

Succession and fate of the spring diatom bloom in Disko Bay, western Greenland

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ABSTRACT: Phytoplankton and copepod succession was investigated in Disko Bay, western Greenland from February to July 2008. The spring phytoplankton bloom developed immediately after the breakup of sea ice and reached a peak concentration of 24 mg chl *a* m⁻³ 2 wk later. The bloom was analyzed during 3 phases: the developing, the decaying, and the post-bloom phases. Grazing impact by the copepod community was assessed by 4 methods; gut fluorescence, *in situ* faecal pellet production, and egg and faecal pellet production from bottle incubations. *Calanus* spp. dominated the mesozooplankton community. They were present from the initiation of the bloom but only had a small grazing impact on the phytoplankton. Consequently, there was a close coupling between the spring phytoplankton bloom and sedimentation of particulate organic carbon (POC). Out of 1836 ± 180 mg C m⁻² d⁻¹ leaving the upper 50 m, 60 % was phytoplankton based carbon (PPC). The composition and quality of the sedimenting material changed throughout the bloom succession from PPC dominance in the initial phase with a POC/PON ratio close to 6.6 to a dominance of amorphous detritus with a higher POC/PON ratio (>10) in the post-bloom phase. The succession and fate of the phytoplankton spring bloom was controlled by nitrogen limitation and subsequent sedimentation, while grazing-mediated flux by the *Calanus*-dominated copepod community played a minor role in the termination of the spring bloom of Disko Bay.

KEY WORDS: Spring bloom · *Calanus* spp. · Grazing · Faecal pellets · Sedimentation · Arctic · Greenland

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INTRODUCTION

The spring phytoplankton bloom is a key event in high-latitude marine ecosystems, responsible for fueling high secondary production. It is triggered by the break-up of sea ice, combined with a warming and freshening of the surface layer stratifying the water column. The bloom is characterized by a short and pronounced growth peak of diatoms that depletes the surface layer of nitrate (Andersen 1981) and subsequently

sediments (Nielsen & Hansen 1995, Poulsen & Reuss 2002).

Sedimentation of phytoplankton from the euphotic zone is the principal mechanism in transport of organic material to benthic communities. Transport occurs directly as intact cells or indirectly as faecal pellets from pelagic grazers, aggregates, and amorphous detritus (Turner 2002, De La Rocha & Passow 2007). During its transit to the sea bed, the organic material is modified by the pelagic heterotrophs in a way that

influences its quantity and quality as a carbon source for the benthic community (Kjørboe et al. 1996, Kjørboe 1998, Wassmann 1998, Turner 2002, Riser et al. 2008).

Grazing is a key process that reduces the sedimentation of phytoplankton while simultaneously accelerating the sedimentation through production of fast-sinking faecal pellets (Juul-Pedersen et al. 2006). However, a large portion of the faecal pellets may be retained in the water column due to grazing, disintegration, and remineralisation (Noji et al. 1991, Riser et al. 2002, 2007, Turner 2002, Reigstad et al. 2008). Copepods often only graze a minor fraction of the spring phytoplankton bloom, because their longer generation time compared to that of phytoplankton limits them in numerically responding to the bloom (Kjørboe 1998). This results in a high sedimentation of ungrazed phytoplankton. In Arctic areas however, where overwintering stocks of copepods like *Calanus* exist, the copepod grazing impact on the bloom dynamics may be larger, depending on whether ascending *Calanus* match the spring phytoplankton bloom or not (Bathmann et al. 1990, Wassmann 1998, Wassmann et al. 2004, 2006).

The pelagic food web structure and production in Disko Bay, western Greenland, has been intensively investigated since the early 1990s (Nielsen & Hansen 1995). The original focus was mainly on baseline descriptions of composition and trophodynamics of the pelagic food webs (Levinsen et al. 2000a, Madsen et al. 2001, Madsen et al. 2008b). Few studies have considered the sedimentation of organic matter from the surface layer in this region (Juul-Pedersen et al. 2006, Sejr et al. 2007), and investigations resolving the contribution from the spring bloom are lacking. A match between copepod ascent to the surface and the spring phytoplankton bloom would lead to a large grazing impact on the phytoplankton biomass, while a mismatch would result in a larger contribution of ungrazed phytoplankton to the sedimentation.

Over the last decades, the physical forcing of the plankton succession in Disko Bay has changed. Recently an acceleration of the melt water input from the Jakobshavn Isbræ glacier to the bay has been documented (Holland et al. 2008), which, in combination with the general reduction of the sea ice cover (IPPC 2008), potentially impacts stratification, light conditions for the plankton, and consequently the timing and succession of the lower trophic levels of the food chain. Therefore baseline knowledge of

carbon flow during the short and pronounced spring diatom bloom needs to be acquired if the ecological impact of future environmental disturbances associated with climate change and increased human activities (e.g. oil exploration) is to be understood. The aim of the present study was to investigate the succession and fate of the spring diatom bloom in Disko Bay, with emphasis on evaluating the role of sedimentation versus grazing impact by the *Calanus* dominated zooplankton community.

MATERIALS AND METHODS

Study site. The study site was located in the Disko Bay off Qeqertarsuaq, western Greenland (Fig. 1). Due to land-connected sea ice coverage during winter, 2 sampling sites were combined. At the first site in winter (21 February to 23 March 2008), sampling was conducted through a hole in the ice at ca. 65 to 160 m depth approximately 0.5 nautical mile (n mile) south of Qeqertarsuaq (69° 14' N, 53° 29' W). In spring and summer (9 April to 18 July), sampling was done at a monitoring station 1 n mile south from Qeqertarsuaq (69° 14' N, 53° 23' W) (Fig. 1) at 300 m depth, onboard RV 'Porsild' (Arctic station, University of Copenhagen) and 'Maja S' (Finn Steffens, Qeqertarsuaq).

Sampling. Sampling was carried out between 10:00 and 17:00 h. Vertical profiles of water temperature and salinity were measured down to 150 m during winter and down to 250 m during spring and summer, using a

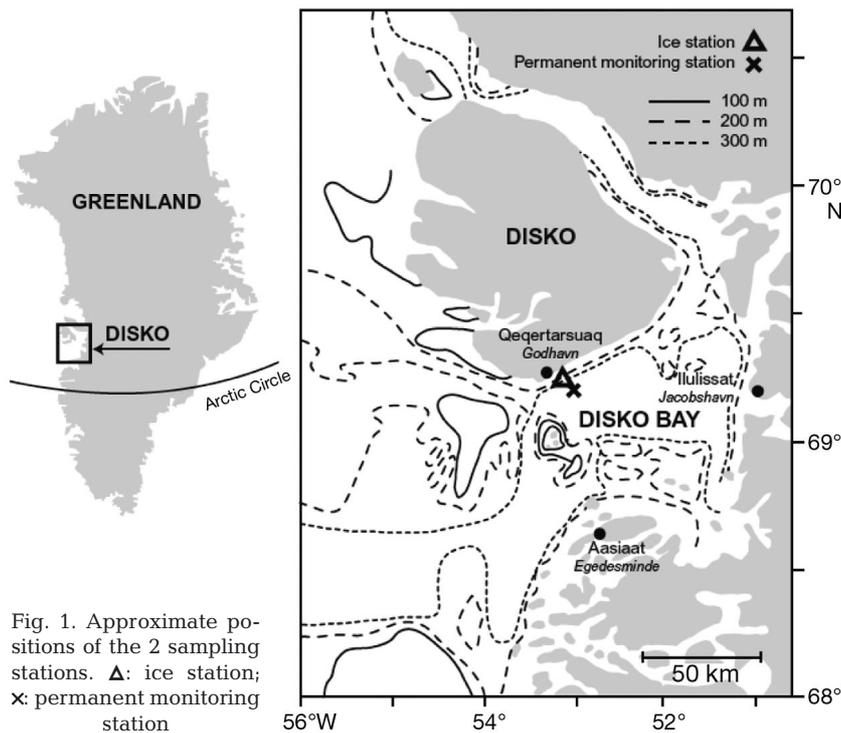


Fig. 1. Approximate positions of the 2 sampling stations. Δ : ice station; \times : permanent monitoring station

Seabird SBE25-01-CTD. Salinity measurements were calibrated against salinity samples taken approximately once a month ($n = 4$) throughout the study phase, and analyzed on an 8410-Portasal salinometer (Guildline).

Samplings for chemical and biological variables were taken at 6 depths (1, 20, 50, 75, 100, and 150 m) using a 10 l Niskin bottle. Two additional depths (200 and 250 m) were included after ship-based sampling was initiated. Chlorophyll *a* (chl *a*), particulate organic carbon (POC), and particulate organic nitrogen (PON) samples were siphoned directly to 10 l acid cleaned dark carboys and kept in the dark until filtering (max. 1 h after arrival to laboratory). Inorganic nutrient samples (phosphate, nitrate, ammonia, and silicate) were immediately frozen (-30°C) for later analysis on a Skalar autoanalyser (Breda), following the procedures of Grasshoff (1976). Nutrient sample precisions were 0.06, 0.1, 0.3, and 0.2 μM for phosphate, nitrate, ammonia, and silicate, respectively. For analysis of size-fractionated chl *a* and phaeopigments (phaeo), triplicate subsamples (200 to 500 ml) were filtered onto Whatman GF/F filters (0.7 μm) and mesh nets (10 μm and 50 μm). The filters were extracted in 5 ml 96% ethanol overnight (Jespersen & Christoffersen 1987), and pigments were measured on a TD-700 fluorometer (Turner) and calibrated against a chl *a* standard before and after acidification (Yentsch & Menzel 1963).

Samples of POC and PON were collected at 3 depths (1, 20, 50 m) from the ice station, with an additional fourth depth (100 m) from 30 April onwards. Samples (0.7 to 4.5 l) were filtered onto pre-combusted (400°C for 2 h) Whatman GF/F filters, which were stored frozen (-30°C) until analysis on a CHNS Automatic Elemental Analyzer (EA 1110, CE Instruments) after drying at 60°C for 24 h. POC was divided into 3 fractions: chl *a*-based POC (PPC), faecal pellet-based POC (FPC), and amorphous detritus-based POC (amorphous detritus). PPC was estimated from a regression line between chl *a* and water column POC (see Eq. 7 in 'Results'). The amorphous detritus fraction was estimated by subtracting PPC and FPC from POC.

Mesozooplankton. During sampling from the ice, mesozooplankton was collected using a modified WP-2 net (45 μm) equipped with a closing mechanism (Hydrobios). Samples were collected in 3 depth strata (0–50, 50–100, and 100–150 m). During ship-based sampling, mesozooplankton was collected with a multi-net (50 μm) equipped with a flow meter (Multinet, Hydrobios type midi), and 2 additional depth strata (150–200 and 200–250 m) were included. In addition to the seasonal study one diurnal investigation with sampling every 6 h was conducted from 29 April at 12:00 h to 30 April 30 at 12:00 h. Samples were immediately preserved in buffered formalin (5% final concentration) for later analyses. Biomass values of the different copepod

species were calculated based on measurements of prosome length, and length/weight relationships. Two regressions for *Calanus* spp. were established for biomass calculations: one applicable prior to and during the phytoplankton bloom until 4 May, and another from 9 May onwards (R. Swalethorp et al. unpubl.). Regressions for the remaining copepod species were taken from the literature (Satapoomin 1999, Thor et al. 2005, and references therein).

Gut fluorescence. Gut fluorescence were measured on females of *Calanus finmarchicus*, *C. glacialis*, and *C. hyperboreus* collected using a WP-2 net towed at 0.5 m s^{-1} from 100 m to the surface. A subsample of the cod-end was immediately concentrated on a piece of 100 μm plankton gauze and frozen to -40°C by freeze spraying (75 super, Kontakt Chemie) and then stored frozen (-30°C). Females of *C. hyperboreus* were gently collected from the rest of the cod-end and transferred individually into vials containing 5 ml 96% ethanol and kept dark for later analysis. The full operation took less than 5 min. In the laboratory, samples of *C. glacialis* and *C. finmarchicus* were gently thawed under dim light, diluted with 0.2 μm ice-cold filtered seawater, and triplicate pools of *C. finmarchicus* ($n = 5$) and *C. glacialis* ($n = 3$) were extracted in 5 ml 96% ethanol. All samples were analyzed for chl *a* and phaeo as described above. Background values of *C. finmarchicus* ($n = 45$), *C. glacialis* ($n = 25$), and *C. hyperboreus* ($n = 20$) were estimated after incubation for 24 h in 0.2 μm filtered seawater that was renewed after 12 h. Background values were subtracted from all the *Calanus* spp. gut fluorescence measurements.

Ingestion rate (I ; $\mu\text{g C individual}^{-1}\text{ d}^{-1}$) was calculated using the following equation (modified from Harris et al. 2000)

$$I = K \times G \times 60 \times 24 \quad (1)$$

where K is the instantaneous gut defecation rate constant (min^{-1}) and G is the gut pigment ($\mu\text{g chl } a$). The gut pigment was converted to carbon by application of a conversion factor of 29.9 measured during the present investigation. Instantaneous gut defecation values of 0.032 for *C. finmarchicus* (Maar et al. 2002), 0.017 for *C. glacialis*, and 0.015 for *C. hyperboreus* (Hansen et al. 1990) were used and corrected to *in situ* temperature by applying a Q_{10} value of 2.8 (Hansen et al. 1997). Specific ingestion rate ($\% \text{ d}^{-1}$) was calculated using *in situ* carbon contents of the 3 *Calanus* spp. (R. Swalethorp et al. unpubl.).

In situ faecal pellet production. Faecal pellet production of the copepod community was measured in short time incubations during ship-based sampling from 23 April to 9 June. Copepods were collected using a WP-2 net (200 μm) from 100 m to the surface. Subsamples of the cod-end were immediately distrib-

uted into 8 fecatrons made of clear PVC (polyvinyl chloride) tubes (inner diameter 8 cm, height 40 cm) filled with 45 μm filtered surface water. One additional subsample was immediately fixed in 2% (v/v) acidic Lugol's solution and served as a start sample. Four fecatrons were equipped with a 200 μm and 4 with a 400 μm false mesh bottom. The procedure from sampling of copepods to incubation start took less than 5 min. Concentrations in the fecatrons were between 20 and 385 copepods l^{-1} , and they were incubated for 1 to 2.5 h in a 100 l container cooled with surface water. Present copepod predators (primarily *Sagitta* spp. and *Pareuchaeta* sp.) were removed before the start of the experiment. At the end of the incubations the samples were preserved with 2% (v/v) acidic Lugol's solution. In the laboratory, faecal pellets and copepods were concentrated on a 20 μm sieve and counted and measured using a stereo light microscope (Olympus SZ40, 40 \times magnification for faecal pellets and 20 \times for copepods). The dimensions of maximum 30 faecal pellets and 60 copepods were measured and the rest were counted. Faecal pellet measurements and counting criteria were determined, so that the faecal pellets had to be a minimum of 3 \times longer than wide. Faecal pellet volume was calculated assuming cylinder shape. FPC was estimated applying a volume to carbon conversion factor of 0.043 $\text{pg C } \mu\text{m}^{-3}$ (R. Swalethorp et al. unpubl.). Copepod carbon content was calculated from a combined length/weight regression of *Calanus finmarchicus* and *C. glacialis* in the 2 time series as described above (R. Swalethorp et al. unpubl.).

Specific faecal pellet production rate (SPP, $\mu\text{g C}_{\text{pellet}} \mu\text{g C}_{\text{cop}}^{-1} \text{d}^{-1}$) was calculated using the following equation:

$$\text{SPP} = \text{FPC}_{\text{prod}} / (\text{C}_{\text{cop}} \times t) \quad (2)$$

where FPC_{prod} is the carbon content of the faecal pellets produced ($\mu\text{g C}_{\text{pellet}}$) during the experiment. C_{cop} is the biomass of the copepods ($\mu\text{g C}_{\text{cop}}$), and t is the duration of the experiment in days. Specific ingestion rate ($\% \text{d}^{-1}$) was calculated assuming an assimilation efficiency of 68.2% (Conover 1966).

Sedimentation. Sedimentation measurements were conducted from 23 April to 9 June using free-floating sediment traps fitted on a buoy (0.5 m^3 , 75 kg buoyancy) and moored with a weight (20 kg) to reduce water resistance and upward drift. The sediment traps (KC Denmark) consisted of 2 parallel acrylic cylinders mounted in a gimballed frame equipped with steering fins and pivot joints to ensure a vertical position perpendicular to the current. The traps had an internal diameter of 0.052 m with an aspect ratio (height:diameter) of 6.35. The sediment traps were deployed at 20, 50, and 100 m, with an average deployment time of 5.35 ± 0.1 h (10:00 to 16:00 h). During deployment, the sediment traps did not drift more than 1 n mile away from the sampling position,

except on one occasion where tracking of the traps was temporarily interrupted and deployment time was 24 h (23 April). Prior to deployment, the traps were filled with 0.2 μm filtered seawater from depths >100 m with added salt to increase the salinity by 5 psu (Knap et al. 1996). Hereby, advective diffusive exchange between the higher density, particle-free water and the ambient seawater was reduced in the traps during deployment. No preservative was added.

Immediately after recovery, the sedimentation traps were sealed with a clean lid and protected from direct light. In the laboratory, samples were visually inspected for swimmers, which were removed, and kept at 5°C in darkness until analysis (<2 h). The trap contents were carefully mixed before subsampling. To ensure minimal disturbance of the collected material, samples for microscopic examination were taken first. Subsamples for faecal pellets (600 to 900 ml) were preserved with 2% (v/v) acidic Lugol's solution. Analyses for total chl *a*, phaeo (100 ml in triplicates), POC and PON (500 ml), and for faecal pellets were performed as described above for the *in situ* faecal pellet production experiment.

Suspended faecal pellets. Parallel to the trap deployment, water samples were collected using a 30 l Niskin bottle at 3 trap deployment depths (20, 50, and 100 m) for quantification of suspended faecal pellets. The water samples were concentrated on a 20 μm sieve, fixed in 2% (v/v) acidic Lugol's solution, and analysed for faecal pellets and copepod abundance and biomass as described above.

Calculations. The sedimentation rate ($\text{mg m}^{-2} \text{d}^{-1}$) of the measured variables was calculated from the following equation (Knap et al. 1996):

$$\text{Sedimentation rate} = (\text{C}_{\text{trap}} \times V_{\text{trap}}) / (\text{A}_{\text{trap}} \times t_{\text{dep}}) \quad (3)$$

where C_{trap} (mg m^{-3}) is the concentration of the measured variable in the sediment trap, V_{trap} (m^3) is the volume of the sediment trap, A_{trap} (m^2) is the sediment trap surface area, and t_{dep} (d) is the deployment time. Sinking velocity (m d^{-1}) was estimated for different variables using the following equation (Kjørboe et al. 1994):

$$\text{Sinking velocity} = \text{Sedimentation rate} / \text{C}_{\text{in situ}} \quad (4)$$

where the sedimentation rate is from Eq. (3) and $\text{C}_{\text{in situ}}$ (mg m^{-3}) is the *in situ* concentration of the variables at 50 m depth.

The daily loss rate ($\% \text{d}^{-1}$) of chl *a* biomass in the 0–50 m depth stratum due to sinking export was estimated using the following equation (Olli et al. 2002):

$$\text{Daily loss rate} = \text{Sedimentation rate} \times 100 / \text{C}_{\text{int}} \quad (5)$$

where sedimentation rate is from Eq. (3) and C_{int} (mg m^{-2}) is the integrated biomass of chl *a*, estimated vertically from integration of the water column measurements at 0–50 m.

The daily loss ($\% \text{ d}^{-1}$) of phytoplankton due to copepod grazing was estimated using Eq. (5) by replacing the sedimentation rate with the copepod grazing calculated from faecal pellet production, i.e. the specific faecal pellet production multiplied by the integrated biomass of copepods (mg C m^{-2}) in the 0–50 m depth stratum and assuming an assimilation efficiency of 68.2% (Conover 1966). The present estimate assumes that the copepod prey is dominated by phytoplankton, which is most valid during the bloom. Outside the bloom, protozooplankton can contribute significantly to the diet of *Calanus* spp. (Levinson et al. 2000b), and the daily ratio will then be overestimated accordingly.

Statistical procedures. Data was tested for normal distribution using the Shapiro-Wilks test and, if necessary, log transformed. Effects of different mesh sizes in the fecatrons were tested with 1-way ANOVA (Statistica 2004, version 7.0; Statsoft). Linear regressions were determined using ANOVA tests (Sigmaplot 2004, version 10.0, Systat Software) to examine potential relationships between carbon and chl *a* of the suspended or settled material as well as the relationship

between food size and specific ingestion rate. Comparisons of regression lines were done with a 2-tailed *t*-test and accepted as significantly different if $p < 0.05$. The comparison of faecal pellet sizes and sedimentation fluxes could not be fitted to a normal distribution, and a non-parametric analysis followed by a Kruskal-Wallis test was used. Velocity difference of the organic materials was tested with 1-way ANOVA as were differences between the 4 methods for estimate specific ingestion rates. All data are presented as means \pm SE.

RESULTS

Hydrography and nutrient distribution

The ice coverage in Disko Bay was 60% until break-up, between 23 March and 9 April, after which it melted and advected out of the bay. During winter, when sampling took place from the ice station, the water column was stratified with colder (-1.7°C) surface water separated from a warmer (1.7°C) bottom

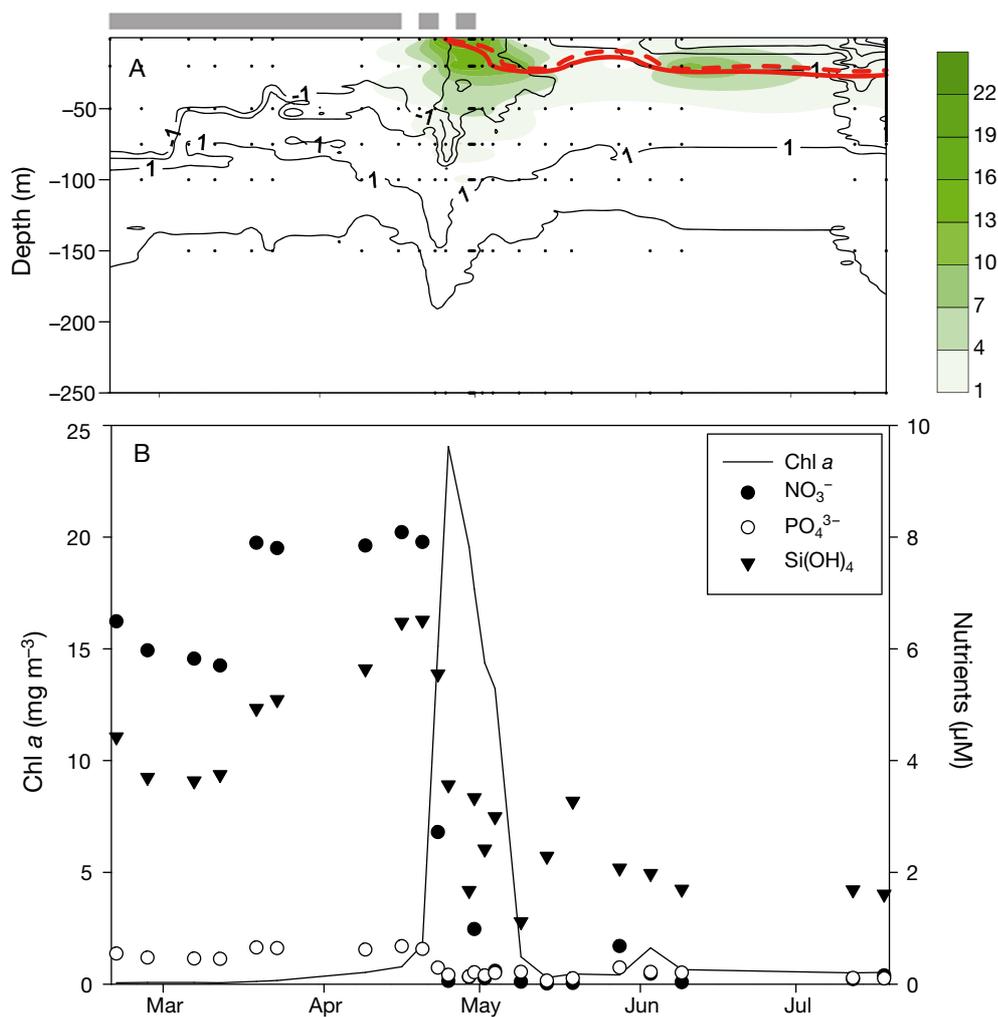


Fig. 2. Water column characteristics over the course of the investigation. (A) Isolines are water temperature ($^\circ\text{C}$) and the green coloration illustrates chlorophyll *a* (chl *a*) concentrations (mg m^{-3}). The concentration of the limiting nutrient nitrate is displayed as red isolines (solid: $1.0 \mu\text{M}$, broken: $0.5 \mu\text{M}$). Points: sampling depths. Grey bars indicate ice cover. (B) Succession in the surface layer (1 m) of chl *a* and nutrient concentrations

layer by a thermocline at approximately 60 m depth (Fig. 2A). During ship-based sampling from 9 April, the weather was dominated by a high pressure system with high irradiance and no wind. The thermocline remained at approximately 60 m depth as the temperature in the upper layers increased to an average of -0.9°C (Fig. 2A). Towards the end of the investigation, from 9 May, the thermocline approached 20 m, separating the water column into 3 water layers with a warm surface layer, an intermediate layer of cold water ($<0.5^{\circ}\text{C}$), and a warm bottom layer (Fig. 2A).

Nitrate was uniformly distributed in the water column at the initiation of the investigation. During the bloom phase the average nitrate concentration in the upper 50 m stratum decreased from 5.4 ± 1.5 to $2.1 \pm 1.0 \mu\text{M}$ as a result of the developing bloom (Fig. 2A,B). Nitrate concentrations decreased below detection level above 20 m after 14 May. Phosphorus and silicate distributions mirrored the nitrate development but were not fully depleted. The surface nitrate concentration remained low after the sedimentation of the bloom, and the nutricline followed the stratification for the rest of the study. Comparison of the major nutrients species (Fig. 3) suggests nitrogen limitation of the primary producers, since nitrate became depleted relative to the Redfield ratios with respect to phosphorus (Fig. 3A) and silicate (Fig. 3B) after the ice break-up and initiation of the bloom.

Phytoplankton biomass

The initiation of the spring bloom (9 April) began at the break-up of sea ice and establishment of the pycnocline. The bloom developed in the upper 20 m from 23 April and averaged at $11.8 \pm 1.7 \text{ mg chl } a \text{ m}^{-3}$ ($n = 20$) during the bloom phase, with a peak of 24.1 mg m^{-3} on 25 April, followed by a rapid sedimentation out of the photic zone (Fig. 4).

In the present study we divided the phytoplankton bloom into 3 phases. The first phase was defined as the bloom developing phase, (23 to 30 April), followed by the decaying bloom phase (2 to 19 May) as the second phase, and the post-bloom phase (28 May to 9 June) as the third phase. The following results are confined to these phases.

Across the 3 depths sampled there was a clear seasonal succession in the size classes of chl *a*. In the beginning of March, chl *a* $<10 \mu\text{m}$ dominated the phytoplankton (Fig. 3). During the spring bloom phase, a succession of the 3 size classes was observed initiated by a peak of chl *a* $<10 \mu\text{m}$, followed by a 10–50 μm peak, and culminating with a peak of chl *a* $>50 \mu\text{m}$. The post-bloom period was dominated by the smallest fraction, i.e. $<10 \mu\text{m}$.

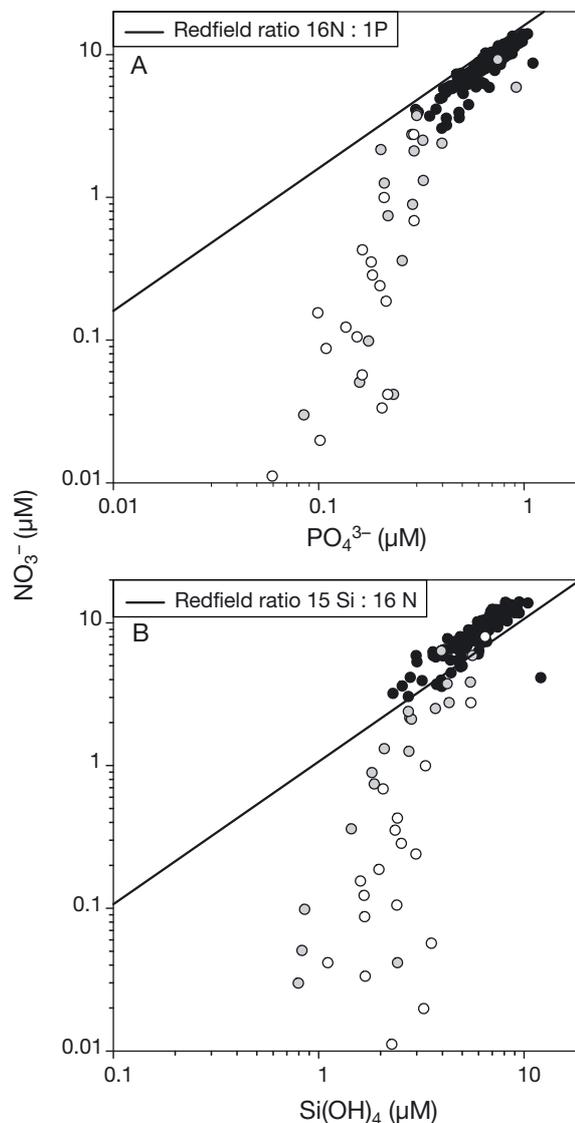


Fig. 3. Relationship between (A) phosphorus and nitrate and (B) silicate and nitrate in the water samples taken from 21 February to 18 July 2008. (●) samples taken throughout the water column before the initiation of the spring bloom and below the photic zone thereafter $>50 \text{ m}$; (○) samples from 20 m after 20 April and (○) surface samples after 20 April. Lines indicate the Redfield ratios of the nutrients in consideration

The same overall patterns were observed at all depths, with the bloom peaking first in the surface. Total chl *a* averaged 12.3 ± 3.4 , 5.9 ± 2.7 and $1.9 \pm 2.2 \text{ mg m}^{-3}$ at 0 to 50 m depth during the 3 bloom phases, respectively.

The smallest chl *a* fraction (0.7 to 10 μm) contributed on average 31% to the total chl *a* in the upper 50 m (Fig. 4). At the surface, this fraction had a high average concentration of $5.0 \pm 2.2 \text{ mg m}^{-3}$ in the bloom developing phase, followed by a subsurface bloom (20 m) with a maximum concentration of $11.9 \pm 0.1 \text{ mg m}^{-3}$ on June 9.

The medium chl *a* fraction (10 to 50 μm) contributed on average 36% to the total chl *a* in the upper 50 m

Table 1. Taxonomic composition of the dominating phytoplankton community during the 3 phases of the bloom (S. Soylyu unpubl.)

Size fraction (µm)	Developing bloom 23–30 April	Decaying bloom 2–19 May	Post bloom 28 May–9 June
<10	<i>Phaeocystis pouchetti</i>	<i>P. pouchetii</i>	<i>Chrysochromulina</i> spp., <i>Pyramimonas</i> spp., <i>P. pouchetii</i>
>10	<i>Fragilariopsis cylindrus</i> , <i>F. pseudonana</i> , <i>Navicula pelagica</i> , <i>P. pouchetii</i>	<i>F. cylindrus</i> , <i>F. pseudonana</i> , <i>Heterocapsa triquetra</i> , <i>N. pelagica</i> , <i>P. pouchetii</i>	
>50	<i>F. cylindrus</i> , <i>F. pseudonana</i> , <i>N. pelagica</i>	<i>F. cylindrus</i> , <i>F. pseudonana</i> , <i>N. pelagica</i>	

(Fig. 4). The fraction showed a clear unimodal distribution, with high average values both at the surface and subsurface of 7.7 ± 1.3 and 6.8 ± 2.1 mg m⁻³, respectively, during the bloom developing phase.

The large chl *a* fraction (>50 µm) contributed 33% to the total chl *a* in the upper 50 m (Fig. 4), with average concentrations at the surface up to 6.4 ± 2.5 mg m⁻³ during the bloom developing phase. Subsurface concentrations showed a clear bimodal distribution during the developing and decaying bloom phase, with a maximum of 14.2 ± 0.04 mg m⁻³ on 9 May, and an average of 5.0 ± 2.5 mg m⁻³.

All chl *a* fractions at 50 m depth had maximum values approximately 3 to 4× lower than the concentrations in the depths above (Fig. 4).

The succession illustrated by the size fractionated chl *a* measurements (Fig. 4) was corroborated by microscopic examinations of the Lugol's fixed samples (Table 1). The winter plankton was dominated by haptophytes (primarily *Phaeocystis pouchetii*). Single cells of *P. pouchetii* also dominated the initial phase (chl *a* <10 µm) of the spring bloom (Fig. 4), followed by a gradual shift to the diatom species of *Fragilariopsis cylindrus* and *Navicula pelagica* after the bloom, first as single cells and later as chain formations during the exponential growth of the bloom. After the bloom peaked and became nutrient-depleted, chains of *F. cylindrus* and *N. pelagica* (>50 µm) sedimented out of the euphotic zone. The summer plankton was dominated by small flagellates <10 µm (Table 1). The same succession patterns were observed in all 3 strata investigated. Further details on the phytoplankton succession and sedimentation are presented in S. Soylyu et al. (unpubl.).

Copepod biomass

In all 5 depth strata sampled, a clear seasonal succession was observed in the species composition of the

copepods. The bulk part of the biomass was mainly located in the upper 50 m, with an average biomass of 3200 ± 800 mg C m⁻², where *Calanus* spp. constituted 96% of the total biomass during all 3 bloom phases. Before the bloom, the biomass consisted primarily of *Oithona* spp., *Pseudocalanus* spp., and *Microcalanus* spp. (Fig. 5). A shift in the biomass composition towards *C. finmarchicus* and *C. glacialis* was observed later in March. During the spring bloom phase, *Calanus* spp. dominated with peaks of high biomasses of *Metridia longa* and *Pseudocalanus* spp. (Fig. 5A). In this phase, the *Calanus*

species was mainly represented as adult females (R. Swalethorp et al. unpubl.). During the decaying bloom phase, the biomass consisted mainly of *C. hyperboreus*, which peaked in early May. During the post-bloom phase the biomass of copepods decreased with *C. hyperboreus* as the dominating *Calanus* spp. and *Pseudocalanus* spp. contributing to the main biomass of the smaller copepods. The biomass decreased with depth, although large biomasses were located in the 50–100 m depth stratum, averaging 2200 ± 600 mg C m⁻², and dominated by *Calanus* (93% of the total biomass) during all 3 bloom phases (Fig. 5B). However, *M. longa* increased in biomass with depth (Fig. 5C,D). A high biomass of *C. hyperboreus* was observed during the spring bloom phase at the deepest sampling depth (Fig. 5E), followed by a low biomass during the decaying bloom phase, which suggests a vertical ascent of *C. hyperboreus* towards the surface during this phase. An increase in the biomass of *C. hyperboreus* was observed during the post-bloom phase at the 2 deepest depth strata (Fig. 5D,E). Here *C. hyperboreus* contributed with 63 and 89% to the biomass, respectively, suggesting a vertical migration of *C. hyperboreus* towards the overwintering depths at the bottom.

Gut fluorescence

Overall, the specific ingestion rate ranged between 0.01 and 74% d⁻¹ for the *Calanus* spp., with an overall average of $10.3 \pm 2.2\%$ d⁻¹ (n = 38) (Fig. 6, Table 2). The specific ingestion rate for *C. finmarchicus* at the initiation of the bloom reached 74% d⁻¹ on 30 April (Fig. 6B). The average specific ingestion rate for *C. finmarchicus* was $17.5 \pm 5.7\%$ d⁻¹ (n = 13) during all 3 bloom phases and decreased significantly in the post-bloom phase (p < 0.01, n = 13).

The specific ingestion rate for *Calanus glacialis* followed the pattern of *C. finmarchicus*, reaching 20%

d^{-1} on 9 May. It decreased significantly in the post-bloom phase ($p < 0.01$, $n = 13$), and the average specific ingestion rate for *C. glacialis* was $7.0 \pm 1.9\% d^{-1}$ ($n = 13$) during all 3 bloom phases (Fig. 6C). The specific ingestion rate for *C. hyperboreus* reached $15\% d^{-1}$ on 4 May, with an average of $2.1 \pm 0.5\% d^{-1}$ ($n = 12$) during all 3 bloom phases (Fig. 6D) and a significant decrease in the post-bloom phase ($p < 0.01$, $n = 12$).

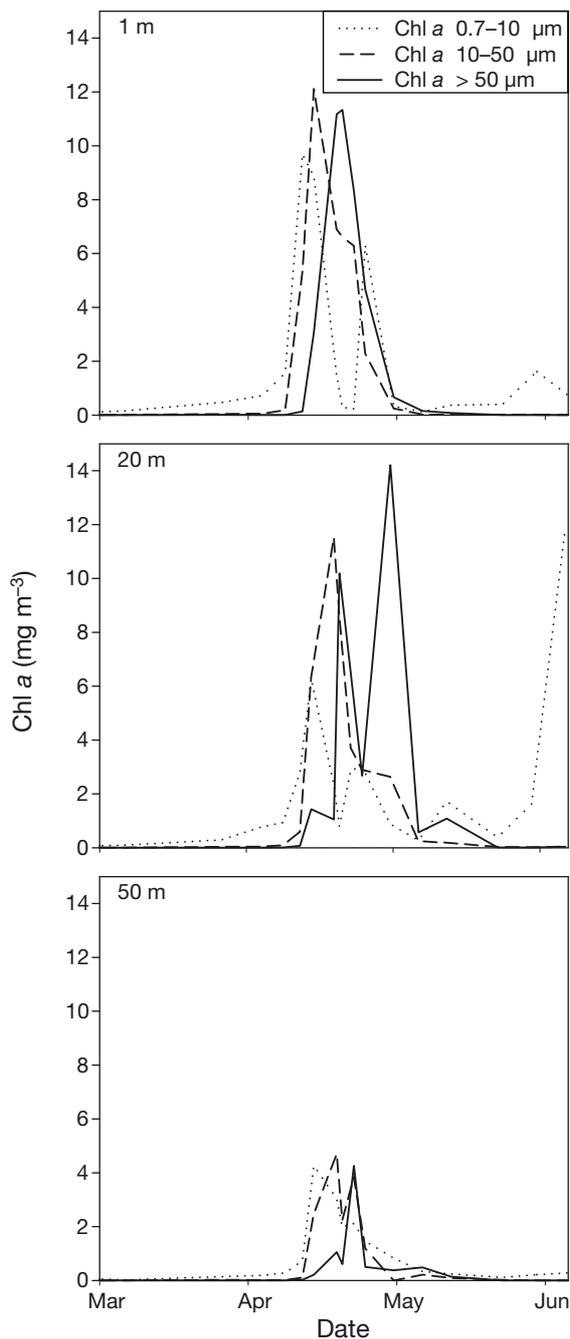


Fig. 4. Size fractionated chl a concentrations (0.7–10 μm , 10–50 μm , and >50 μm) at 1, 20, and 50 m depths

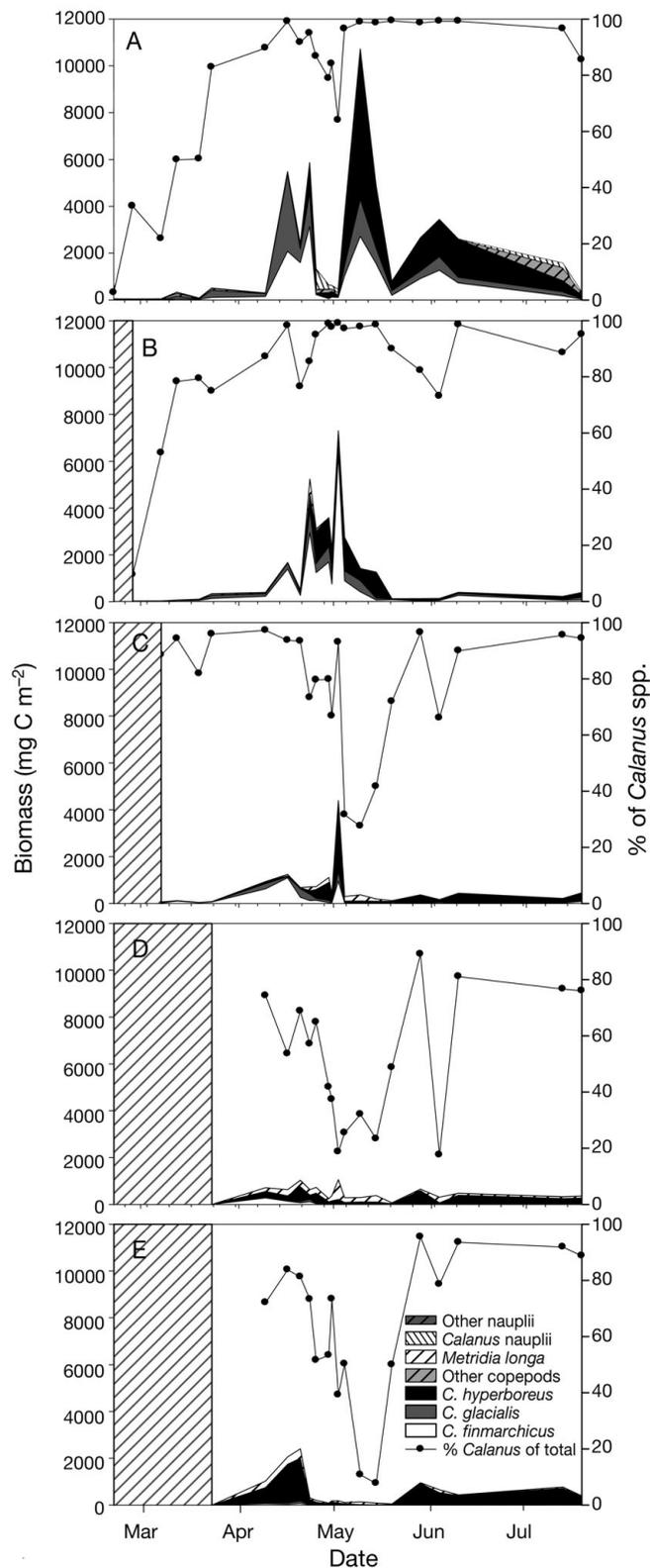


Fig. 5. Integrated biomasses of copepods ($mg C m^{-2}$) in (A) 0–50 m depth, (B) 50–100 m depth, (C) 100–150 m depth, (D) 150–200 m depth, and (E) 200–250 m depth stratum. Solid line: contribution of *Calanus* spp. (%) to total copepod biomass. Vertical black line: initiation of sampling at depths

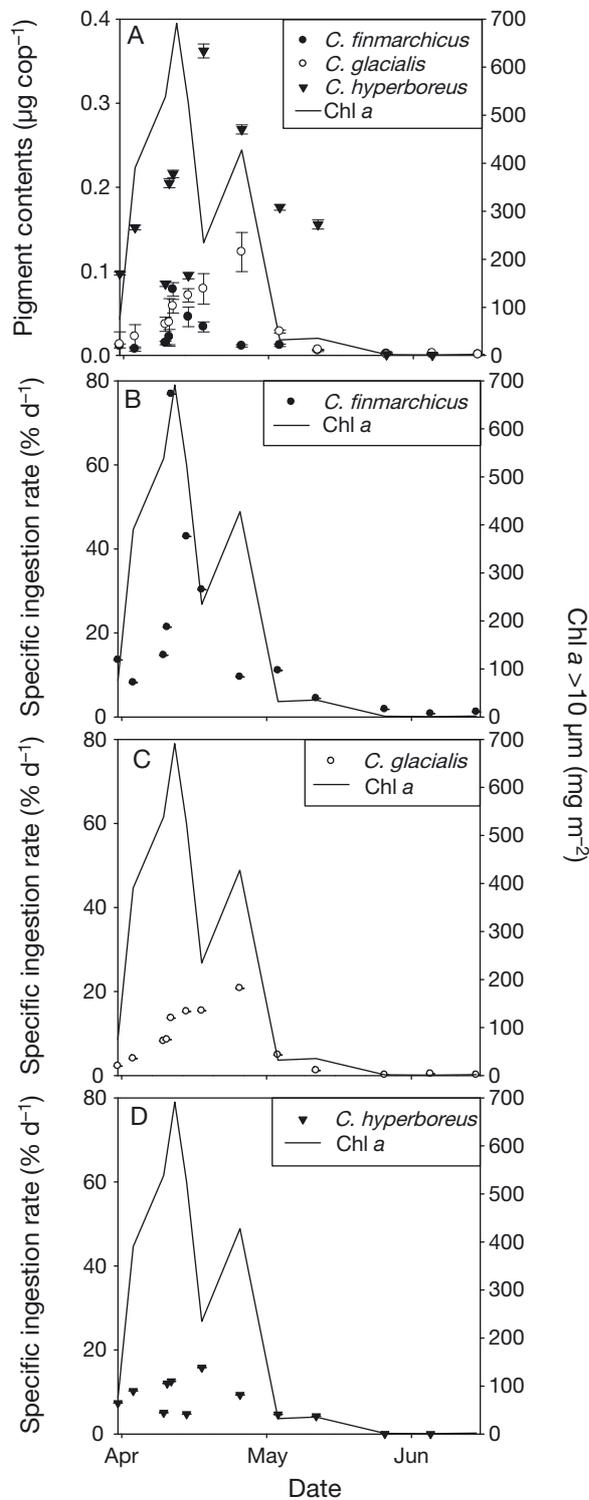


Fig. 6. (A) Gut pigment content (μg) of *Calanus* spp. from gut fluorescence measurements. (B,C,D) Specific ingestion rates ($\% \text{d}^{-1}$) calculated from gut fluorescence measurements for (B) *C. finmarchicus*, (C) *C. glacialis*, and (D) *C. hyperboreus*. Data are averages \pm SE. Solid line: integrated chl *a* $>10 \mu\text{m}$ (mg m^{-2}) for the 0–50 m depth stratum

A correlation analysis of the specific ingestion rates of *Calanus* spp. and the integrated chl *a* size fractions from 0 to 50 m showed that 74, 72, and 57 % of the variation in specific ingestion rates could be explained by the integrated concentration of the chl *a* $>10 \mu\text{m}$ for *C. finmarchicus*, *C. glacialis*, and *C. hyperboreus*, respectively, although not significantly for *C. hyperboreus* ($p > 0.05$, $n = 12$). Furthermore, correlation analyses for chl *a* $>50 \mu\text{m}$ revealed a significant correlation of 89 % of specific ingestion rate by *C. glacialis*, while the correlations of *C. finmarchicus* and *C. hyperboreus* remained unchanged compared to the chl *a* $>10 \mu\text{m}$. Consequently, the ingestion calculated from the gut pigment of *Calanus* spp. was compared with chl *a* $>10 \mu\text{m}$ representing the potential food for *Calanus* (Fig. 6).

All the *Calanus* spp. responded positively to the developing bloom (Fig. 6). A clear response was observed in the gut pigment content of *C. hyperboreus* at the initiation of the bloom, while the responses were more modest for *C. finmarchicus* and *C. glacialis* (Fig. 6A). Likewise, the gut pigment content of all *Calanus* spp. decreased with the termination of the chl *a* bloom.

All background values measured were low; 0.001 ± 0.02 ($n = 45$), 0.002 ± 0.02 ($n = 25$), and $0.01 \pm 0.001 \mu\text{g female}^{-1}$ ($n = 20$) for *C. finmarchicus*, *C. glacialis*, and *C. hyperboreus*, respectively.

In situ faecal pellet production experiment

There was no difference between the *in situ* faecal pellet production measured using the 2 types of fecatrons with different mesh sizes ($p > 0.05$, $n = 89$). Consequently, the results represent an average of the 2 fractions ($n = 8$). No correlation was found between specific ingestion rate by copepod spp. and chl *a* size fractions from the surface water (1 m) ($p > 0.05$, $n = 11$). It was assumed that the food size spectra for the copepod spp. were the same as observed for the specific ingestion rate during the gut pigment measurements and observed in the incubations of individual *Calanus* for faecal pellet production measurements from R. Swalethorp et al. (unpubl.). Consequently, production measurements by the *in situ* faecal pellet productions experiment of *Calanus* spp. are presented along with chl *a* $>10 \mu\text{m}$ representing the potential food for the copepods (Fig. 7).

Faecal pellet production rate (FPR) followed the bloom, with a peak after the culmination of the bloom and a decrease to a stable FPR during the post-bloom phase (Fig. 7A). Overall average FPR was 20 ± 2.2 pellets $\text{ind.}^{-1} \text{d}^{-1}$ ($n = 89$), with a maximum during the decaying bloom of 27 ± 2.9 pellets $\text{animal}^{-1} \text{d}^{-1}$ ($n = 37$). Faecal pellet volumes increased significantly ($p < 0.01$,

Table 2. Integrated phytoplankton and copepod biomass in the upper 50 m and specific ingestion rates and corresponding grazing impact (% of phytoplankton biomass d^{-1} ; PB) estimated by 4 methods: (A) *in situ* faecal pellet production experiment, (B) average gut fluorescence in 3 *Calanus* spp., (C) average faecal pellet (FP) production from bottle incubations of 3 *Calanus* spp., and (D) egg production by *C. finmarchicus* and *C. glacialis*. Data are averages \pm SE. No. of samples given in parentheses

Phase	Biomass ($g\ C\ m^{-2}$)		Specific ingestion rate and grazing impact								(A,B,C,D) Average of produc- tion (% d^{-1})	% of PB
	Phyto- plankton	Cope- pod	(A) <i>In situ</i> FP pro- duction (% d^{-1})	% of PB	(B) Gut fluores- cence (% d^{-1})	% of PB	(C) FP pro- duction (% d^{-1})	% of PB	(D) Egg pro- duction (% d^{-1})	% of PB		
Developing bloom												
23–30 April	18.3 \pm 2.7 (5)	2.7 \pm 1.1 (5)	2.9 \pm 0.6 (29)	0.4	14.1 \pm 4.4 (15)	2.1	2.2 \pm 0.3 (164)	0.3	5.6 \pm 0.4 (93)	0.8	6.4	1.0
Decaying bloom												
2–19 May	9.4 \pm 3.3 (5)	4.0 \pm 1.9 (5)	6.0 \pm 1.3 (37)	2.5	12.5 \pm 2.8 (15)	5.3	2.1 \pm 0.3 (197)	0.9	7.4 \pm 0.5 (133)	3.2	7.2	3.4
Post bloom												
28 May– 9 June	3.8 \pm 2.7 (3)	2.9 \pm 0.3 (5)	3.1 \pm 0.9 (23)	2.3	0.7 \pm 0.3 (9)	0.5	0.5 \pm 0.1 (164)	0.4	1.8 \pm 0.3 (47)	1.4	1.6	1.6

$n = 5563$) throughout the 3 phases, with averages of $1.1 \pm 0.02 \times 10^6\ \mu m^3$ ($n = 1920$), $1.5 \pm 0.02 \times 10^6\ \mu m^3$ ($n = 2283$), and $1.6 \pm 0.04 \times 10^6\ \mu m^3$ ($n = 1360$), respectively (Fig. 8A).

Specific ingestion rates showed a delayed response to increased chl *a*. Specific ingestion rates peaked during the decaying bloom phase at $15.6 \pm 3.8\%$ d^{-1} ($n = 8$), then decreased to $4.3 \pm 0.6\%$ d^{-1} ($n = 8$) and remained constant until the last sampling date (Fig. 7C). The overall average was $6.7 \pm 0.7\%$ d^{-1} ($n = 89$) (Fig. 7C, Table 1).

From the 2 independent methods, *in situ* faecal pellet production experiment and gut fluorescence, copepod specific ingestion rates were evaluated and compared. Furthermore, 2 other measures from a concurrent study were included in the comparison (R. Swalethorp et al. unpubl.). Egg and faecal pellet production were measured for adult females of the 3 *Calanus* species individually in bottle incubations. Here, the average of the 3 *Calanus* spp. is presented for faecal pellet production, and the average of *C. finmarchicus* and *C. glacialis* for egg production, since *C. hyperboreus* were not producing any eggs during the time of the study.

Estimated specific ingestion rates from the bottle incubations were calculated from the specific egg and faecal pellet productions assuming a growth and assimilation efficiency of 33 and 68.2%, respectively (Conover 1966, Hansen et al. 1997) and that specific ingestion rates by *Calanus* could be considered representative for the entire copepod community. Grazing impacts on the phytoplankton community assessed using the 4 methods are estimated by multiplying specific ingestion rate by copepod biomass (Table 2). Differences between the estimated ingestion rates of the 4 methods are evaluated in the 'Discussion'.

Sedimentation

POC sedimentation averaged $1.4 \pm 0.1\ g\ C\ m^{-2}\ d^{-1}$ ($n = 68$) from the 0–100 m depth during all 3 phases (Fig. 9). Sedimentation rates at 20 m were approximately 2 \times higher than at 50 and 100 m. The bulk part of PPC sedimentation was observed in the upper 20 m, with an increasing trend during the developing bloom, ranging from 0.3 to 3.0 $g\ C\ m^{-2}\ d^{-1}$ (Fig. 9). This was reflected by high phytoplankton biomass in the euphotic layer and relatively low grazing rates, suggesting passively sinking PPC from the productive layer. PPC sedimentation rates increased in relation to the subsurface bloom at the end of the post-bloom phase (Fig. 9). PPC contributed with an average of $1.1 \pm 0.4\ g\ C\ m^{-2}\ d^{-1}$ ($n = 5$), equivalent to 60% of POC sedimentation in the upper 50 m during the developing bloom. A relatively lower sedimentation of PPC was observed at 100 m with $0.4 \pm 0.2\ g\ C\ m^{-2}\ d^{-1}$, yet this was equivalent to 57% of POC at 100 m in the developing bloom phase. PPC sedimentation decreased towards the post-bloom phase, with contributions of 18 to 25% of POC in the upper 50 m and 11 to 35% at 100 m in the post-bloom phase. The overall average of PPC was $0.5 \pm 0.06\ g\ C\ m^{-2}\ d^{-1}$ ($n = 78$) at 0 to 100 m depth contributing 36% to the total POC sedimentation.

Seasonal development in sedimentation of amorphous detritus followed the PPC (Fig. 9). Sedimentation of amorphous detritus was of high importance during the post-bloom phase, contributing 71 to 79% to POC sedimentation in the upper 50 m and 10 to 70% at 100 m. On average, the relative importance of sedimenting amorphous detritus exceeded that of PPC both in the decaying and post-bloom phases. The overall average of amorphous detritus was $0.7 \pm 0.1\ g\ C\ m^{-2}\ d^{-1}$ ($n = 68$), contributing 52% to total POC sedimentation.

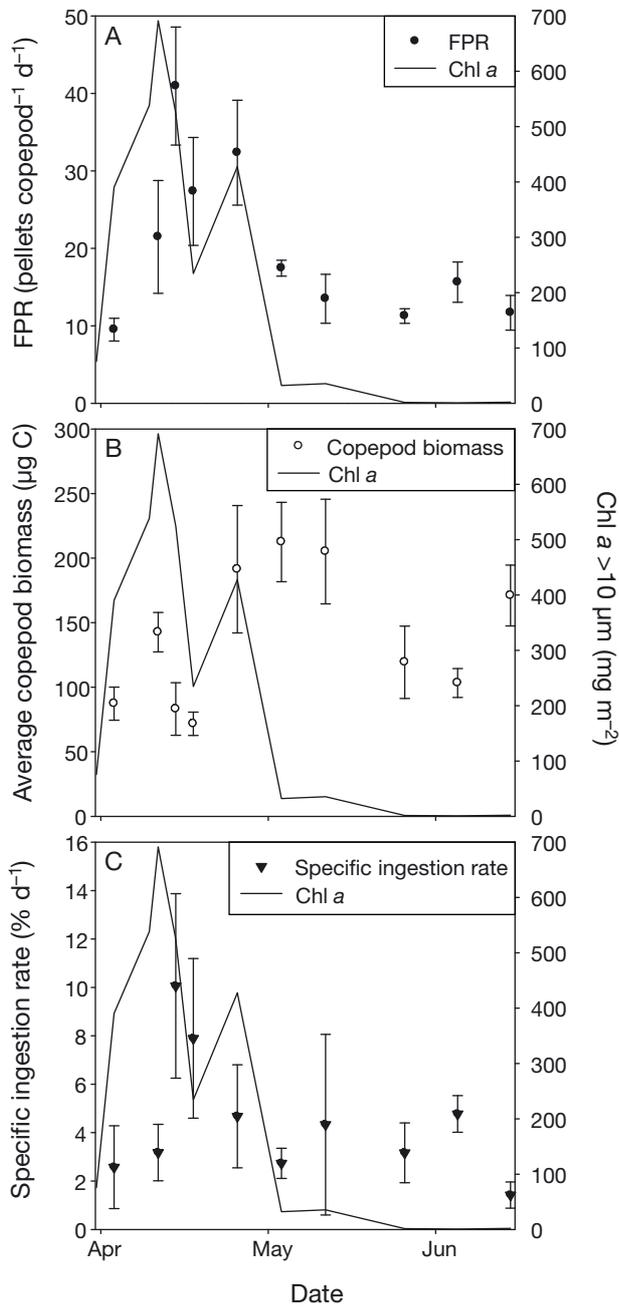


Fig. 7. (A) Faecal pellet production rate (FPR; number of pellets copepod⁻¹ d⁻¹) of the *in situ* copepod community. (B) Individual copepod biomass (µg C copepod⁻¹), (C) specific ingestion rates (% d⁻¹). Data are averages ± SE. Solid line: integrated chl a > 10 µm (mg m⁻²) for the 0–50 m depth stratum

The highest FPC sedimentation was measured at 100 m depth in the decaying bloom phase, ranging from 0.3 to 0.4 g C m⁻² (Fig. 9) corresponding to 10 to 43 % of POC. Sedimentation of FPC at 100 m was significantly different between the decaying and post-bloom phases ($p < 0.05$, $n = 26$). These findings correspond well with larger faecal pellets found in the sediment

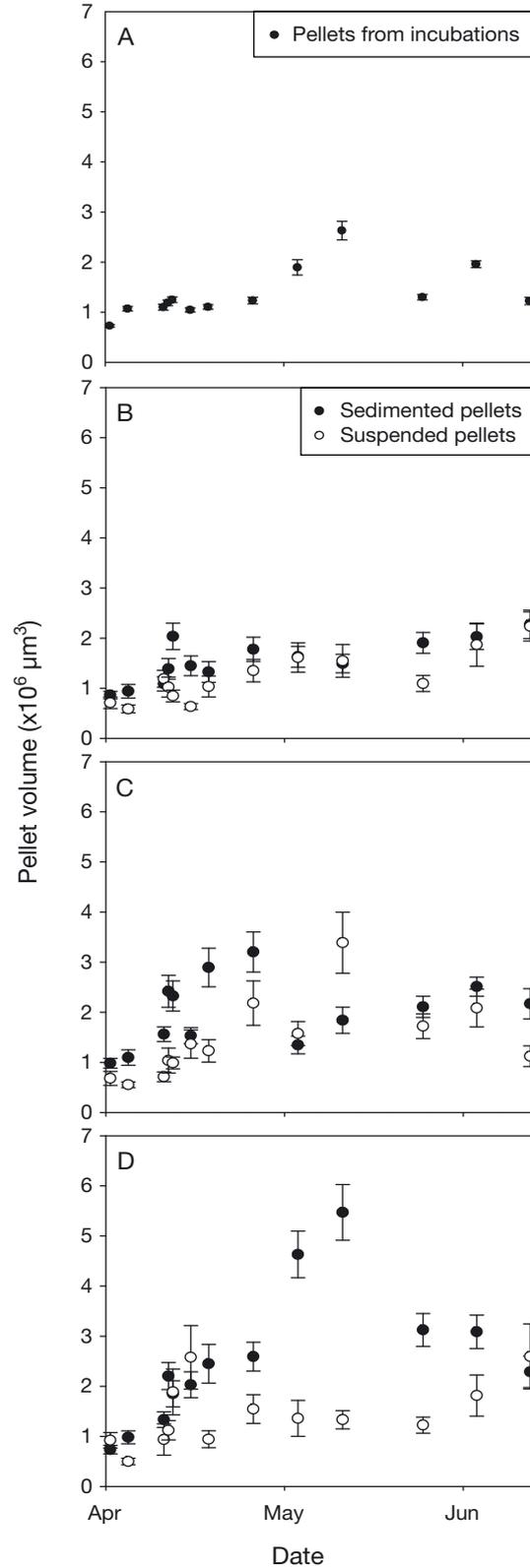


Fig. 8. (A) Volume of faecal pellets from *in situ* faecal pellet production experiments. (B,C,D) Volume of faecal pellets from sediment traps and water column at (B) 20 m, (C) 50 m, and (D) 100 m depths. Data are averages ± SE

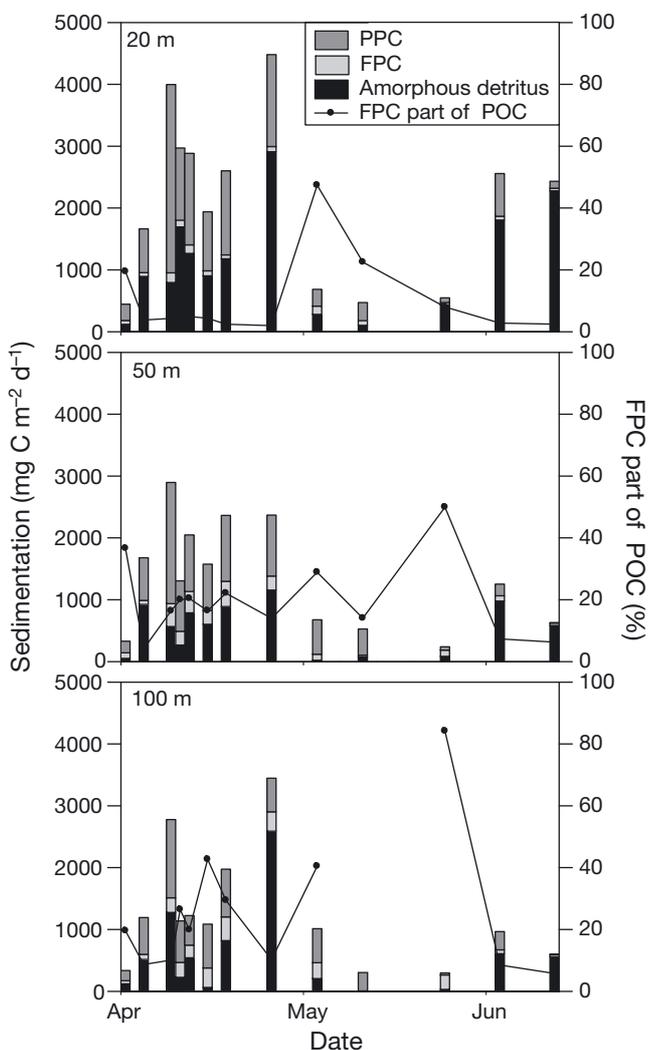


Fig. 9. Seasonal trends in sedimentation of phytoplankton based carbon (PPC), faecal pellet based carbon (FPC), and amorphous detritus fractions of POC ($\text{mg C m}^{-2} \text{d}^{-1}$) at 20, 50, and 100 m depths. Solid lines: FPC part of POC sedimentation. The FPC sample is missing on May 19 at 100 m

traps at 100 m during the decaying phase (Fig. 8D). Faecal pellets found in the sediment traps also differed significantly in size from the suspended faecal pellets at 20, 50, and 100 m depths during the developing and post-bloom phases ($p < 0.01$, $n = 1349$) and ($p < 0.01$, $n = 806$), respectively (Fig. 7B,C,D). The overall average of FPC sedimentation was $0.16 \pm 0.1 \text{ g C m}^{-2} \text{d}^{-1}$ ($n = 78$), contributing 12% to total POC sedimentation.

Biochemical composition of the suspended and settled biomass

In general, the bulk of suspended POC and PON was collected in the upper 20 m, where POC peaked at 887 mg C m^{-3} on 29 April. The POC/PON ratio indicates

the quality of organic material, as degradation increases the ratio from the Redfield ratio of 6.6. The general trend was that the elementary composition of POC/PON ratios increased with depth and throughout the season (Table 3).

The average suspended POC/PON ratio was ~ 7 in the upper 50 m during the developing and decaying phases, with an increase in the post-bloom phase. The increase suggests a nitrate limitation, but no significant correlation was observed between the POC/PON ratio and nitrate concentrations ($p > 0.05$, $n = 9$). None of the suspended POC/PON ratios were significantly different between depths during all 3 phases ($p > 0.05$, $n = 40$) (Table 3). The POC/PON ratio of the settled material during the developing bloom phase at 100 m depth was 6.9 (Table 3). The ratio increased during the decaying bloom phase and ended at 16 in the post-bloom phase at 100 m. This suggests that the settling phytoplankton may have been nutrient-limited during the last phase. However, no significant correlation was observed between POC/PON ratio and nitrate concentration ($p > 0.05$, $n = 9$).

The relation between suspended POC and chl *a* was fitted to a linear regression:

$$\text{POC} = 29.9 \pm 1.3 \times \text{chl } a + 113.9 \pm 10.3 \quad (7)$$

$(r^2 = 0.87, n = 88, p < 0.01)$

The slope of the regression line was used to convert suspended and sedimented chl *a* measurements into PPC.

The suspended POC/chl *a* ratio increased from ~ 50 , measured during the developing bloom phase, to 300 during the post-bloom phase. The ratio did not change significantly with depth during all 3 phases ($p > 0.05$, $n = 56$).

For the settled material, the linear regression between the POC and chl *a* measurements was:

$$\text{POC} = 40.8 \pm 4.2 \times \text{chl } a + 191.8 \pm 55.2 \quad (8)$$

$(r^2 = 0.6, n = 68, p < 0.01)$

The POC/chl *a* ratio of 300 measured for the settled material in the post-bloom phase was significantly higher than for the developing and decaying bloom phases ($p < 0.01$, $n = 36$), which suggests that the settling material consisted mainly of degraded material (data not shown).

The chl *a*/phaeo ratio can be taken as a proxy for grazing, since increased degradation of chl *a* to phaeo pigment led to a decrease of the chl *a*/phaeo ratio. The suspended chl *a*/phaeo ratio decreased from 30 in the bloom developing phase to 5 in the post-bloom phase (data not shown). The ratio in the settled material was 2 in the bloom developing phase at 100 m, after which it decreased to < 1 during the decaying bloom and the post-bloom phases.

Table 3. Concentrations and ratios of particulate organic material (POC and PON) from water column and sediment traps at 20, 50, and 100 m depths during 3 sampling phases. Sedimented POC and subsequent amorphous detritus at 100 m on 23, 25, and 29 April are missing, as is suspended POC at 100 m on 29 April. Data are averages \pm SE. No. of samples given in parentheses

Phase	Depth (m)	Suspended (mg m ⁻³)		Sedimented (mg m ⁻²)		POC:PON ratio (g:g)		r ²
		POC	PON	POC	PON	Suspended	Sedimented	
Developing bloom 23–30 April	20	520.7 \pm 57.7 (7)	90.2 \pm 10.6 (7)	1022.8 \pm 103.9 (10)	158.8 \pm 14.7 (10)	6.0 \pm 0.6 (7)	6.6 \pm 0.5 (10)	0.5
	50	236.0 \pm 23.2 (7)	40.2 \pm 6.1 (7)	644.0 \pm 60.9 (10)	103.5 \pm 10.2 (10)	6.3 \pm 0.6 (7)	6.4 \pm 0.3 (10)	0.6
	100	93.9 \pm 13.9 (2)	11.4 \pm 2.1 (2)	503.3 \pm 27.7 (10)	77.6 \pm 6.6 (10)	9.9 \pm 0.2 (2)	6.9 \pm 0.7 (10)	0.01
Decaying bloom 2–19 May	20	538.3 \pm 90.5 (5)	79.5 \pm 15.5 (5)	822.6 \pm 192.6 (10)	114.0 \pm 25.5 (10)	7.6 \pm 1.6 (5)	7.9 \pm 0.7 (10)	0.9
	50	201.8 \pm 36.8 (5)	34.0 \pm 10.2 (5)	499.9 \pm 99.0 (10)	70.5 \pm 14.0 (10)	6.9 \pm 1.1 (5)	7.4 \pm 0.7 (10)	0.7
	100	120.9 \pm 13.7 (5)	13.9 \pm 1.5 (5)	491 \pm 147.6 (7)	45.3 \pm 10.3 (7)	8.6 \pm 0.4 (5)	11.6 \pm 1.8 (7)	0.4
Post bloom 28 May–9 June	20	269.6 \pm 131.0 (3)	38.0 \pm 17.5 (3)	795.8 \pm 304.0 (4)	50.9 \pm 17.6 (4)	6.9 \pm 0.2 (3)	24.0 \pm 4.4 (4)	0.9
	50	111.7 \pm 30.5 (3)	10.4 \pm 1.8 (3)	296.1 \pm 179.3 (3)	21.2 \pm 4.5 (3)	10.8 \pm 2.9 (3)	13.6 \pm 4.5 (3)	0.9
	100	89.6 \pm 11.5 (3)	8.4 \pm 0.6 (3)	298.3 \pm 114.8 (4)	18.9 \pm 4.7 (4)	10.7 \pm 0.9 (3)	16.0 \pm 3.0 (4)	0.7

Loss rates

Sinking velocity of particulate material increased with increasing depth (except amorphous detritus) (Table 4). Sinking velocity of FPC was approximately one order of magnitude higher than PPC and amorphous detritus (Table 4).

Estimated daily loss rates of PPC due to sedimentation were generally high in the first 2 phases (Fig. 10), with an average daily loss rate of $5.1 \pm 1.6\% \text{ d}^{-1}$ (n = 13) equivalent to a loss of $0.6 \pm 0.01 \text{ g C m}^{-2} \text{ d}^{-1}$ during all 3 bloom phases. The average daily loss rate of PPC due to copepod grazing was $4.1 \pm 1.5\% \text{ d}^{-1}$ (n = 13). This was equivalent to an average loss of $0.2 \pm 0.06 \text{ g C m}^{-2} \text{ d}^{-1}$ during all 3 bloom phases.

Copepod community grazing impact

The specific ingestion rates calculated from the applied methods were variable; however they showed similar trends, with higher rates during the bloom relative to the post-bloom period (Table 2). In general, the lowest estimates originate from bottle incubations (i.e. the egg and faecal pellet production), while the *in situ* measurements (i.e. gut fluorescence and *in situ* faecal pellet production method) resulted in higher rates. Variation in the specific ingestion rate estimates of the different methods varied with bloom phase. The gut fluorescence method values was significantly higher than those originating from the faecal pellet production method, with bottle incubations during the first 2 bloom phases (p < 0.01, n = 21) and (p < 0.01, n = 25), respectively. Ingestion measured from *in situ* faecal pellet production was significantly higher than that from the faecal pellet production from bottle incuba-

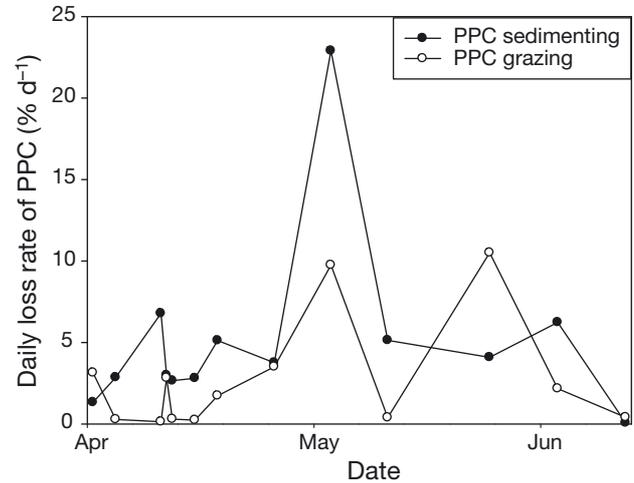


Fig. 10. Daily loss rates (% d⁻¹) of phytoplankton based carbon (PPC), caused by grazing by copepods (Table 1) and sedimentation at the 0 to 50 m depth stratum

Table 4. Sinking velocities of total particulate organic carbon (POC) and its fractions (PPC, FPC, and amorphous detritus) at 20, 50, and 100 m depths during 3 sampling phases. POC and, subsequently, amorphous detritus at 100 m on 23, 25, and 29 April are missing. Data are averages \pm SE. No. of samples given in parentheses

Phase	Depth (m)	Velocity (m d ⁻¹)			Amorphous detritus
		POC	PPC	FPC	
Developing bloom 23–30 April	20	4.1 \pm 0.6 (10)	2.7 \pm 0.6 (10)	117.4 \pm 35.0 (10)	25.5 \pm 14.7 (10)
	50	6.0 \pm 1.0 (10)	5.3 \pm 1.6 (10)	174.1 \pm 59.7 (10)	19.6 \pm 18.3 (10)
	100	10.5 \pm 1.0 (4)	18.9 \pm 2.7 (10)	108.0 \pm 13.7 (10)	6.0 \pm 2.1 (4)
Decaying bloom 2–19 May	20	3.3 \pm 0.7 (10)	3.2 \pm 0.5 (10)	35.5 \pm 5.1 (10)	17.6 \pm 15.5 (10)
	50	6.5 \pm 2.0 (10)	4.7 \pm 0.7 (10)	48.2 \pm 8.8 (10)	29.2 \pm 26.3 (10)
	100	9.4 \pm 2.8 (10)	10.9 \pm 2.9 (10)	188.8 \pm 39.1 (10)	9.1 \pm 5.4 (10)
Post bloom 28 May–9 June	20	8.4 \pm 2.8 (6)	3.0 \pm 1.2 (6)	85.3 \pm 20.0 (6)	13.2 \pm 5.6 (6)
	50	5.3 \pm 3.1 (6)	3.8 \pm 1.7 (6)	140.0 \pm 12.0 (6)	2.8 \pm 1.2 (6)
	100	6.9 \pm 2.1 (6)	10.0 \pm 5.6 (6)	321.7 \pm 104.5 (6)	6.3 \pm 3.9 (6)

tions during the decaying bloom ($p < 0.01$, $n = 25$). During the post bloom, the estimate from the *in situ* faecal pellet production was significantly higher than the other methods (except the egg production [$p < 0.01$, $n = 9$]). The overall averaged grazing rates were significantly lowest in the post-bloom phase ($p < 0.01$, $n = 13$).

Quantification of the daily grazing impact of the copepod community on the phytoplankton based on only daytime measurements of gut fluorescence and *in situ* faecal pellet production requires knowledge about diel feeding periodicity and vertical migration. During the investigation of vertical migration (Fig. 11) we measured gut fluorescence and *in situ* faecal pellet production at 12:00 and 24:00 h (Table 5), and no significant diurnal differences in these rates were detected. However, the biomass of copepods in the upper 50 m doubled from our normal sampling time to around 24:00 h, from 1455 ± 853 to 3147 ± 1168 mg C m⁻² (Fig. 11) due to vertical migration from deeper layers (data not shown). Consequently, our estimates of grazing impact by the copepod are conservative by not considering the increase in biomass around 24:00 h, and the actual grazing impact could be twice as high as our estimates (Table 2).

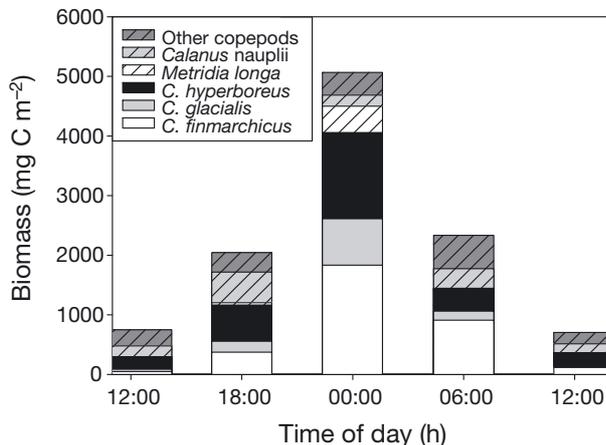


Fig. 11. Average biomass of copepods (mg C m⁻²) in the upper 50 m during a diurnal study from 29 to 30 April

DISCUSSION

Hydrography and plankton succession

In Disko Bay, the sea ice conditions are highly variable (Hansen et al. 2003). In some years, no stable sea ice establishes (Madsen et al. 2008b), while in other years, the sea ice breaks up as late as June (Nielsen & Hansen 1995). In the present study, the sea ice broke up in early April. The spring phytoplankton bloom developed immediately after the breakup of sea ice. Thus, sea ice conditions are very important for the match between the arrival of overwintering copepods to the surface layers and the high spring phytoplankton production, and therefore also for the subsequent sedimentation.

In 2008 the bloom started on 9 April, and the main bloom event (from 23 April) was triggered by a thermocline at 60 m combined with increasing irradiance. Chl *a* concentration was in the same order of magnitude as previously reported from a shallow cove of Disko Bay (Sejr et al. 2007). It was, however, nearly twice as high as previously recorded spring concentrations from the Disko Bay proper (Levinsen et al. 2000a, Madsen et al. 2001). Generally the 3 chl *a* fractions in this study contributed equally to the phytoplankton biomass. However, distinct underlying successions were observed. Prior to the spring bloom chl *a* < 10 μ m

Table 5. Comparison of gut pigment contents (GC; ng total pigment fem⁻¹) of *Calanus* spp. and the specific egestion rate (% d⁻¹) of the copepod community from the *in situ* faecal pellet experiment around 12:00 h (29 April) and 24:00 h (30 April). Data are averages \pm SE. No. of samples given in parentheses

	12:00 h	24:00 h
<i>Calanus finmarchicus</i> (GC)	40 \pm 6 (8)	31 \pm 9 (5)
<i>C. glacialis</i> (GC)	60 \pm 10 (8)	89 \pm 28 (5)
<i>C. hyperboreus</i> (GC)	157 \pm 20 (20)	204 \pm 24 (10)
Specific egestion rate	1.0 \pm 0.3 (13)	0.8 \pm 0.2 (8)

dominated; during the spring bloom a succession of the 3 size classes was observed initiated by a peak of chl *a* <10 μm , followed by a 10–50 μm peak and culminating with a peak of chl *a* >50 μm (diatoms) and a post-bloom dominance of chl *a* <10 μm (haptophytes). The same succession was observed throughout the water column. The spring bloom lasted for approximately 1 wk after nitrate depletion. During the second half of May, melt water from glaciers reduced sea surface salinity and the upper 20 m became additionally stratified. At the end of the investigation chl *a* concentration was low in the surface, and the phytoplankton were concentrated at the nutricline/pycnocline.

Generally, the timing of the ascending copepods appears to match the spring bloom (Diel & Tande 1992), which was also confirmed by the present study, where *Calanus* spp. were present in the surface from the initiation of the bloom. Early appearances of *Calanus* in April have been observed in Disko Bay in 1996 and 1997 (Madsen et al. 2001) and in 2005 (Madsen et al. 2008b). However, the main difference between the present study and the 2 latter was the higher biomass of *Calanus* observed already in April and an early maximum biomass peak in early May. It has been suggested that previously sampling of the copepod community with large mesh size nets (e.g. 200 μm) has led to overestimation of the importance of large species like *Calanus*. We sampled the copepod community with small mesh size nets (50 μm), and still species of *Calanus* comprised up to 99% of the biomass, which is typical for Arctic areas during spring (Hirche 1991, Riser et al. 2007, Madsen et al. 2008a,b). The copepods remained abundant in the upper 100 m throughout the study and did not decrease until early July. *C. hyperboreus* was the only species observed descending to deep waters, whereas *C. finmarchicus* and *C. glacialis* disappeared from the surface but did not aggregate above the bottom as did *C. hyperboreus*. It is possible that they have dispersed out into the bay proper by deeper currents (Holland et al. 2008). This observation is in contrast to the suggestion of a self-sustaining population of *C. finmarchicus* and *C. glacialis* in this area as suggested earlier (Madsen et al. 2001, Niehoff et al. 2002).

Seasonal variability of sedimentation in Disko Bay

During the present study, the sedimentation of organic material increased considerably for all the measured variables. High sedimentation of particulate organic material linked to spring phytoplankton production is common in Disko Bay (Juul-Pedersen et al. 2006, Sejr et al. 2007) and in other Arctic areas (Wassmann 1990, Andreassen et al. 1999, Turner 2002). The

POC sedimentation reported in the present study is, to our knowledge, among the highest reported values during an Arctic spring bloom. It resembles estimates from the spring bloom in a shallow cove in Disko Bay in April (ranging from 1200 to 3000 $\text{mg C m}^{-2} \text{d}^{-1}$ in the upper 5 to 45 m; Sejr et al. 2007), but with a substantial higher PPC fraction of the POC (60% as opposed to 36%), despite the same level of phytoplankton biomass in that study. Sejr et al. (2007) suggested a mismatch between ascending copepods and the spring phytoplankton bloom leads to subsequent low grazing impact by the copepods. In addition, the carbon to chl *a* conversion factor for the sedimented phytoplankton used here was 50% lower, which implies that our PPC contribution to POC could have been 50% higher. Sedimentation of PPC has been reported in similar ranges to ours, during a spring bloom in the central Barents Sea in May-June (Olli et al. 2002, Caron et al. 2004, Reigstad et al. 2008). These studies revealed a close coupling between phytoplankton succession and sedimentation as suggested in the present study. Furthermore, our results of POC sedimentation in the post bloom resembled the values previously found in Disko Bay in June (Juul-Pedersen et al. 2006) and in the central Barents Sea in May (Wassmann et al. 1990, Andreassen & Wassmann 1998). In agreement with the present study, the latter examples report a low contribution of phytoplankton sedimentation relative to carbon in May-June.

Sedimented POC in the present study differed remarkably in composition between two of the bloom phases (developing and post bloom). The above results indicate that much of the vertical reduction (i.e. vertical attenuation) of the settling organic material takes place over a short vertical distance (Fig. 9). The vertical attenuation seems strongest in the upper 50 m, as reported earlier (Riser et al. 2002, Reigstad et al. 2008). The vertical attenuation in the developing bloom of POC was mirrored by amorphous detritus and, to a lesser extent, PPC and FPC. This suggests a selective attenuation of particles where low density particles are being retained above the pycnocline. It is supported by the general lower velocity of PPC in the upper 50 m relative to 100 m depth. This is in contrast to the much higher velocities of the denser faecal pellets observed throughout the water column. If the pycnocline is acting as a barrier to the sedimentation of certain particles, the vertical changes of sedimentation rate would not directly reflect the transformation of the sinking material with depth but rather a differential sedimentation of particles.

In the present study, average FPC contribution to sedimenting POC was low (16%) but within values reported for the Arctic areas in March-May (Wassmann et al. 1999, Riser et al. 2002) and in June

(Andreassen et al. 1996). The relative importance of FPC sedimentation is dependent on copepod abundance, FPR, and faecal pellet size (Besiktepe & Dam 2002, Turner 2002). In the present study, the highest FPC sedimentation was found during the decaying bloom, which is in accordance with the highest FPR (Fig. 7A) and copepod abundance measured.

Nitrogen has previously been shown to be limiting the spring bloom in Disko Bay (Andersen 1981, Nielsen & Hansen 1995, Juul-Pedersen et al. 2006). When nitrate deficiency in phytoplankton growth occurs, an increase in the POC/PON ratio of 6.6 can be expected (Redfield et al. 1963). The high PPC sedimentation during the bloom was probably caused by nitrate limitations, as nitrate decreased in the photic zone (Fig. 2), although average POC/PON ratios found in the suspended and sedimented material were still close to the Redfield ratio. It is likely that phytoplankton aggregation and density dependent coagulation during growth caused the high sedimentation rate (Kjørboe et al. 1994). This is supported by the increased velocity observed at 100 m depth and indicates a depth-related formation of phytoplankton aggregates. During the second half of the decaying bloom, decreased phytoplankton biomass and nitrate depletion led to lower POC sedimentation. PPC attenuation was not observed in the upper 50 m, which suggests that the subsurface bloom was located below the pycnocline and depth of mixing. A majority of the sedimented POC (80%) from the upper 50 m was found at 100 m, thus supporting this notion. Consequently, sedimentation occurs with reduced resuspension and density dependent particle restrictions through the pycnocline. A post-bloom increase of POC sedimentation coincided with a subsurface phytoplankton bloom supported by reintroduced nitrate. The vertical attenuation of POC fractions resembled that observed in the bloom-developing phase. When nitrate was depleted, at the end of the post-bloom phase, both suspended and sedimented POC/PON ratios increased with depth. Phytoplankton growth was probably nitrate-limited, as indicated by POC/PON ratios well above the Redfield ratio in the sedimented material. We cannot, however, exclude a potential influence of increased carbon concentrations from sedimentation of amorphous detritus and faecal pellets, which would increase the POC/PON ratio (Olesen & Lundsgaard 1995, Daly 1996, Daly et al. 1999). Furthermore, potential size dependent sedimentation of phytoplankton (Bienfang 1980, Sarthou et al. 2005) or aggregation (Kjørboe & Hansen 1993, Kjørboe et al. 1994) is not accounted for, which would increase the POC/PON ratio (Olli et al. 2002). The relations between suspended and sedimented carbon and chl *a* were not significantly different ($p > 0.01$, $n = 152$), and the carbon concentration of suspended phyto-

plankton relative to the sedimented biomass did not change. Sedimented POC/PON ratios found in Greenland Sea in June-August consisting of nutrient deficient diatoms (Daly et al. 1999) resembled the findings of the present study. Accordingly, we did not find any correlation between nitrate concentration and sedimented POC/PON ratios.

The high sedimentations reported here underline the importance of the process as a path of carbon transport in Disko Bay. The results indicate a seasonal variability in magnitude and composition of POC sedimentation, where fresh phytoplankton sedimented in the developing bloom and the quality and quantity of the sedimented material decreased towards the post-bloom phase.

Grazing impact of the copepod community — a comparison of methods

The seasonal variation in composition, biomass, and feeding activity of the copepods strongly influenced the benthic-pelagic coupling by shaping the quality and quantity of organic material sinking to the depths. The timing of mesozooplankton migration to match the spring phytoplankton bloom is essential for an efficient pelagic turnover of phytoplankton and for the magnitude of the secondary production (Diel & Tande 1992, Niehoff et al. 2002). Here we applied 4 methods to quantify the grazing impact on the phytoplankton community. The reliability of the measurements depended on season (i.e. the quantitative and qualitative composition of the prey plankton community). The gut fluorescence method directly quantifies the amount of phytoplankton ingested, while pellet and egg production rates through the application of different conversion factors estimate the carbon ingested (i.e. both autotrophic and heterotrophic prey). In addition, the rates are impacted by diel periodicity in feeding and migration. In the present study, no differences could be detected between midday and midnight rate measurements (Table 5), but significant vertical migration took place increasing the night biomass in the upper 50 m. Consequently, our estimated grazing impact could be higher than reported here (Table 2) based on our normal sampling program.

Ingestion calculated from the egg production rate assumes a direct transfer of the ingested food to egg production. This is not always true for copepods that store large amounts of lipids like *Calanus* (Lee et al. 2006 and references therein). Before the bloom, at low food availability, they may start to produce eggs based on stored lipids (Hirche & Kattner 1993, Hirche & Niehoff 1996). At the termination of the bloom, egg production ceases for *C. glacialis*, as the lipid stores

are refueled (S. Kjellerup et al. unpubl.). During the bloom, however, the assumption of egg production depending on food availability holds true for the 2 smallest *Calanus* spp. (Nielsen & Hansen 1995).

In contrast to egg production, faecal pellet production is more directly related to ingestion (Besiktepe & Dam 2002, Seuthe et al. 2007). Hence, lower faecal pellet production was measured during the developing bloom than later in the bloom where more food was available. The individual faecal pellet production of the 3 *Calanus* species was measured in incubations of 24 to 48 h, and there were indications of chl *a* depletion in the incubation bottles in the post-bloom phase (R. Swailethorp et al. unpubl.) leading to underestimations of the faecal pellet production. Another potential problem is ingestion of faecal pellets during the incubation (coprophagy), as no mesh false bottom was used. However, experiments with *Calanus finmarchicus* have showed that faecal pellets were fragmented (coprorhexy) rather than ingested (Noji et al. 1991). Faecal pellet loss due to coprophagy might be unimportant in the present study, but if coprorhexy disintegrate the faecal pellets completely (Lampitt et al. 1990, Noji et al. 1991) the result would be an underestimated faecal pellet production. The *in situ* faecal pellet production experiment measures the instantaneous faecal pellet production in short time incubations, where coprophagy and food limitation should be of minor importance. Another difference to the bottle experiments was that the *in situ* incubations were made on a mixture of the larger stages (i.e. including copepodites) of the dominating copepods. The smaller copepodites will, on average, have a higher specific pellet production than the adult females (Hansen et al. 1997). Accordingly, the *in situ* pellet production was consequently higher than the faecal pellet production from the long-duration bottle incubation (Table 2).

The gut fluorescence method gave the highest grazing estimate of all the methods during the first 2 bloom phases, mainly as a result of a high specific production by *Calanus finmarchicus*, approximately 60% higher than estimated for *C. glacialis* and *C. hyperboreus*. During the 2 first bloom phases the potential food was completely dominated by diatoms. In such situations, the gut fluorescence method can therefore be expected to produce very reliable estimates of copepod grazing. During the post-bloom phase, the highest estimates came from the *in situ* production. Gut fluorescence may underestimate the grazing rate when phytoplankton biomass is low and/or dominated by small spp., since copepods then ingest non-fluorescent food particles such as protozooplankton to sustain production (Ohman & Runge 1994, Hirche & Kwasniewski 1997, Hansen et al. 1999, Levinsen et al. 2000b). Consequently, successful application of the gut fluorescence

method is restricted to phases when herbivory dominates, i.e. during the spring phytoplankton bloom (Kiørboe et al. 1985, Peterson et al. 1990). Thus, a direct correlation between mixed food availability and faecal pellet production seemed to give the most reliable value from the *in situ* production during the post-bloom phase.

The 2 faecal pellet production estimate methods based on direct counting are much more time-consuming than gut fluorescence, but they are a much more reliable tool for estimating the grazing throughout the season. Furthermore, the *in situ* faecal pellet production reduces potential coprophagy and coprorhexy on produced faecal pellets by separating copepods and their pellets. In addition, a short-term incubation period minimized the risk of food depletion during incubation. In contrast, during a long-term experiment, a loss of phytoplankton due to sedimentation and/or grazing during the incubation period can lead to negative effects on the faecal pellet production over time (Besiktepe & Dam 2002, Seuthe et al. 2007), but the long duration is nonetheless necessary when measuring egg production of batch spawning copepods.

In summary, all 4 methods have their strengths and limitations. The choice of method depends on the season, i.e. condition and composition of the phytoplankton community as well as the specific research objective and the time available. Here the overall average was applied to evaluate the copepod community grazing impact on phytoplankton (Fig. 10). Despite a match between the developing bloom and the arrival of *Calanus* to the surface layer, the grazing impact of the copepods was insignificant in the termination of the spring bloom (Table 1, Fig. 10).

The short exponential spring phytoplankton bloom in Disko Bay in 2008 resulted in high phytoplankton sedimentation. The 4 methods used to evaluate the copepod grazing potential concurrently show that grazing impact by copepods played a minor role on the observed phytoplankton succession. Therefore the termination of the phytoplankton spring bloom was primarily caused by sedimentation of nitrogen-limited phytoplankton rather than grazing from the *Calanus*-dominated copepod community.

Acknowledgements. This study was financed by the National Environmental Research Institute (NERI), Aarhus University, Roskilde University (RU), Carlsberg Foundation, Oticon Foundation, ECOGREEN, the Danish Natural Sciences Research Council, and the University of Southern Denmark (SDU). We thank the Arctic station in Qeqertarsuaq and the scientific leader Outi Maria Tervo, University of Copenhagen, who provided us with excellent laboratory facilities and logistical support. At sea, MS 'Porsild' and crew, and captain Finn Steffensen on RV 'Maja S.' with crew provided a great working platform. Also we greatly acknowledge Birgit Søborg in her logistical and technical support.

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Editorial responsibility: Hans Heinrich Janssen, Oldendorf/Luhe, Germany

Submitted: October 19, 2009; Accepted: September 7, 2010
Proofs received from author(s): November 1, 2010