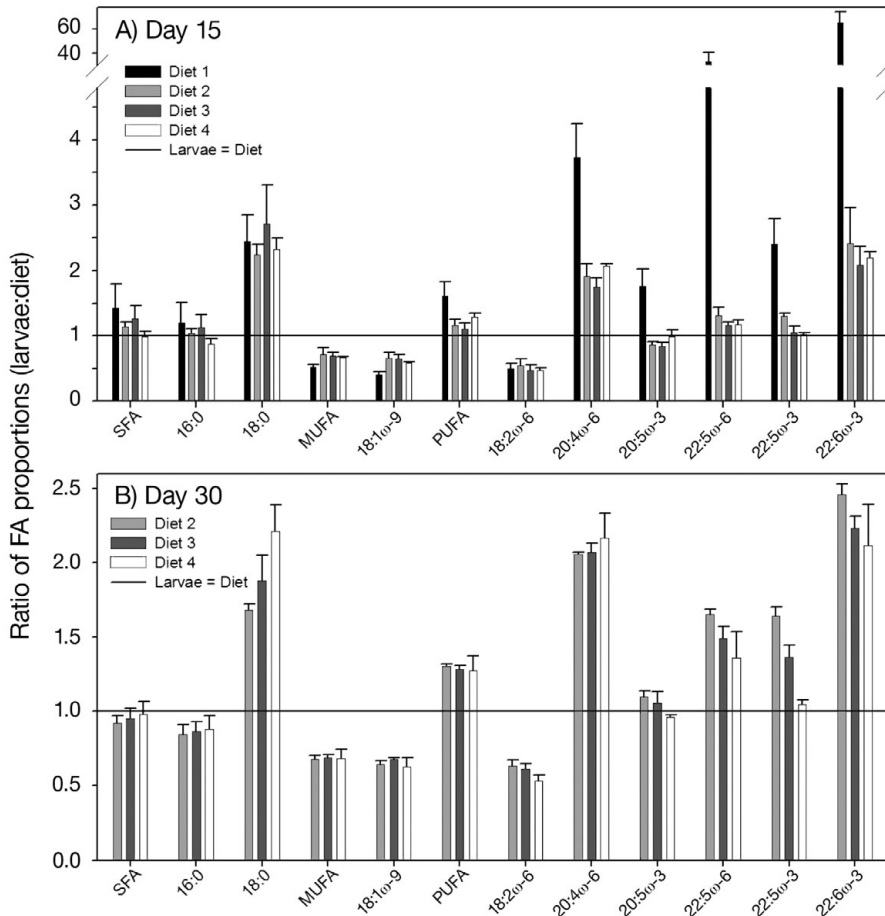


Fig. 3. *Gadus macrocephalus*. Ratio of specific fatty acids (FA) in larvae compared to dietary levels (larvae:diet) after (A) 2 wk and (B) 4 wk of feeding on 4 differentially enriched rotifer diets. Data are mean ± SEM, n = 3. Solid line: proportion of FA in the larvae equals that in the diet (1:1)

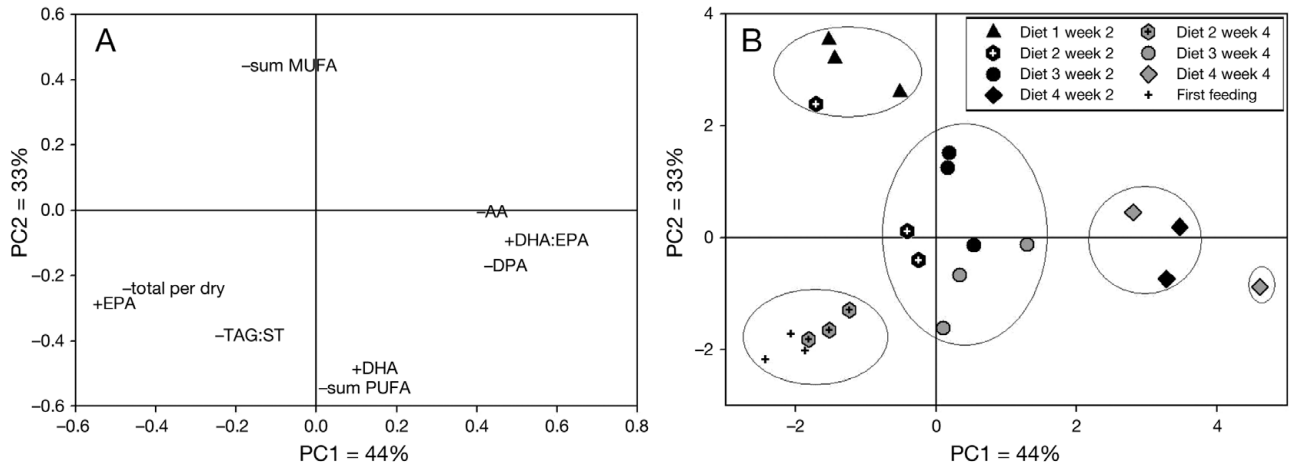
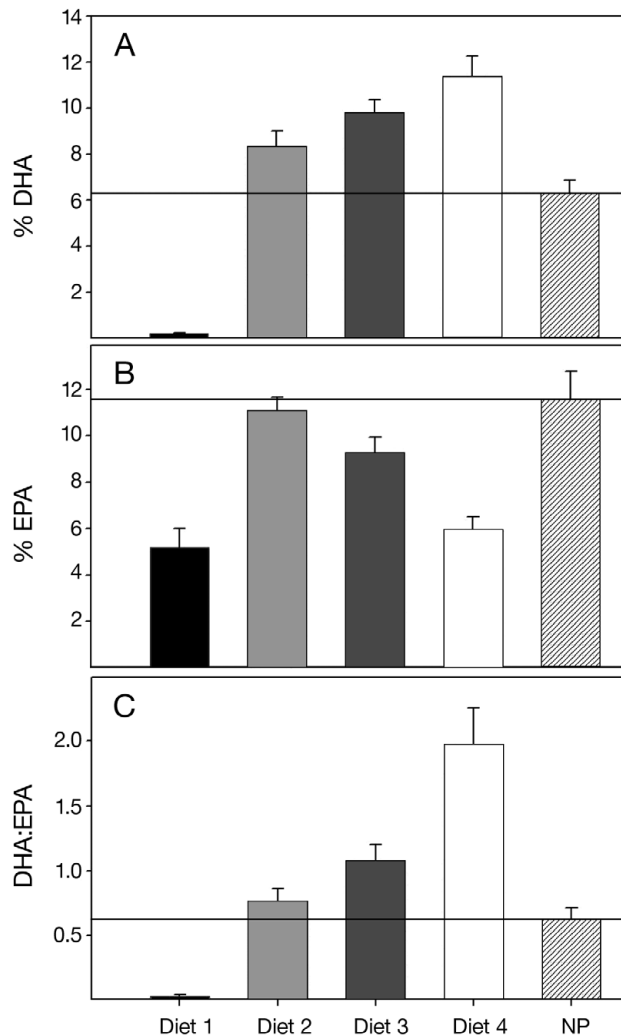


Fig. 4. *Gadus macrocephalus*. Analysis of the first 2 principal components of lipid data from larvae (first-feeding and feeding for 2 and 4 wk on 4 differentially enriched rotifer diets). The fatty acid and lipid class parameters used were: AA (20:4 ω -6), DHA (22:6 ω -3), ω 6DPA (22:5 ω -6), EPA (20:5 ω -3), DHA:EPA, Σ MUFA = monounsaturated fatty acids, total per dry = total fatty acids per dry weight $\mu\text{g mg}^{-1}$, TAG:ST = triacylglycerols:sterols, Σ PUFA = polyunsaturated fatty acids. Groups were determined by cluster analysis of (A) lipid parameter coefficients, symbols represent orientation along the third principal component axis and (B) larval fish scores for the first 2 principal components



DISCUSSION

The results of our experiment support our predictions that larval Pacific cod do require high levels of ω -3 PUFA for normal growth and development. Further, energetically similar diets with different DHA: EPA ratios did affect size-at-age and lipid composition. However, Pacific cod larvae did not show a high requirement for DHA relative to EPA but, rather, demonstrated highest growth at ratios ranging from 1.1:1 to 0.8:1. Diets 2 and 3 produced elevated growth and increased lipid condition indices (TAG:ST). After 4 wk of feeding, larvae from Diet 2 most closely resembled the levels of PUFA found in wild-spawned first-feeding Pacific cod larvae (Laurel et al. 2010). Further, the DHA:EPA ratio in Diet 2 rotifers (0.8:1) resembled that reported for 4 species of cold-water Pacific copepods (*Neocalanus plumchrus*, *Calanus marshallae*, *Euchaeta elongate* and *Eucalanus bungii*, El-Sabaawi et al. 2009b).

The rotifer *Brachionus plicatilis* is not a natural prey item for Pacific cod. Rotifers, however, are commonly used in both ecological and aquaculture studies on small marine fish larvae (Puvanendran & Brown 1998,

Fig. 5. *Neocalanus plumchrus* and rotifers. Relative proportions of (A) docosahexaenoic acid (DHA), (B) eicosapentanoic acid (EPA), and (C) the ratio DHA:EPA in 4 differently enriched rotifer diets and in the calanoid *Neocalanus plumchrus* (NP; averaged from zooplankton collected over 6 yr in the Strait of Georgia, British Columbia, Canada, see El-Sabaawi et al. 2009a)

Jordaan & Brown 2003, Imsland et al. 2006), because they are more easily mass-cultured than wild zooplankton and can be enriched with lipids or FAs to resemble specific prey types. Further, rotifers do not show significant retro-conversion of long chain PUFA into shorter chain PUFA like other live-cultured prey e.g. *Artemia* (Navarro et al. 1999). Still, fish larvae often grow and survive better on natural prey in the laboratory (e.g. Imsland et al. 2006), likely because lipid-enriched rotifers lack some of the amino acids, vitamins, minerals and digestibility characteristics of wild zooplankton (Sargent et al. 1999, Evjemo et al. 2003). Although Pacific cod larvae would be ideally cultured on marine copepods with variable ratios of EFAs, the techniques for controlled lipid enrichment of marine copepods have not been fully developed (Olivotto et al. 2008). For these reasons rotifers still represent the best vehicle to experimentally examine the effects of prey quality on fish larvae.

Larvae of Pacific and Atlantic cod feed opportunistically on protozoa, copepod nauplii or copepodites in the wild (van der Meeren & Naess 1993, Takatsu et al. 2002). Takatsu et al. (2002) showed that Pacific cod larvae off Japan consumed a variety of copepod nauplii and copepodites. Over a 3 year study, small larvae (3.6 to 7.0 mm TL) had many nauplii (41 to 83% of prey items) while large larvae (7.1 to 15.5 mm TL) had mostly copepodites (80.4 to 99.5%) in their guts. Here we examined just the effect of FA composition of 1 live-food on larval size-at-age and condition. Although Pacific cod larvae are capable of growing on rotifers up to 6 wk at similar temperatures (Laurel et al. in press), we cannot rule out the possibility that prey size constrained growth in our larvae during the last week of the experiment. However, this factor would likely have been more significant in larger larvae from Diet 2 & 3 than for smaller larvae in Diet 1. Therefore, any potential constraint by prey size in our experiment would have made our conclusions about dietary EFA on size-at-age somewhat conservative.

We intentionally formulated diets ranging in DHA:EPA ratios to represent the natural annual FA variation in plankton food webs based on diatoms and dinoflagellates (Budge & Parrish 1998, El-Sabaawi et al. 2009a). Previous studies on lipid nutrition in gadids have largely been based on commercial live-food enrichments, which often vary simultaneously in factors such as total lipids per dry wt, protein, lipid classes as well as multiple FAs (Park et al. 2006, Garcia et al. 2008a). Controlled studies using formulated experimental enrichment oils have been conducted to examine the importance of the essential FAs DHA (22:6 ω -3), EPA (20:5 ω -3) and AA (20:4 ω -6). However, most of these studies focus on commercially important Atlantic species (Copeman et al. 2002, Villalta et al. 2005a, Lund et al. 2007).

The functional significance of dietary DHA:EPA can be observed in terms of competitive interactions between FAs for incorporation into PLs; specifically, competition for the enzymes that esterify FAs onto the glycerophospho-based backbone (Sargent et al. 1999). The functional significance of the ratio of DHA:EPA has now been investigated in both primary consumers (Arendt et al. 2005, El-Sabaawi et al. 2009a) and fish larvae (Sargent et al. 1999, Izquierdo et al. 2000) as well as at the ecosystem level (Litzow et al. 2006). Specifically, this ratio has been well studied in relation to the dietary requirements of many marine fish that are candidates for aquaculture. Nutritional requirements for DHA and EPA have been found to be both species- and developmentally-specific (Copeman 2001, Villalta et al. 2005b).

From a population perspective, differences in reproduction and early life history suggest that Pacific cod larvae may be more susceptible to changes in prey quality in the field than Atlantic cod larvae. Although both species are highly fecund, and likely susceptible to high variation in survival during the first few weeks of life (May 1967, McCain 2003), Pacific cod eggs are semi-adhesive and are released in 1 batch during spring (Mecklenburg et al. 2002). This contrasts to Atlantic cod eggs which are pelagic and are released in batch spawning events across several months in spring and fall (Kjesbu 2006). Temporal (i.e. single batch-spawning) and spatial (i.e. reduced dispersal potential of eggs) spawning characteristics have been hypothesized to make Pacific cod more vulnerable to changes in their prey than Atlantic cod (Laurel et al. in press).

Pacific cod also differ from Atlantic cod in their ability to synthesize and convert lipids and FAs during egg development. Pacific cod eggs synthesize large amounts of lipid (presumably from protein) just prior to hatch, and demonstrate extreme conservation and possible synthesis of DHA as yolk-sac larvae (Laurel et al. 2010). This conservation of DHA by Pacific cod may reflect a unique ability to produce DHA from short chain precursors, however, the mechanism for this synthesis needs further investigation. DHA:EPA ratio in wild Atlantic cod eggs has been reported to be 2:1 (Finn et al. 1995) while levels in pre-feeding larvae have been found to be 1.8:1 (Garcia et al. 2008a) or 2.6:1 (Finn et al. 1995). For wild Pacific cod from 2006 and 2008, DHA:EPA ratios are slightly lower in both eggs and newly hatched larvae, 1.5:1 and 1.8:1 respectively (Laurel et al. 2010). Therefore, some of the differences in the natural history of these 2 cod species, coupled with lower levels of DHA:EPA in wild caught samples may indicate a lower dietary requirement for DHA and a higher for EPA in Pacific cod than in their Atlantic congener.

EPA is an important FA both for inclusion in cell membranes and for the production of biologically active compounds called eicosanoids or 'localized hormones'. AA (20:4 ω -6) is also used in the production of these localized hormones that include prostaglandins, thromboxanes, and leukotrienes. In fish, AA is the preferred substrate for the formation of eicosanoids and has been found to produce eicosanoids of higher biological activity than EPA (Bell et al. 1994). Flatfish fed high levels of AA relative to EPA have been found to develop high rates of malpigmentations (Estevez et al. 1999, Copeman et al. 2002). AA is always found in low levels in wild zooplankton and Pacific cod embryos (<2.5%; Budge & Parrish 1998, Laurel et al. 2010, El-Sabaawi pers. comm.). We offered a range of EPA:AA ratios in our diets with Diet 4 showing the lowest (3:1) and Diet 2 the highest levels (6.9:1). Van der Meeren et al. (2008) measured levels of EPA:AA in a number of copepod species and found them to be never less than 7.5:1 and as high as 49.5:1 due to very low levels of AA in the wild. The importance of this FA has led fish nutritionists to discuss optimum ratios in terms of a three-part index, DHA:EPA:AA, which has been hypothesized to be 10:1:1 (Park et al. 2006) or 11:1.5:1 (Garcia et al. 2008a) in Atlantic cod. Based on our higher weight and lipid condition indices for larvae in Diet 2, we suggest a preliminary dietary ratio of DHA:EPA:AA = 5:7:1 for first-feeding Pacific cod.

Rotifers enriched with all 4 experimental emulsions had the same total lipids per DW and the same proportions of different lipid classes. This ensured that the proximate lipid composition of our rotifers did not differ between dietary treatments. We enriched our rotifers once every 12 h. However, for maximal lipid retention and growth potential these experimental emulsions should ideally be added to rotifer cultures more repeatedly e.g. every 4 h. This is likely why the levels of total lipid per DW in our data were ~11% while Copeman et al. (2002) reported levels of ~16% using similar experimental emulsions. Enrichment every 12 h provided the variation in DHA:EPA ratios that we required to mimic natural variation reported in zooplankton of the North Pacific, despite possible reductions in growth and survival potential.

Temperature and food availability are often emphasized as the most limiting factors regulating the vital rates of fish larvae (Buckley et al. 2004). However, our experiment indicated that prey quality can explain similar variance in growth and survival. Our experiment was conducted at 8°C, yet Pacific cod larvae exposed to Diet 1 grew at rates similar to Pacific cod larvae reared at 3°C (2% d⁻¹; Laurel et al. in press). In Diets 2 & 3, the observed growth rates (i.e. ~5% d⁻¹) were on the lower range of those reported for Pacific cod larvae reared at similar temperatures (5 to 12%

d⁻¹), but this is likely attributable to using experimental enrichment emulsions as opposed to commercially formulated rotifer enrichments. While Diet 1 was an extreme and unlikely scenario for Pacific cod to face in the field, the growth variation among the 3 PUFA diets was measurable and would likely have significant survival consequences for Pacific cod larvae in the field when faced with size-dependent predation.

Survival at the end of the experiment was low in all treatments, with an average of 6% in the 3 PUFA diets and only ~1% in Diet 1. Mortality of marine fish larvae is extremely high in the field, and in laboratory experiments can be driven by numerous uncontrolled factors and tank effects. Interestingly, survival was much lower after Week 2 than after Week 1, most notably in Diet 1. Further, a reduction in the TAG:ST ratio and proportion of TAG was observed at Week 2. This was followed by a dramatic increase in condition and lipids at Week 4. Week 2 samples therefore likely contained larvae that had not successfully started feeding or were starving due to inadequate nutrition. Day 13 to 18 post-hatch at 8°C is the period at which 100% mortality occurs in non-feeding Pacific cod larvae (Laurel et al. 2008). Lipid class analysis showed that larvae fed low PUFA had a significantly lower TAG:ST ratio than larvae in all other treatments. Relative improvements in larval condition in other species, such as herring or Atlantic cod, have been attributed to elevated total lipid, TAG per DW, and TAG:ST ratios (Fraser 1989, Lochman et al. 1995).

Although levels of DHA, EPA, and AA have been well investigated for their effects on the early survival, growth, and development in fish and marine invertebrates (Sargent et al. 1999, Arts et al. 2001), more recently ω 6DPA (22:5 ω -6) has received attention as an EFA. Parrish et al. (2007) used stable isotope data and FA proportions to show that this FA was conserved at very high levels in larval tissue. Further isotopic evidence showed that these high levels were due to conservation of this long chain ω -6 PUFA rather than increases due to chain elongation of shorter chain precursors. Our results confirm that this FA was also conserved at high levels in Pacific cod larvae, at a rate of 1.5 \times what was found in the diet. This is similar to the level of conservation of ω 3DPA and lower than the levels of 2.5 \times seen for DHA in all larvae at Week 4. Despite the conservation of this FA in larval tissues, inclusion in the diet of Pacific cod did not result in increased growth or survival. This is contrary to reports for both larval Atlantic cod and larval scallops (*Argopecten irradians*), where ω 6DPA has been associated with increased growth (Milke et al. 2006, Garcia et al. 2008b). However, the addition of this FA to the diet of larval haddock did not increase growth or survival despite retention of this FA at high levels within larval tissues

(Garcia et al. 2008b). Future work is required to test the importance of this FA to the growth and survival of marine species without simultaneous variation in other highly essential FAs such as DHA and EPA. It has been suggested that this FA can be used as a C22 PUFA substitute in larval tissues when inadequate levels of DHA are present in the diet (Garcia et al. 2008a).

Conclusions

In the North Pacific, shifts between EPA-rich diatoms and DHA-rich dinoflagellates result in variable DHA:EPA ratios in zooplankton (El-Sabaawi et al. 2009a) but the effects on Pacific marine fish larvae remain poorly studied. We have shown that Pacific cod larvae are sensitive to changes in the ratios of essential PUFA in their diet and that optimum dietary levels are comparable to those seen (on average) in the wild. Therefore, given the sensitivity of marine fish larvae to the nutritional composition of zooplankton, further efforts should be made to determine not only the effect of changes in zooplankton abundance and species composition (Beaugrand et al. 2003, Batten & Welch 2004, Mackas et al. 2007) but also the effect of changes in essential parameters of zooplankton nutritional quality for fish larvae. Furthermore, it will be important to determine when such prey quality is most critical in developing fish larvae. Given the sensitivity of zooplankton to phytoplankton species composition and FA proportions (El-Sabaawi et al. 2009a) it is likely that food quality will help explain a portion of variability in year class strength observed in Pacific cod throughout the North Pacific.

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