COMMENT

Stable isotope analysis of Humboldt squid prey: Comment on Miller et al. (2013)

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ABSTRACT: Miller et al. (2013; Mar Ecol Prog Ser 477:123−134) used bulk stable isotope analysis (SIA) and a Bayesian isotope-mixing model (SIAR) to assess the diet of the Humboldt squid Dosidicus gigas in the Northern California Current. Their conclusions starkly contrast decades of food habits and other research on this organism. The methods they employed are sensitive to a range of assumptions and parameter estimates, and the trophic model they used is confounded by the interactive effects of movement and diet on D. gigas isotopic values. This is compounded by the sensitivity of the migratory model to critical assumptions of the trophic level of D. gigas (specifically the assumption that large D. gigas are approximately 1 trophic level above copepods), which are inconsistent with the literature. Therefore, Miller et al.’s (2013) conclusions regarding the contribution of specific prey to D. gigas in the Northern California Current are unsupported.

KEY WORDS: Humboldt squid · Dosidicus gigas · Stable isotope analysis · Bayesian mixing models · California Current

Introduction

Miller et al. (2013) assessed the diet of the Humboldt squid Dosidicus gigas in the Northern California Current (NCC) using bulk stable isotope analysis (SIA) and a Bayesian isotope-mixing model (SIAR) to assess the diet of the Humboldt squid Dosidicus gigas in the Northern California Current. Their conclusions starkly contrast decades of food habits and other research on this organism. The methods they employed are sensitive to a range of assumptions and parameter estimates, including the assumption of isotopic equilibrium with prey resources, trophic enrichment factors (TEFs, Δδ15N and Δδ13C), adequate isotopic characterization of prey resources and their variance, the geometry of prey resource isotopic values, and isotopic turnover rates (Martínez del Rio et al. 2009, Parnell et al. 2010, Boecklen et al. 2011, Bond & Diamond 2011, Layman et al. 2012, Ruiz-Cooley & Gerrodette 2012, Smith et al. 2013). Critical assumptions of both the migratory and trophic model in Miller et al. (2013) are inconsistent with the literature. For instance, the trophic model is confounded by the interactive effects of movement and diet on D. gigas isotopic values, and therefore, the conclusions regarding the contribution of specific prey to D. gigas in the NCC are unsupported.

The conclusions reached by Miller et al. (2013) relied on a sequential approach to developing an
understanding of *D. gigas* feeding habits. This included (1) a migratory model to examine the residency of *D. gigas* across 4 regions of the California Current and (2) a trophic model to examine the relative importance of prey resources in the diet of *D. gigas* within the NCC. These modeling approaches have different inherent *a priori* assumptions regarding *D. gigas* being in isotopic equilibrium with their diets at a given time and within a given region. For the migratory approach, there is no assumption of equilibrium; rather, the expectation is that *D. gigas* feeding across regions with isotopically distinct food webs would have isotopic signatures that are intermediate to those food webs, reflecting residency patterns and tissue turnover times (in the case of metabolically active tissues). Thus, mixing model results from the comparison of *D. gigas* isotopic signatures to their potential prey across regions (or sources that reflect production at the base of food webs within each region) are a relative measure of *D. gigas* residency among those locales (Hansson et al. 1997, Hobson 1999, Graham et al. 2010, Carlisle et al. 2012). Conversely, the trophic mixing model implicitly assumes that *D. gigas* is in isotopic equilibrium with prey resources within the NCC over the temporal and spatial scale of the model (Martinez del Rio & Wolf 2005).

The only case where identical *D. gigas* isotopic data can be used in both approaches is when the migratory model indicates that *D. gigas* tissues reflect residency within (i.e. are in isotopic equilibrium with) the region of interest for which potential prey resources are included within the trophic model. This is not the case in the migratory model of Miller et al. (2013), where southern California Current (SCC) end-members contribute considerably (>25%) to the *D. gigas* isotope values (see Fig. 3 in Miller et al. 2013), indicating that *D. gigas* are not in isotopic equilibrium with prey resources within the NCC. Nevertheless, the trophic model only includes isotope data for prey from the NCC, violating a basic assumption of this approach. Although Miller et al. (2013) claim that the contribution of NCC end-members (74%) is sufficient to utilize the trophic model to estimate the contribution of NCC prey to the *D. gigas* they collected, they provide no evidence or reference to support their view that this level of disequilibrium with the NCC did not significantly bias their results.

Furthermore, in their trophic model, Miller et al. (2013) only include isotopic data for prey from nearshore areas (less than 70 km offshore) to isotopically characterize prey across the entire NCC, despite the fact that the distribution of squid is known to extend at least 300 km offshore (Field et al. 2013, Ruiz-Cooley et al. 2013). Due to the strong isotopic gradient between inshore and offshore areas within the NCC (Fig. 3 in Miller et al. 2008, as well as Olson et al. 2010, Ruiz-Cooley & Gerrodette 2012), it would have been important to account for these gradients in the absence of information on isotopic values of potential offshore prey. In particular, the relative effect of foraging in regions isotopically distinct from the coastal NCC—as opposed to prey preferences within the NCC—on *D. gigas* isotopic values needed to be considered, and the inshore–offshore gradient in prey isotopic values within the NCC should have been fully characterized. In the absence of these analyses, Miller et al.’s (2013) conclusions on the contribution of specific prey to *D. gigas* in the NCC are doubtful. Moreover, recent analysis has demonstrated that squid inhabiting the NCC could have originated from multiple regions of Baja California, the Northeastern tropical Pacific and even far offshore waters of the California Current (Ruiz-Cooley et al. 2013), further complicating the assumptions of the migratory model.

**Sensitivity of the migratory model**

While the migratory model as presented by Miller et al. (2013) demonstrates that *D. gigas* are not in isotopic equilibrium with NCC prey resources, another key issue is that the *a priori* assumption that large (~60 cm) *D. gigas* feed at a low trophic level likely inflated the contribution of NCC end-members to *D. gigas* isotope values. While not explicitly stated in the methods, the choice of copepods as baselines for migratory end-members and a single TEF in the migratory model *a priori* presumes that the isotopic signatures of *D. gigas* are a result of feeding exclusively on copepods (or other organisms at the same trophic level as copepods) across their ontogeny and in all regions. As discussed comprehensively in the next paragraph, this assumption is incorrect. While copepods can indicate regional differences in isotopic baselines, the assumed trophic level (TL) difference between a consumer and the species chosen as the baseline will greatly affect mixing model results (Graham et al. 2010, Bond & Diamond 2011).

Decades of research have demonstrated that Humboldt squid are opportunistic and voracious predators, preying primarily on mesopelagic fishes (particularly in oceanic habitats) throughout their distribution (Nigmatullin et al. 2001, Markaida et al. 2008, Rosas-Luis et al. 2011, Field et al. 2007, 2013,
and references therein). In a comparison among a number of these food habits studies between the Gulf of California and the central California Current (essentially between the ‘northern’ and ‘southern’ regions of Miller et al. 2013), mesopelagic fishes were present in 80% of stomachs in both regions, accounting for approximately 80% of the larger (fish and cephalopod) prey items in the Gulf of California and 50% in the California Current (Stewart et al. 2013). Although krill and other macrozooplankton are occasional prey, and in fact their contribution is potentially underestimated due to differences in digestibility and other factors, these studies demonstrate that prey items at a considerably higher trophic level than copepods represent a substantial fraction of the diet of D. gigas.

Moreover, the habitat affinities of D. gigas, which overlap strongly with the range of mesopelagic fishes, provide additional strong evidence of the tight interactions among these species (Stewart et al. 2013). Contrary to the suggestion of Miller et al. (2013) that the discrepancy in food source could have been a consequence of ‘net-feeding’ in trawl-caught squid, <30 of the over 2000 squid stomachs analyzed in the studies summarized by Stewart et al. (2013) were trawl-caught. Instead, nearly all were caught using vertical jigg ing gear that is widely acknowledged to be the least likely method to induce bias in empirical food habits studies of cephalopods.

Finally, in the California Current, the diet composition of Humboldt squid varies substantially across space, particularly between inshore and offshore domains, as well as across seasons, with greater predation on larger prey at larger D. gigas sizes (Field et al. 2013). SIA has also revealed such ontogenetic changes in the trophic position of D. gigas in various ecosystems from the Eastern Pacific, and these patterns differ from system to system (Ruiz-Cooley et al. 2010). As a consequence of this wide body of literature and empirical results, an assumption of feeding at >1 TL above copepods in the migratory model would better reflect what is known about the feeding habits of Humboldt squid—particularly for the larger (mean ± SD: 59 ± 5.6 cm) D. gigas that Miller et al. (2013) collected in the NCC.

We simulated the effect of the assumed trophic level of D. gigas on Miller et al.’s (2013) migratory mixing model results by randomly selecting 76 δ¹⁵N and δ¹³C values from the empirical distribution of D. gigas reported in Table 1 of Miller et al. (2013). We then ran SIAR using the regional copepod end-member values reported in Miller et al. (2013; see their ‘Methods: Data analysis’ section), the simulated D. gigas data, and mean (±1 SD) Δδ¹⁵N and Δδ¹³C of 3.4 ± 1.0 and 0.8 ± 1.3‰, respectively. In the methods section, Miller et al. (2013) state that they used Δδ¹³C of 0.4 ± 1.3‰ in their migratory mixing models but the legends of both their Figs. 2 & 3 state that 0.8‰ were used to generate the results. Because the results graphically depicted in Miller et al.’s (2013) Fig. 3 seem to reflect those given in the written results (their ‘Results: Migratory end-members’ section), we used mean Δδ¹³C of 0.8‰ in our simulation. When we assume 1 TL between D. gigas and copepods in all regions, our results are similar, though not identical, to Miller et al.’s (2013), with a median contribution of 36% for NCC offshore, 26% NCC nearshore, 34% SCC nearshore and 4% SCC offshore (regions as defined in Miller et al. 2013).

However, if we modify Miller et al.’s (2013) TEF assumptions, such that D. gigas arriving in NCC feed just 0.5 TL higher, i.e. 1.5 TL above copepods (adjusted TEF with propagated error: Δδ¹⁵N and Δδ¹³C of 5.1 ± 1.1 and 1.2 ± 1.5‰, respectively), the results change substantially: the contribution of the offshore SCC region rises to 27%, and the SCC overall to 45%, with the 2 NCC regions contributing evenly to D. gigas isotope values. Under this latter scenario, D. gigas is clearly not in equilibrium with any particular region, and this in turn precludes limiting a trophic mixing model analysis of prey contributing to D. gigas diet to only those prey from NCC regions. Assuming a TL of 1.5 above copepods for D. gigas in both the SCC and NCC would further increase the estimated contribution of the SCC region (50% total), particularly SCC offshore (43%), to D. gigas isotope values.

In either of our scenarios (TL of 1.5 for D. gigas in the NCC only or SCC and NCC), the results indicate that the isotopic values of D. gigas collected by Miller et al. (2013) in the NCC are likely and to a great extent the result of foraging in other isotopically distinct regions, rather than solely due to prey preference within the NCC. This strengthens our assertion that the conclusions of Miller et al. (2013) on the contribution of specific prey to D. gigas in the NCC are unsupported. While the seasonal and spatial variability in the relative contribution of prey to D. gigas may not be well known due to the squids’ opportunistic behavior, the assumption that D. gigas feed exclusively 1 TL above copepods is unreasonable; an assumed TL >1 for the migratory model is more appropriate. At a minimum, Miller et al. (2013) should have undertaken the same level of sensitivity analysis on the effect of TL in the migratory model as they did for TEF in the trophic approach (their Fig. 6).
Timing of sample collection

Regardless of the degree of contribution of SCC prey to *D. gigas* (~25 to 50%), additional evidence suggests that squid used in the Miller et al. (2013) study were not in the NCC long enough to reach isotopic equilibrium. Miller et al. (2013) collected samples in July, August and September in Oregon, and they cite Litz et al. (2011) as evidence that *D. gigas* arrived in Oregon sufficiently early to be in isotopic equilibrium with regional prey resources. Litz et al. (2011) report that in over 900 trawls over a 6 yr period (May to November, 2004 to 2009), 2612 squid were encountered in 60 different trawls. However, only 3 of the trawls encountering *D. gigas* (<5% of the total) and 15 of the individual squid (0.5%) occurred before August, despite proportionately greater sampling effort in the May–July period. These data demonstrate an extreme rarity of *D. gigas* in this region prior to August, particularly in nearshore waters. Humboldt squid is a highly mobile predator, capable of horizontal migrations of ~40 km d⁻¹ (Gilly et al. 2006, Stewart et al. 2012). Their virtual absence in the May–July period contradicts the assertion of Miller et al. (2013) that *D. gigas* collected in August and September were all likely resident in the NCC long enough, at least 100 d according to Miller et al. (2013), to approach (~80%), but not reach, isotopic equilibrium with regional prey resources.

Conclusions

Mixing models are sensitive to a wide range of assumptions and parameter estimates (Martínez del Rio et al. 2009, Boecklen et al. 2011, Bond & Diamond 2011, Layman et al. 2012, Smith et al. 2013), and Parnell et al. (2010) explicitly caution that ‘SIAR will always attempt to fit a model’ (p. 5). Miller et al.’s (2013) results do not demonstrate that the *D. gigas* collected were in equilibrium with prey resources of the NCC, under any assumption of trophic level, rendering any assessment of prey contribution to *D. gigas* in the NCC using the approach of Miller et al. (2013) unsupported. No single method is without limitations, and there are rarely sufficient data available in any study to rigorously address every uncertainty. We suggest that additional sampling of squid late into the fall and winter, perhaps combined with retrospective isotopic analysis via sequential sampling of *D. gigas* gladii, could help resolve the timing of migration and residency patterns in order to evaluate if isotopic equilibrium has been reached with prey resources (Ruiz-Cooley et al. 2010, Lorrain et al. 2011, Argüelles et al. 2012).

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