

Sensitivity of the mussel *Mytilus edulis* to substrate-borne vibration in relation to anthropogenically generated noise

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ABSTRACT: Many anthropogenic activities in the oceans involve direct contact with the seabed (for example pile driving), creating radiating particle motion waves. However, the consequences of these waveforms to marine organisms are largely unknown and there is little information on the ability of invertebrates to detect vibration, or indeed the acoustic component of the signal. We quantified sensitivity of the marine bivalve *Mytilus edulis* to substrate-borne vibration by exposure to vibration under controlled conditions. Sinusoidal excitation by tonal signals at frequencies within the range 5 to 410 Hz was applied during the tests, using the 'staircase' method of threshold determination. Thresholds were related to mussel size and to seabed vibration data produced by anthropogenic activities. Clear behavioural changes were observed in response to the vibration stimulus. Thresholds ranged from 0.06 to 0.55 m s⁻² (acceleration, root mean squared), with valve closure used as the behavioural indicator of reception and response. Thresholds were shown to be within the range of vibrations measured in the vicinity of anthropogenic operations such as pile driving and blasting. The responses show that vibration is likely to impact the overall fitness of both individuals and mussel beds of *M. edulis* due to disruption of natural valve periodicity, which may have ecosystem and commercial implications. The observed data provide a valuable first step to understanding the impacts of such vibration upon a key coastal and estuarine invertebrate which lives near industrial and construction activity, and illustrate that the role of seabed vibration should not be underestimated when assessing the impacts of noise pollution.

KEY WORDS: Substrate-borne vibration · Anthropogenic noise · Seismic energy · *Mytilus edulis* · Sensitivity threshold · Marine energy

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INTRODUCTION

Sound energy travels as a longitudinal (compressional) wave, alternately compressing and rarefying the particles across the medium (pressure), and causes an oscillation of molecules parallel to the direction of travel (particle motion) (Van der Graaf et al. 2012). For an underwater sound source encountering a solid, the particle motion may disperse not only via

the water column, but also by the substrate (Hazelwood 2012, Hazelwood & Macey in press); hence, causing 'water-borne' and 'substrate-borne' particle motion. Once in the seabed, the energy may be propagated as longitudinal (compressional, 'P') waves, shear (transverse, 'S') waves, or surface (Rayleigh, 'ground roll') waves (Markl 1983, Aicher & Tautz 1990, Hazelwood & Macey in press), with energy being transmitted in one or multiple waveforms depending

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on the substrate boundary layers, and connection to the substrate (reviewed by Aicher & Tautz 1990). For Rayleigh waves, the energy is confined to the surface of the seabed and the waves are likely to propagate for large distances from the source (Hazelwood & Macey in press). Energy in the substrate may also re-enter the water column at high levels at distance from the original source (Popper & Hastings 2009). Anthropogenic activities, especially those directly in contact with the seabed such as pile driving and drilling, may produce substrate-borne vibrations. Underwater noise has been identified as a major stressor in marine systems and is subject to recent governance initiatives, for example, the European Marine Strategy Framework Directive, which includes underwater energy and noise as one of its 11 descriptors against which 'Good Environmental Status' is measured (Borja et al. 2013). Seabed vibration is not specifically mentioned, yet the consequences of these waveforms to marine life are largely unknown; indeed, there is little information on the ability of invertebrates to detect these waves in general (Roberts 2015).

Detection of substrate-borne vibration has been described in various terrestrial organisms such as spiders, snakes, lizards, scorpions and insects (reviewed by Hill 2001). Semi-terrestrial fiddler crabs *Uca* spp. have also been shown to be receptive to, and indeed to communicate using such substrate vibrations (Salmon & Atsides 1969, Salmon & Horch 1973, Popper et al. 2001). In the marine environment, other crustaceans have detection systems for water-borne particle motion, which may also be used for substrate-borne stimuli (Tautz & Sandeman 1980, Breithaupt & Tautz 1988, 1990, Roberts & Breithaupt in press). Indeed vibration reception, and perhaps communication, seems likely in marine invertebrates since vibrations can propagate large distances through solids, making the seabed an ideal medium for transmission, yet this area is still relatively unstudied (Hill 2001).

There are few data regarding vibration detection in benthic invertebrates; indeed, the research field has not progressed greatly since the review of Frings & Frings (1967). Of the data available, reactions to unquantified vibration stimuli <500 Hz have been observed in decapods, coelenterates, and nudibranchs (Frings & Frings 1967), but the focus has predominantly been upon semi-terrestrial crustaceans, which use vibration during courtship (Salmon & Atsides 1969, Horch 1971, Salmon & Horch 1973).

There is relatively little information regarding the reception and use of vibration by molluscs, as com-

mented upon by Markl (1983). Work has largely focussed upon the more active cephalopods, where reception to water-borne particle motion has been described, with the epidermal lines on the head proposed as the analogue to the fish lateral line (reviewed by Budelmann 1988). More recently ink-jetting by cephalopods has been linked to sound exposure (Fewtrell & McCauley 2012). In bivalves, the specific receptors to detect acoustic and vibrational stimuli are relatively unstudied though there is some support for their vibration reception ability (Mosher 1972, Kastelein 2008), and of detecting particle motion rather than pressure (Ellers 1995). Responses to stimuli described include siphonal retraction, closure of the valves and, in the more active pectinids, jumping from the substrate (Mosher 1972, Ellers 1995, Kastelein 2008), although in many cases the precise levels of vibration are unspecified.

The auditory evoked potential technique (Nedwell et al. 2007) to determine vibro-acoustic sensitivity has been successfully used in cephalopods and one crustacean species (Lovell et al. 2005, Mooney et al. 2010); however, behavioural conditioning produces more accurate thresholds (Ladich & Fay 2013, Sinneros et al. in press). Such behavioural conditioning is difficult in invertebrates, but has been successfully achieved in crustaceans (Offutt 1970). An alternative to this approach is to use small behavioural changes as markers for reception; for example, postural changes, antenna movement and walking leg displacement are commonly used as response indicators in crustaceans (Heinisch & Wiese 1987, Goodall 1988, Breithaupt 2002), or monitoring of respiratory action as demonstrated in cephalopods (Kaifu et al. 2008).

The current study tested the hypothesis that a common intertidal bivalve, *Mytilus edulis* (Mytilidae), would be sensitive to a precise, repeatable and quantifiable source of vibration, and that the stimulus would affect its behaviour. The species occurs on both sheltered and wave-exposed shores (Seed & Suchanek 1992), creating biogenic reefs (Borthagaray & Carranza 2007), and is a common biofouling species of great commercial importance. The sensitivity of this species to vibration has not been previously determined, but, due to a lack of any inner ear or ear-like structure, is likely to involve an array of mechanoreceptors across the body, or a statocyst, as found in other bivalves (Cragg & Nott 1977, Zhadan 2005). To our knowledge, vibration reception has not yet been described for *M. edulis*, an organism adapted to low, moderate and high-energy shores.

MATERIALS AND METHODS

Experiments were undertaken in one session with mussels collected from the intertidal area of Filey Brigg peninsula shore, NE England (54° 13' 02.5" N, 0° 16' 28.3" W). The animals were transported in seawater and placed directly in a glass holding tank (600 × 300 × 300 mm) with a partially sandy substrate, strewn with small rocks for attachment. Mussels were retained in natural groups until testing days and were not specifically fed for the duration of their time in the laboratory. However, the seawater supply to the tank was unfiltered, therefore it is likely that some algae were present in the water, allowing some limited feeding. Valves were not cleared of external fouling organisms (e.g. barnacles), to avoid stressing the animal. Partial water changes (1–2) were undertaken during the period in the laboratory. Subjects were held, at minimum, 72 h prior to experiments.

Experimental setup

A shaker system was used to expose animals to primarily substrate-borne particle motion, with minimal pressure or water-borne particle motion elsewhere in the tank. External ground vibrations affecting the experimental tank (400 × 600 mm) were minimised using a purpose-built layered structure (Fig. 1). A weighted steel frame, completely separate to the base, held an electromagnetic shaker (LDS v.101, sine force 8.9 N, 5–12000 Hz) above the tank, with a carbon fibre stinger rod descending to the substrate. The rod terminated in a plastic cap (35 mm) buried in the substrate to increase vibration propagation. At the other end of the tank an arena (100 × 50 mm) was positioned, consisting of a cylindrical piece of plastic without a base. The arena was screened to eliminate visual disturbance. The experimental tank had a substrate of fine white aquarium sand (depth 30 mm) and a water depth of 150 mm. While mussels attach to hard substratum, this can be a shell or stone on sand. However, such an attachment was avoided here as it would have (1) required cutting the byssal attachment during the experiment, and (2) influenced the vibration received by the animal depending on the strength of byssal attachment.

Inside the arena each mussel was placed with the umbo (adjacent to the hinge margin) into the substrate and the exhalant siphon pointing upwards, and was not restrained in any way. A camera (Microsoft Lifecam web-camera in a subsea housing) was situated above the arena allowing behaviour to be monitored live, and the presentation signal to be modified accordingly. The external monitor for the camera eliminated disturbance by the experimenter.

Each mussel was acclimated in the experimental tank for 1 h without vibration prior to threshold determination. Preliminary tests indicated that a response of an individual to vibration could be classed as full or partial valve closure (a reduction in valve gape by approximately half), hereafter termed 'valve closure' as the vibration reception indicator. Additional responses were observed such as retraction of the foot, a 'twitch' of the valves (minor movement of the valve as if to close, but remaining open) and digging in the sandy substrate.

Shell length (maximum anterior-posterior axis) and shell width (maximum lateral axis) were measured

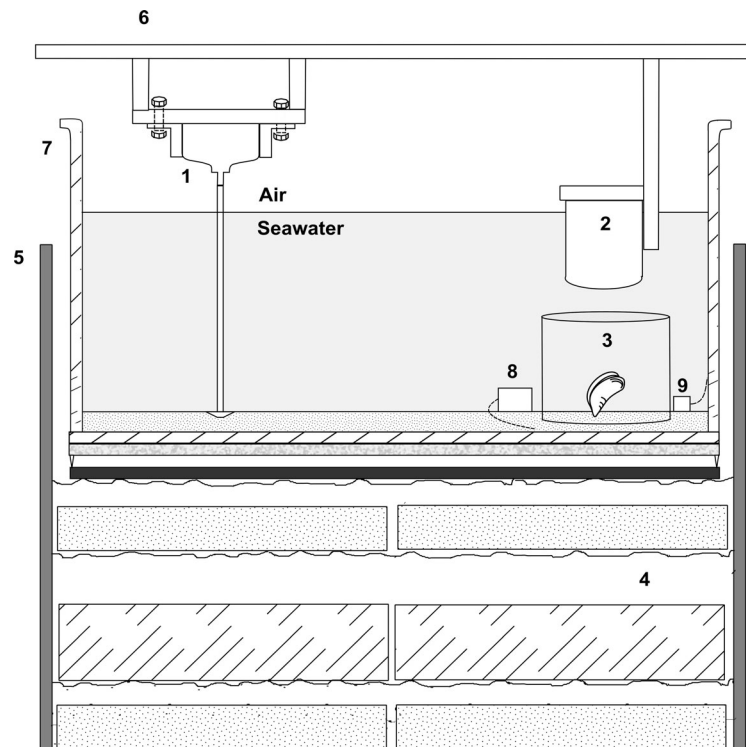


Fig. 1. Experimental setup (not to scale) showing electromagnetic shaker and stinger rod (1), underwater camera (2), experimental arena (3), layered base made up of mixed hard and soft insulation, acoustic dampening and concrete (4), wooden support structure (5), steel frame separate from the base (6), experimental tank with needlepoint legs and sandy substrate (7), position of geophone system (8) and position of accelerometer (9)

after testing (using Vernier callipers, ± 0.1 mm), and a length:width ratio was derived.

Vibration stimuli and threshold determination

Sinusoidal signals (8 s duration), with a 1 s rise and decay time to prevent distortion, were presented at 11 amplitudes (incrementally 6 dB below the maximum amplitude the shaker could produce) (Fig. S1 in the Supplement at www.int-res.com/articles/suppl/m538p185_supp.pdf). Seven frequencies, equally spaced across a frequency range of 5 to 410 Hz, were presented. Signals were played back through a Roland R-09HR MP3 recorder, after being created in AUDACITY (open source, v.2.0.5) and exported on an SD card. The recorder was connected to the electromagnetic shaker and a car amplifier (JL Audio XD 200/2 200 W 2 channel, full range 12–22 kHz) and battery.

Threshold determination was undertaken using the standard 'staircase' method (Cornsweet 1962), which involves exposing the subject to the stimulus and choosing the next signal according to the observed response. A negative response, in this case no valve movement, prompted an increase of the signal and vice versa, until 2 amplitudes were repeatedly presented, with positive and negative responses consistent, i.e. the staircase had reached a plateau. An average of 10 iterations was taken to be the threshold (Cornsweet 1962) (Fig. S2 in the Supplement). Full and partial valve closures were used as the response indicators to calculate the threshold.

A threshold value was calculated at each frequency. At a random point across each test session animals were also exposed to a 'blank' clip (a zero amplitude file, no vibration) to investigate the effect of the equipment itself (hereafter termed control trial). The presentation of frequencies was randomised and an interval of 10 to 15 min was given between frequencies to allow for recovery. Each individual was tested at 7 different frequencies at 11 amplitude levels. Amplitudes were presented 2 to 5 min apart, depending on the duration of response. Two mussels were tested per day, one per session (morning and afternoon) respectively. There was no indication of habituation across the tests, which typically lasted 4 to 5 h. As such no further tests for habituation were undertaken.

Stimulus measurements and signal analysis

Vibrations emitted to the substrate were measured in the vertical axis (m s^{-2} , 1 k s^{-1} sampling rate) using

a waterproofed Brüel & Kjær piezo-electric accelerometer (Type 4333, sensitivity 20.60 mV g^{-1}) connected to a Brüel & Kjær Charge Amplifier type 2635. The accelerometer was placed next to the arena, on the outside, throughout the experiments, as the subjects were likely to interrupt the signal if they came into contact with the sensor (Fig. 1).

Since particle motion is a vector quantity, a 3-dimensional geophone sensor system was used to demonstrate the relative magnitude of velocity in all 3 planes (m s^{-1}) (Sensor Nederland, SM-7 370 Ω , I/O sensor, $28.8 \text{ V m}^{-1} \text{ s}^{-1}$). The geophone was adjacent to the arena on the sandy substrate and was connected to an ADInstrument Powerlab data acquisition module and an IBM Laptop with CHART software (v.5.5.6). The positioning of the geophone was such that the x-axis was between the shaker stinger rod and the arena, the y-axis vertical and the z-axis perpendicular, across the tank. The signal was of greatest amplitude in the vertical axis (Fig. 2), although at 5 Hz the z-axis was slightly greater, perhaps due to interference.

Data from both sensors were recorded simultaneously (and continuously) in CHART. All measurements were made as root mean square (RMS), defined as the square root of the sum of the squared amplitude of the points. All 4 sensor channels were selected simultaneously allowing RMS calculations for the accelerometer and the geophone signals (x, y, z axis). Exactly 6 s of each signal were used for the measurements, with the 1 s rise and fall part of the signal omitted. These values were then adjusted using a correction value (calculated as the difference in RMS between inside and outside the arena) to calculate the vibration received inside the arena, and then were averaged to calculate the threshold value for each frequency.

Spectra of the excitation signals were calculated from all time periods using a 1024 fast Fourier transform and a Blackman window (1 k s^{-1}). In each stimulus frequency there was a prominent peak at the desired frequency with slight variation of signal per experimental session. It is of note that in some cases at 40 Hz there were harmonic peaks due to resonance. At the maximum these peaks were 10 to 30% of the maximum peak amplitude, and as such the 40 Hz results were viewed with some caution (Fig. S3 in the Supplement).

A 6 s sample of background level for each day and frequency was used and averaged to calculate average background levels (RMS) across each experimental day, for the main experimental periods. There was no significant difference between background

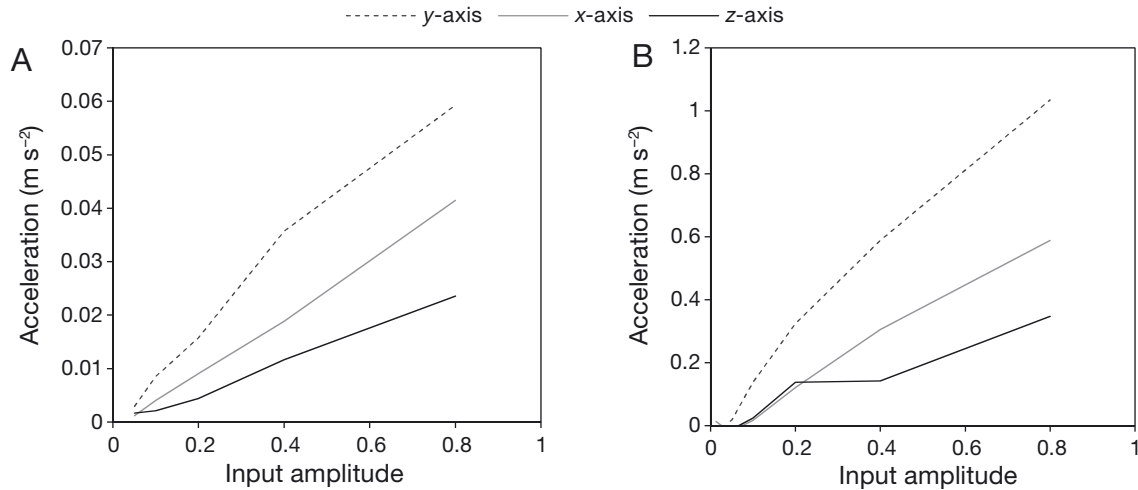


Fig. 2. Relative proportion in each axis (x , y , z ; RMS) of the sinusoidal signal; for example, (A) at 40 Hz and (B) at 210 Hz. Maximum shaker input amplitude is 0 dB, denoted as 1

levels of the compared periods (Kruskal-Wallis $H = 0.68$, $df = 2$, $p = 0.71$); hence, the average background level across all periods (0.0074 m s^{-2} , RMS) was compared to threshold values.

At the end of the experiments, the 4333 accelerometer and the geophone data were calibrated against a type 4370 accelerometer (Brüel & Kjær, sensitivity 80 mV g^{-1}), which was used for the sole purpose of calibration, see Roberts (2015) for details.

Statistical analysis

All data sets were tested for normality (Shapiro-Wilks test) and log transformed as appropriate to fulfill the assumption of parametric tests. Where this was not possible non-parametric tests were used. Mussel thresholds were averaged across individuals at each frequency. Shell width (mm), length (mm) and the shell length:width ratio were correlated with average threshold values (m s^{-2} , RMS) using Pearson's (data separated according to frequency) or Spearman's correlation when appropriate.

Thresholds were related to measurements of vibration taken in the vicinity of anthropogenic operations (measured as velocity, m s^{-1}) (Roberts 2015, Roberts et al. in press). Therefore sine wave equations were used to convert the thresholds from the current work into velocity (m s^{-1}) using the sinusoidal wave equation for amplitude:

$$A = 2\pi fV \quad (1)$$

where A = acceleration (m s^{-2} , RMS), f = frequency (Hz) and V = velocity (m s^{-1} , RMS).

Anthropogenic vibration levels used here are given as the maximum peak amplitude across all axes—the axis of the maximum was not provided in the source literature and therefore it is not known which axis (x , y or z) was predominant in the given signals.

RESULTS

Fifteen adult mussels, shell length 35.7 to 43.8 mm, were tested for sensitivity to sinusoidal waves at 7 frequencies ranging from 5 to 410 Hz. The mussels were deemed healthy as there was no mortality and valve gapes were frequent, gills and siphons were visible, and the foot was sometimes active, leading to partial digging behaviour. Clear valve gape changes were observed in all mussels in response to the vibration stimulus, which were distinct from the valve movements during natural rhythms of feeding. No reactions were observed during control trials. Full and partial valve closure responses were frequent and clearly visible throughout the experiment. On average each mussel reacted to 5 out of the 7 frequencies tested ($n = 15$), regardless of individual and the day tested. The response was similar across all frequencies with an average of 12 reactions per frequency out of 15 (11.57 ± 2.15 , mean \pm SD) (Table 1). With regard to observer bias, given the well-defined criteria for open, closed and partially closed valves, there was little ambiguity in the response and so an independent verification was not needed. However, all experiments were filmed for later reference if required. Furthermore, real-time determination of re-

Table 1. Descriptive statistics for the mussel *Mytilus edulis* threshold experiments, with closure and partial closure used as the indicator of response (n = 15)

| Frequency (Hz) | Threshold (m s ⁻²) (mean ± SD) | n ind. responding |
|----------------|--|-------------------|
| 5 | 0.07 ± 0.008 | 9 |
| 10 | 0.06 ± 0.002 | 11 |
| 20 | 0.08 ± 0.010 | 15 |
| 40 | 0.10 ± 0.012 | 12 |
| 90 | 0.09 ± 0.041 | 13 |
| 210 | 0.55 ± 0.092 | 12 |
| 410 | 0.12 ± 0.014 | 9 |

sponses was necessary given that the observer also had to adjust the exposure level after each presentation. Finally, there was no incentive to bias a yes or a no response, since there were no previously known threshold values to aim towards, or other results to disprove.

The greatest sensitivity to vibration was measured at 10 Hz, with an average threshold of 0.06 m s⁻² (RMS, n = 11) in the vertical direction. Thresholds ranged from 0.06 to 0.55 m s⁻², with an approximately consistent level, but a prominent peak (reduction in sensitivity) at 210 Hz of 0.55 m s⁻² (RMS) (Fig. 3A).

There was a significant correlation between length of mussel (mm) and average threshold value (m s⁻²) (Pearson's r = 0.59, n = 13, p < 0.05, log transformed) (Fig. 3B), but not between width (mm) or length:width ratio and average threshold values (all frequencies together, Pearson's r = 0.50, n = 13, p = 0.08 and r = -0.002, n = 13, p = 0.10 respectively, log transformed). When the data were subdivided according

Table 2. Correlation coefficients (Spearman's ρ) between shell morphology (mm) and average thresholds per frequency (Hz) for *Mytilus edulis*. None of the results were statistically significant (p > 0.05)

| Frequency (Hz) | Length (mm) (ρ) | Width (mm) (ρ) | Length:width ratio (ρ) |
|----------------|-----------------|----------------|------------------------|
| 5 | 0.17 | 0.34 | -0.14 |
| 10 | 0.24 | 0.30 | -0.90 |
| 20 | 0.07 | -0.07 | 0.17 |
| 40 | 0.03 | -0.06 | -0.30 |
| 90 | 0.07 | 0.24 | -0.20 |
| 210 | -0.12 | -0.57 | 0.08 |
| 410 | 0.25 | 0.46 | -0.15 |

to frequency (Hz), there were no significant correlations between the threshold and the morphological variables (Table 2, Spearman's).

DISCUSSION

Sensitivity of mussels to vibrations

Sounds in the oceans are produced naturally by various abiotic sources including waves, bubbles, wind and turbulence. In addition to this, biotic sources can include incidental feeding and communication sounds of marine organisms, hydrodynamic sounds created by shoals and even byssal thread movement of bivalves (Di Iorio et al. 2012). Detection of ambient levels of substrate-borne vibration (and water-borne in some cases) may be advantageous, for example, for detection of waves (e.g. Ellers 1995) or predators.

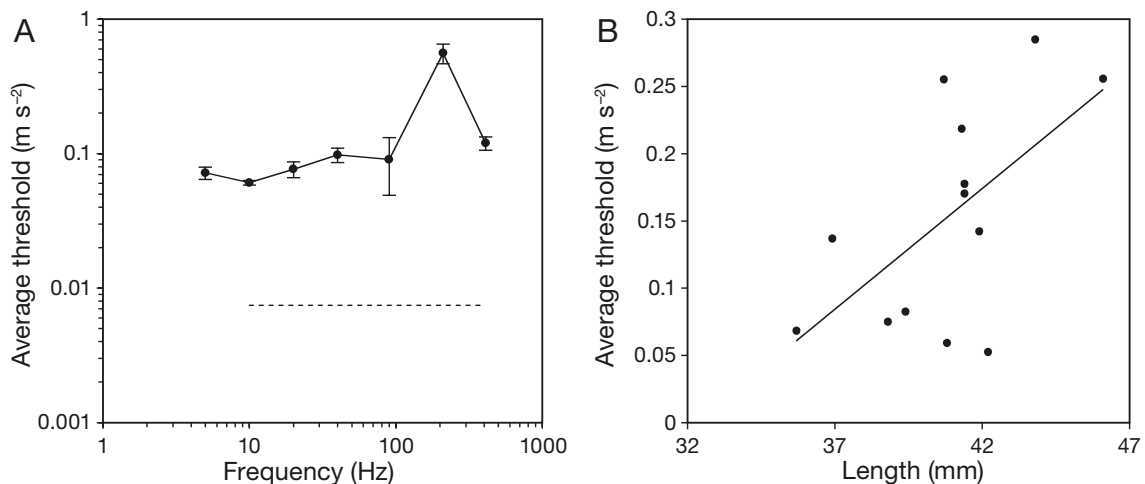


Fig. 3. (A) Sensitivity threshold (m s⁻², RMS, vertical plane) of *Mytilus edulis* (n = 15; mean ± SE) to substrate-borne sinusoidal vibration frequencies (Hz). Average background levels are denoted by the dotted line. (B) Correlation (Pearson's) of shell length (mm) and average threshold (m s⁻²)

In this study, mussels responded to sinusoidal vibratory signals in the frequency range of 5 to 410 Hz. Responses were relatively constant across all frequencies, with a prominent decrease in sensitivity at 210 Hz (0.55 m s^{-2} , RMS). A reduction in sensitivity with increasing frequency such as this has been demonstrated in crustaceans, fish and cephalopods (Salmon & Atsides 1969, Packard et al. 1990, Hughes et al. 2014). In our study, the sensitivity increased again at 410 Hz. Spectral analysis indicated that the 410 Hz was relatively 'pure' in terms of frequency composition; therefore, the reason for the anomaly cannot be determined. Alternatively, it is possible that the 210 Hz value was over-estimated by the accelerometer, possibly affected by resonance within the tank. Use of a non-contact transducer such as a laser Doppler vibrometer (e.g. Breithaupt 2002) to measure the vibration on the mussel valve itself would have been valuable to further understand these results.

Sensitivity to vibration decreased with size (and hence weight) of the mussel. This effect may be caused by the higher inertia of larger mussels. Rayleigh waves will accelerate a mussel resting on the ground in the vertical axis. At a given force a heavier mussel needs more acceleration to be moved in the vertical plane than a lighter mussel. This dependence on size (or weight) is a consequence of the current experimental set up and may not be of biological significance. In nature, mussels are attached to solid objects and depend on transmission characteristics of the solid.

There have been few studies investigating sensitivity of bivalves to vibration (Frings 1964, Mosher 1972, Kowalewski et al. 1992, Ellers 1995, Zhadan 2005, Kastelein 2008). Of these, only one provides detailed measurements of the exposure stimulus (Kowalewski et al. 1992), but focusses upon mortality of larval forms rather than responses of adults. Incomplete or undisclosed descriptions of exposures in other studies make it difficult to fully interpret the results, for example, when references to sensitivity are given without data to confirm these observations (discussed in Roberts 2015). Therefore, there are insufficient data with which to compare the current sensitivity results. Other studies do not provide sufficient details of the vibration stimulus in terms of amplitude, but do indicate the frequency range of reception. For example, vibration sensitivity of *Cerastoderma edule* (Cardiidae) and the swift scallop *Chlamys swifti* (Pectinidae) has been demonstrated in the region of 20 to 64 kHz (Zhadan 2005, Kastelein 2008); and sensitivity <1000 Hz was shown for *Macoma balthica* (Tellinidae), *Mizuhopecten yesso-*

ensis (Pectinidae) and *Donax variabilis* (Donacidae) (Mosher 1972, Ellers 1995, Zhadan 2005).

Whilst threshold data for bivalves are sparse, they are available for other more active molluscs, for example cephalopods (Packard et al. 1990, Kaifu et al. 2008), although these have a more complex nervous system. All of these studies indicate a greater sensitivity to particle motion than *Mytilus edulis*, with threshold amplitudes ranging from 0.0003 to 1.1 m s^{-2} (water-borne particle motion, 1–300 Hz) (Kaifu et al. 2008, Mooney et al. 2010). It is of note that the cephalopod studies used different methodologies, and water-borne stimuli rather than the vibration stimuli used here. However, although cephalopods are more mobile than sessile bivalves, they are still in contact with the substrate. In general, most research focus has been upon crustaceans where threshold sensitivities are reported to be in the range of 0.002 to 0.81 m s^{-2} (RMS acceleration, 20–1600 Hz) (Salmon & Atsides 1969, Horch 1971, Salmon & Horch 1973, Breithaupt & Tautz 1988, Breithaupt 2002, Hughes et al. 2014), which is within the range observed in the current work.

Due to a lack of any specific ear-like structure, 2 receptor systems are likely to be involved in the detection of vibration in non-cephalopod molluscs—the internal system (statocyst) and external system (superficial receptors such as mechanoreceptors on the epidermal layer) (LaCourse & Northrop 1978, Budelmann 1992) (see 'Supplemental text' in the Supplement at www.int-reds.com/articles/suppl/m538p185.pdf). Epidermal sensory cells may be stimulated by hydrodynamic and vibrational changes causing deflection of cilia cells on the body surface (Cragg & Nott 1977, Zhadan 2005) and in some cases may involve specialised abdominal sense organs (Budelmann 1988, Zhadan 2005). It is also likely that vibration travelling through the body may stimulate movement of the statocyst system (LaCourse & Northrop 1978, Ellers 1995, Kaifu et al. 2008, Mooney et al. 2010), as observed in other invertebrates (Budelmann 1988). However, there are few data available on these detection systems in bivalves (Budelmann 1992), although responses to water movements have been observed (Frings & Frings 1967), and there are statocyst descriptions for other bivalve species (Cragg & Nott 1977, Zhadan 2005).

Behavioural responses and implications

In the current work, responses were clear and occurred at onset of the stimulus. The response of *M.*

edulis to fully calibrated vibration sources has not been recorded previously. Responses may be similar across bivalves, where valve closure, siphon retraction and burrowing have been documented (Mosher 1972, Kádár et al. 2005, Kastelein 2008).

With all behavioural experiments involving presentation of stimuli, there is a risk of habituation. As with *Macoma balthica* (Mosher 1972), there was no evidence of habituation in the current work. Nevertheless it would be valuable to explore this further, in particular to assess the repercussions of the response, especially as habituation would not protect them from predators. Furthermore, an investigation into the response variability of the same animal on different days would also be valuable, an aspect already explored with other invertebrates (Roberts 2015).

The closure of the valves in response to a stressor, as seen here, is a costly behaviour in terms of energy, respiratory and heart rate disruption, and impaired excretion ability. For example, a 3 h valve closure has been demonstrated to halve oxygen concentrations within the shell and double carbon dioxide levels (Akberali & Trueman 1979). The energy balance changes with feeding, respiration and excretion; consequently scope for growth (energy balance) and body condition index (longer nutritional and energetic status) are also likely to be affected by valve movement changes; indeed, such changes have been observed in response to chemical pollutants (Widdows et al. 1984, Widdows et al. 2002, Mazik et al. 2013). It is therefore possible that the valve closures exhibited here could affect the overall fitness of the individuals eventually leading to population effects (Widdows et al. 1984). These aspects, and those in relation to synergistic effects of stressors (Mazik et al. 2013) require further study.

Relation to anthropogenic vibration levels

The frequency range tested in the current work (5–410 Hz) was chosen since energy of key anthropogenic acoustic signatures is concentrated at low frequencies (Nedwell et al. 2003), as are many natural sounds (NRC 2005). The frequency range of such energy within the substrate is also likely to be <100 Hz (Subacoustech unpubl., Roberts et al. in press). The longer wavelengths of low frequencies are likely to propagate further and, therefore, are perhaps more likely to be present close to and at greater distances from anthropogenic operations. However, the lack of published field information makes it difficult to relate thresholds to actual values

of anthropogenic signals (Hazelwood & Macey in press). Due to the complexities of underwater sound measurement, many studies only measure sound pressure, without considering water-borne particle motion, or indeed the energy in the seabed. Anthropogenic activities that specifically contact the seabed are of most relevance to the current work—for example, pile driving or those which intentionally produce high levels of substrate vibration, which produce vibrations as compressional, Rayleigh and shear waves (Athanasopoulos & Pelekis 2000, Thandavamoorthy 2004, Hazelwood 2012, Hazelwood & Macey in press).

The threshold of sensitivity determined here ranged from 0.00005 to 0.002 m s⁻¹ (RMS) after conversion to velocity. The sensitivity data determined in our study fall within levels measured near to anthropogenic operations, indicating that the mussels are able to detect such stimuli and show behavioural responses. For example, vibrations measured at 296 m from blasting are greater than the threshold range (6.25 kg charge weight) (Edwards & Kynoch 2008), indicating that the energy would be detectable and would be likely to elicit a behavioural change in mussels. Similarly, vibration levels at 35 m from pile driving (0.9 m diameter pile, muddy substrate, 5 to 50 Hz, Subacoustech unpubl.) are several orders of magnitude higher than the threshold levels, as are those at 22 m from impact drilling (Parvin & Brooker 2008), indicating they would be detectable by mussels at considerable distances from the source. Vibrations measured at 45 m from auger piling (0.75 m diameter auger, 30 m deep) (Parvin et al. 2007) also fall well within the threshold range determined in this study. Details of the measurement procedures for the above data are described in Roberts et al. (in press), and summarised in Roberts (2015) alongside additional measurements. Close to source, strong vibrations probably elicit stronger behavioural changes in mussels, and perhaps injury, although the threshold for damage was not investigated here and there is a lack of published vibration data for comparison. It is of note that a stimulus does not need to be 'detected' to cause damage to an organism, nor does detection necessarily always elicit a response.

Levels of vibration produced by man-made operations will vary significantly according to, for example, the sea-bed composition, type of source and environmental parameters (Thandavamoorthy 2004). Therefore, whilst the data in this study indicate potential detection at specific distances from various source types, actual detection would be scenario-specific. Impulsive signals such as pile driving and

seismic surveys additionally produce a water-borne particle motion and a sound pressure component, which were deliberately not replicated in the current work, as they would confound the observed responses due to interacting factors. Furthermore, the noise from some activities which do not have specific contact with the seabed (such as shipping) may also produce seismic waves in the seabed after propagation through the water (Hazelwood 2012) and therefore be relevant, although levels of these are relatively unknown. The current methodology could be adjusted to incorporate other vibration stimuli, including different anthropogenic recordings, and variation in pure tones such as the duration of the signal. Repeating the tests in the natural environment would also be valuable.

The lack of information regarding seabed vibration is important given the inclusion of underwater noise and energy as an environmental pressure in the OSPAR convention (Oslo/Paris convention guiding international co-operation for protection of the North-East Atlantic) and within the European Marine Strategy Framework Directive (Van der Graaf et al. 2012). These aim to set levels of sound exposure to protect marine species and, for example, by ensuring 'Good Environmental Status' (Borja et al. 2013) even though seabed vibration is not mentioned specifically. By collating sensitivities of a key invertebrate and actual vibration data, the current work emphasises that substrate-borne vibration has a role within noise assessments and therefore the setting of criteria for both substrate-borne and water-borne energy.

Stimulus presentation

In the current work, *M. edulis* was exposed to sinusoidal waves, which were greatest in the vertical plane (horizontal waves were also present to a much smaller degree), although it is difficult to determine the wave type present without further investigation. Rayleigh waves, whilst involving circular motion of particles, excite the substrate in the horizontal and the vertical plane, hence these may be most relevant to the current work (Hazelwood 2012, Hazelwood & Macey in press). These waves are detectable by semi-terrestrial crustaceans such as *Uca pugilator* (Aicher & Tautz 1990), by using receptors in the walking appendages, but data for bivalves are unavailable.

Whilst the vibratory signal here was predominantly substrate-borne, it may also have included water-borne particle motion and perhaps even sound pressure in the experimental tank. However, by using a

shaker directly contacting the substrate, the sound pressure and interference phenomena found in small tanks (Rogers in press) were likely to be minimal compared to the substrate signal. The energy of the signal was predominantly in the vertical axis, but energy was also present in the other 2 planes, and hence the animals may have been responding to this additionally. The present experimental set up was a trade-off between allowing natural behavioural responses and creating a relevant stimulus. Further tests could involve a tank where water-borne particle motion and pressure could also be controlled allowing an investigation of acoustics as well as substrate vibration.

CONCLUSIONS

As with all vibrational and acoustical studies, the results here should be taken within the experimental context, involving a particular exposure duration, frequency range, substrate, vibration stimulus, and species. Extrapolation of the results is not possible since propagation of vibration energy varies according to, for example, substrate, environment and propagation conditions (Kim & Lee 2000). Furthermore, behavioural responses of an individual may be affected by other individual-specific cues such as energy availability, size, respiratory requirements, interactions with conspecifics and perhaps even consistent individual behaviours (reviewed in Roberts 2015). It is not known how energetically costly the behaviours exhibited in the current work were, or to what extent they would affect the long-term fitness of the animals.

The present work has provided a valuable first indication of the sensitivity of a common intertidal species, which is important on an ecological and a commercial scale. The methods are fully reproducible and the vibration stimulus was described in 3 axes; this allows comparisons with future studies. Vibration sensitivity is important within the context of marine noise pollution due to the prevalence of activities contacting the seabed. By comparing sensitivities to field measurements, the data presented here demonstrate that *Mytilus edulis* is likely to detect such vibrations and is likely to exhibit behavioural changes at vibration levels actually produced by operations. This is highly relevant since the shallow, coastal areas occupied by *M. edulis* are also those frequently used for man-made activities. The productivity of mussel beds may be therefore affected by exposure to vibrations, which could have both eco-

system and commercial implications. Hence, this valuable first step towards demonstrating the sensitivity of a common bivalve species to substrate-borne energy clearly illustrates that the effect of seismic waves cannot be underestimated when considering the impact of anthropogenic noise in the marine environment.

Acknowledgements. We are grateful to the UK Department for Environment, Food and Rural Affairs (DEFRA) for funding the doctorate which incorporated this work, and to the following people for discussions of experimental ideas, technical discussions and support throughout: Professor A. Hawkins (Loughine Ltd.), Dr A. Franco (IECS, University of Hull), and Dr R. Pérez-Domínguez (APEM). The authors would also like to thank Mrs S. Jennings and Mr V. Swetez (University of Hull) for animal collection and husbandry. We are also thankful to those who allowed the use of the anthropogenic vibration data: Rockfall Ltd., Shear Design and AECOM UK. Finally we thank 2 anonymous referees for their valuable comments, which have helped improve the manuscript.

Ethical note. The experiments of this study were approved by Hull University Ethics Committee (university ethics reference no. U034). There were no obvious adverse effects on the animals before, during or after experiments. After use, animals were either kept for future experiments in the Hull University aquaria or returned to the shore. Mussels were handled as little as possible throughout the work.

LITERATURE CITED

- Aicher B, Tautz J (1990) Vibrational communication in the fiddler crab, *Uca pugilator*. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 166:345–353
- Akberali HB, Trueman ER (1979) pO_2 , and pCO_2 changes in the mantle cavity of *Scrobicularia plana* (Bivalvia) under normal and stress conditions. *Estuar Coast Mar Sci* 9: 499–507
- Athanasopoulos GA, Pelekis PC (2000) Ground vibrations from sheetpile driving in urban environment: measurements, analysis and effects on buildings and occupants. *Soil Dyn Earthq Eng* 19:371–387
- Borja A, Elliott M, Andersen JH, Cardoso AC and others (2013) Good environmental status of marine ecosystems: what is it and how do we know when we have attained it? *Mar Pollut Bull* 76:16–27
- Borthagaray AI, Carranza A (2007) Mussels as ecosystem engineers: their contribution to species richness in a rocky littoral community. *Acta Oecol* 31:243–250
- Breithaupt T (2002) Sound perception in aquatic crustaceans. In: Wiese K (ed) *The crustacean nervous system*. Springer-Verlag, Berlin, p 548–559
- Breithaupt T, Tautz J (1988) Vibration sensitivity of the crayfish statocyst. *Naturwissenschaften* 75:310–312
- Breithaupt T, Tautz J (1990) The sensitivity of crayfish mechanoreceptors to hydrodynamic and acoustic stimuli. In: Wiese K, Krenz WD, Tautz J, Reichert H, Mulloney B (eds) *Frontiers in crustacean neurobiology*. Springer-Verlag, New York, NY, p 114–120
- Budelmann BU (1988) Morphological diversity of equilibrium receptor systems in aquatic invertebrates. In: Atema J, Fay RR, Popper AN, Tavolga WN (eds) *Sensory biology of aquatic animals*. Springer-Verlag, New York, NY, p 757–782
- Budelmann BU (1992) Hearing in nonarthropod invertebrates. In: Webster DBF, Popper AN, Fay RR (eds) *The evolutionary biology of hearing*. Springer-Verlag, New York, NY, p 141–155
- Cornsweet TN (1962) The staircase-method in psychophysics. *Am J Psychol* 75:485–491
- Cragg SM, Nott JA (1977) The ultrastructure of the statocysts in the pediveliger larvae of *Pecten maximus* (L.) (Bivalvia). *J Exp Mar Biol Ecol* 27:23–36
- Di Iorio L, Gervaise C, Jaud V, Robson AA, Chauvaud L (2012) Hydrophone detects cracking sounds: non-intrusive monitoring of bivalve movement. *J Exp Mar Biol Ecol* 432–433:9–16
- Edwards B, Kynoch JE (2008) Underwater sound pressure and ground vibration measurements taken during blasting in the Ben Schoeman Dock in Cape Town, February 2008. Subacoustech report 780R0102
- Ellers O (1995) Discrimination among wave-generated sounds by a swash-riding clam. *Biol Bull* 189:128–137
- Fewtrell JL, McCauley RD (2012) Impact of air gun noise on the behaviour of marine fish and squid. *Mar Pollut Bull* 64:984–993
- Frings H (1964) Problems and prospects in research on marine invertebrate sound production and reception. In: Tavolga WN (ed) *Marine bio-acoustics*. Pergamon Press, Oxford, p 151–173
- Frings H, Frings M (1967) Underwater sound fields and behavior of marine invertebrates. In: Tavolga WN (ed) *Marine bio-acoustics*. Pergamon Press, Oxford, p 261–281
- Goodall CA (1988) The sensory detection of water borne vibrational stimuli and their motor effects in the Norway lobster, *Nephrops norvegicus* (L.). PhD thesis, Glasgow University
- Hazelwood RA (2012) Ground roll waves as a potential influence on fish: measurement and analysis techniques. (Effects of Noise on Aquatic Life). *Adv Exp Med Biol* 730: 449–452
- Hazelwood RA, Macey P (in press) The intrinsic directional information of ground roll waves. (Effects of Noise on Aquatic Life II). *Adv Exp Med Biol* 875
- Heinisch P, Wiese K (1987) Sensitivity to movement and vibration of water in the North Sea shrimp *Crangon crangon* L. *J Crustac Biol* 7:401–413
- Hill PSM (2001) Vibration and animal communication: a review. *Am Zool* 41:1135–1142
- Horch K (1971) An organ for hearing and vibration sense in the ghost crab *Ocypode*. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 73:1–21
- Hughes AR, Mann DA, Kimbro DL (2014) Predatory fish sounds can alter crab foraging behaviour and influence bivalve abundance. *Proc R Soc B Biol Sci* 281:20140715
- Kádár E, Costa V, Santos RS, Lopes H (2005) Behavioural response to the bioavailability of inorganic mercury in the hydrothermal mussel *Bathymodiolus azoricus*. *J Exp Biol* 208:505–513
- Kaifu K, Akamatsu T, Segawa S (2008) Underwater sound detection by cephalopod statocyst. *Fish Sci* 74:781–786
- Kastelein RA (2008) Effects of vibrations on the behaviour of cockles (bivalve molluscs). *Bioacoustics* 17:74–75

- Kim DS, Lee JS (2000) Propagation and attenuation characteristics of various ground vibrations. *Soil Dyn Earthq Eng* 19:115–126
- Kowalewski J, Patrick PH, Christie AE (1992) Effect of acoustic energy on the zebra mussel (*Dreissena polymorpha*). In Nalepa TF, Schloesser DW (eds) *Zebra mussels biology, impacts and control*. CRC Press, Boca Raton, FL, p 657–666
- LaCourse JR, Northrop RB (1978) A preliminary study of mechanoreceptors within the anterior byssus retractor muscle of *Mytilus edulis* L. *Biol Bull* 155:161–168
- Ladich F, Fay RR (2013) Auditory evoked potential audiometry in fish. *Rev Fish Biol Fish* 23:317–364
- Lovell JM, Findlay MM, Moate RM, Yan HY (2005) The hearing abilities of the prawn *Palaemon serratus*. *Comp Biochem Physiol A Mol Integr Physiol* 140:89–100
- Markl H (1983) Vibrational communication. In: Huber F, Markl H (eds) *Neuroethology and Behavioral Physiology*. Springer-Verlag, Berlin, p 332–353
- Mazik K, Hitchman N, Quintino V, Taylor CJL, Butterfield J, Elliott M (2013) Sublethal effects of a chlorinated and heated effluent on the physiology of the mussel, *Mytilus edulis* L.: a reduction in fitness for survival? *Mar Pollut Bull* 77:123–131
- Mooney TA, Hanlon RT, Christensen-Dalsgaard J, Madsen PT, Ketten DR, Nachtigall PE (2010) Sound detection by the longfin squid (*Loligo pealeii*) studied with auditory evoked potentials: sensitivity to low-frequency particle motion and not pressure. *J Exp Biol* 213:3748–3759
- Mosher JI (1972) The responses of *Macoma balthica* (Bivalvia) to vibrations. *Proc Malacol Soc* 40:125–131
- Nedwell JR, Langworthy J, Howell D (2003) Assessment of sub-sea acoustic noise and vibration from offshore wind turbines and its impact on marine wildlife; initial measurements of underwater noise during construction of offshore windfarms, and comparison with background noise. Subacoustech report 544R0423
- Nedwell J, Turnpenny A, Lovell JM, Parvin SJ, Workman R, Spinks JAL, Howell D (2007) A validation of the dB_{nt} as a measure of the behavioural and auditory effects of underwater noise. Subacoustech report 534R1231
- NRC (National Research Council) (2005) *Marine mammal populations and ocean noise: determining when noise causes biologically significant effects*. The National Academic Press, Washington, DC
- Offutt GC (1970) Acoustic stimulus perception by the American lobster, *Homarus americanus* (Decapoda). *Experimentia* 26:1276–1278
- Packard A, Karlsen HE, Sand O (1990) Low frequency hearing in cephalopods. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 166:501–505
- Parvin SJ, Brooker A (2008) Measurement and assessment of underwater noise and vibration during operations with the Tidal Generation Ltd impact drilling device, Vobster Lake. Subacoustech report no. 810R0204
- Parvin SJ, Workman R, Brooker A (2007) Assessment of underwater noise and vibration during auger piling operations, the River Usk, Newport. Subacoustech report 722R0106
- Popper AN, Hastings MC (2009) The effects of anthropogenic sources of sound on fishes. *J Fish Biol* 75:455–489
- Popper AN, Salmon M, Horch K (2001) Acoustic detection and communication by decapod crustaceans. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 187:83–89
- Roberts L (2015) *Behavioural responses by marine fishes and macroinvertebrates to underwater noise*. PhD thesis, University of Hull
- Roberts L, Breithaupt T (in press) Sensitivity of crustaceans to substrate borne vibration. (Effects of Noise on Aquatic Life II). *Adv Exp Med Biol* 875
- Roberts L, Cheesman S, Elliott M, Breithaupt T (in press) Sensitivity of *Pagurus bernhardus* (L.) to substrate-borne vibration and anthropogenic noise. *J Exp Mar Biol Ecol*
- Rogers P (in press) Parvulescu revisited: small tank acoustics for bio-acousticians. (Effects of Noise on Aquatic Life II). *Adv Exp Med Biol* 875
- Salmon M, Atsides SP (1969) Sensitivity to substrate vibration in the fiddler crab, *Uca pugilator* bosc. *Anim Behav* 17:68–76
- Salmon M, Horch K (1973) Vibration reception in the fiddler crab, *Uca minax*. *Comp Biochem Physiol A Mol Integr Physiol* 44:527–541
- Seed R, Suchanek TH (1992) Population and community ecology of *Mytilus*. In: Gosling E (ed) *The mussel Mytilus: ecology, physiology, genetics and culture*. Developments in Aquaculture and Fisheries Science 25. Elsevier, Amsterdam, p 87–169
- Sisneros J, Popper A, Hawkins A, Fay R (in press) Evoked potential audiograms compared to behavioral audiograms in aquatic animals. (Effects of noise on aquatic life II). *Adv Exp Med Biol* 875
- Tautz J, Sandeman DC (1980) The detection of waterborne vibration by sensory hairs on the chelae of the crayfish *Orconectes limosus*. *J Exp Biol* 88:351–356
- Thandavamoorthy TS (2004) Piling in fine and medium sand—a case study of ground and pile vibration. *Soil Dyn Earthq Eng* 24:295–304
- Van der Graaf AJ, Ainslie MA, André M, Brensing K and others (2012) Report of the technical subgroup on underwater noise and other forms of energy. European Marine Strategy Framework Directive Good Environmental Status (MSFD GES)
- Widdows J, Donkin P, Salkeld PN, Cleary JJ, Lowe DM, Evans SV, Thomson PE (1984) Relative importance of environmental factors in determining physiological differences between two populations of mussels (*Mytilus edulis*). *Mar Ecol Prog Ser* 17:33–47
- Widdows J, Donkin P, Staff FJ, Matthiessen P and others (2002) Measurement of stress effects (scope for growth) and contaminant levels in mussels (*Mytilus edulis*) collected from the Irish Sea. *Mar Environ Res* 53:327–356
- Zhadan PM (2005) Directional sensitivity of the Japanese scallop *Mizuhopecten yessoensis* and swift scallop *Chlamys swifti* to water-borne vibrations. *Russ J Mar Biol* 31:28–35