

Effects of temperature and latitude on larval traits of two estuarine fishes in differing estuary types

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ABSTRACT: Variations in abiotic conditions across large latitudinal gradients can strongly influence the early life history of coastal marine organisms. We investigated the effects of temperature and latitude on the larval traits of 2 estuarine fish species. The arrow goby *Clevelandia ios* and the endangered tidewater goby *Eucyclogobius newberryi* were studied in 18 estuaries along the coast of California, spanning ~8 degrees of latitude. These 2 species were selected because of their dissimilar preferences for estuary type: the arrow goby prefers cooler, fully tidal systems, whereas the tidewater goby prefers warmer lagoons that experience some degree of seasonal isolation from the ocean. Recently settled individuals were collected from July to October 2011, and temperatures within each estuary were recorded to determine how temperature affected larval duration, settlement, and growth rates. Temperatures were more variable among estuaries inhabited by the tidewater goby (10°C range) than among those inhabited by the arrow goby (5°C range). Larval traits of both species differed among sites, but more so for the tidewater goby, a difference that was tied to the greater differences in temperatures among sites in the seasonally closed estuaries it inhabited. In both species, fish that experienced warmer temperatures had shorter larval durations and faster growth rates and were smaller at settlement. Since the length of the larval period has been related to survival and dispersal distance, future variations in temperature due to climate change could have predictable influences on population density and connectivity in estuarine species.

KEY WORDS: Temperature · Latitude · Estuarine fishes · Larval duration · Settlement · Growth rate

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INTRODUCTION

The vast majority of marine fishes begin their life as planktonic larvae, a critical stage that is highly sensitive to changes in the abiotic environment, such as temperature, which can influence larval traits associated with development, survival, and dispersal potential (Houde 1989, Cowen 1991, Gillooly 2001, Gillooly et al. 2002, Brown et al. 2004, McCormick & Hoey 2004, O'Connor et al. 2007, Takahashi et al. 2012). The predicted increases in coastal temperatures due to climate change will likely affect fish

reproduction and larval development, but these effects will vary with latitude and habitat type (Pankhurst & Munday 2011). Controlled laboratory experiments have revealed effects of temperature on the pelagic larval duration (PLD), size at settlement, and growth rates in tropical reef fish, caused by as little as a 3 to 5°C variation (McCormick & Molony 1995, Green & Fisher 2004). In addition, variations in temperature have been found to significantly affect metamorphic success, behavior, and dispersal distance in marine fish species (Rutherford & Houde 1995, Wilson & Meekan 2002, Meekan et al. 2003,

O'Connor et al. 2007), which has the potential to significantly influence population dynamics and community level processes (Gillooly 2001, O'Connor et al. 2007). Since temperature naturally varies over large latitudinal gradients, studying the effects of temperature on fish larvae across these gradients may be useful in predicting some of the long-term impacts of climate change.

In regions with Mediterranean climates, such as California, climate variations are predicted to increase with accelerated human population growth and continued climate change, resulting in warmer, drier, and more variable weather patterns (Valiela et al. 2009, Klausmeyer & Shaw 2009). These predicted changes in climate have already been observed in the coastal zones of southern and central California (Hayhoe et al. 2004). As a consequence of their broad spatial distribution (>1000 km north to south) and tidal patterns, California's estuarine habitats experience significant variability in temperature, both spatially and temporally.

The majority of these estuaries are partially or completely isolated from marine tidal influence either seasonally or episodically due to formation of a sandbar at the estuary mouth (Jacobs et al. 2011). Such isolation typically occurs in the summer months when reduced rates of precipitation lead to reduced freshwater input (e.g. Cooper et al. 2013). In addition, the reduction of tidal exchange causes longer residence time of water, permitting warmer and more variable temperature regimes (Cousins et al. 2010). This phenomenon of estuary closure provides beneficial conditions for a number of endemic and endangered species that use the closed estuaries at some point during their life history (Swift et al. 1989, 1993, Bond et al. 2008).

We studied 2 closely related species of small fish that are year-round residents of estuaries or protected coastal bays, the tidewater goby *Eucyclogobius newberryi* and the arrow goby *Clevelandia ios*. The tidewater goby is found almost exclusively in estuaries that close seasonally or episodically (Jacobs et al. 2011), whereas the arrow goby is found primarily in estuaries and protected bays that retain their connection to the ocean throughout the year. Both species do not exceed 55 to 60 mm standard length (SL) as adults (Miller & Lea 1972) and are benthic microcarnivores that feed on small invertebrates. Where they are present, these 2 fishes are often the most abundant fish in the community (Brothers 1975, Swenson 1999). Their high abundances (exceeding 100 m⁻²; Steele et al. 2006, B. T. Spies pers. obs.) suggest they are important secondary consumers in the

habitats they occupy and are important prey for a variety of larger species, primarily larger fishes. Both species lay demersal eggs from which larvae hatch that likely complete their entire larval development within their natal site before settling and becoming benthic juveniles (Brothers 1975, Swenson 1999).

The tidewater goby is endemic to California, where it is found in a variety of shallow, brackish water estuaries that experience seasonal closure (Swift et al. 1989, Moyle 2002). It was listed as federally endangered in 1994 (USFWS 2005), mainly due to the loss and degradation of suitable habitat. The tidewater goby is considered an annual species, with reproduction occurring year-round, typically peaking in the early spring and late summer (Swenson 1999, USFWS 2005). The estuaries that this species occupies, which are partially or completely isolated from marine tidal influence at times, have lagoonal conditions that are highly variable in salinity, water quality, and temperature. This habitat preference requires the tidewater goby to tolerate highly variable conditions, including salinities ranging from 0 to 41 ppt and temperatures from 9 to 25°C (Swift et al. 1989, Swenson 1999). Larval tidewater gobies are typically restricted to lagoonal habitats during summer months when the estuary mouth is closed (Lafferty et al. 1999a,b), which has created extensive local genetic subdivision due to lack of larval dispersal among estuaries (Barlow 2002, Earl et al. 2010). Thus, it is likely that tidewater goby larvae experience much greater variation in temperature among estuaries than arrow goby larvae, due to the isolation of the lagoons they inhabit.

The arrow goby *Clevelandia ios* is the sympatric sister species of the tidewater goby (Dawson et al. 2002). It has a much broader geographic range, extending from the Bahia Magdalena, Baja California Sur (C. Swift pers. comm.), to British Columbia (Miller & Lea 1972). Arrow gobies also have a prolonged breeding season, usually peaking between February and June (Prasad 1959, Brothers 1975), with a lifespan of 1 to 3 yr depending on locality (Brothers 1975). The open, fully tidal bays and estuaries that the arrow goby typically occupies are usually cooler and higher in salinity than those occupied by the tidewater goby. This differing habitat preference of the arrow goby provides it with a greater dispersal potential via marine larvae, which likely explains its broader geographic range and shallow phylogeographic structure (Dawson et al. 2002). However, a lack of genetic subdivision among populations does not imply frequent dispersal among them (Wang 2004) because as few as 1 migrant per

generation can cause them to be genetically homogeneous (Wright 1931, Mills & Allendorf 1996). Currently, it is unclear how frequently marine dispersal occurs in this species; however, arrow goby larvae are believed to develop and settle primarily in their natal estuarine habitats. This conclusion is based on the limited number of larvae and transitional specimens that have been collected offshore (Brothers 1975) and the common occurrence of all larval stages in estuaries (authors' pers. obs.). Therefore, in this study, we assumed that arrow gobies were collected in their natal estuaries.

The goal of our study was to determine the effects of temperature and latitude on tidewater and arrow gobies, which differ in habitat preference and larval dispersal potential. Although there is a strong correlation between temperature and latitude, which has been used to describe many patterns in larval ecology and development, there are many other environmental factors that change with latitude, including precipitation, day length, the degree of seasonality, and primary production. In this study, we treat latitude as a proxy for a suite of environmental factors that vary over a latitudinal gradient, and we measure temperature directly. We examined 3 main questions: (1) Do larval traits (larval duration, settlement, growth rate) vary latitudinally in both species across their geographic range of overlap? (2) How do temperature regimes vary latitudinally in seasonally closed vs. fully tidal systems? (3) Are variations in larval traits related to variations in the average temperature experienced during the larval phase or to other environmental factors related to latitude?

MATERIALS AND METHODS

Study sites

A total of 18 study sites along the California coast were sampled from July to October of 2011 (Fig. 1, Table 1). Study sites consisted of 8 fully tidal estuaries inhabited by the arrow goby and 10 seasonally closed estuaries inhabited by the tidewater goby. Study sites for the arrow goby listed from north to south were Arcata Bay, Bodega Bay, San Lorenzo River, Elkhorn Slough, Morro Bay, Carpinteria Salt Marsh, Colorado Lagoon, and Los Peñasquitos. Study sites for the tidewater goby from north to south were Ten Mile River, Salmon Creek, San Gregorio Creek, Rodeo Lagoon, Moore Creek, San Luis Obispo Creek, Santa Ynez River, Arroyo Burro Lagoon, Santa Clara River, and Las Flores Marsh. Sites were

chosen based on 2 characteristics: (1) the presence of healthy and abundant populations of the arrow goby or tidewater goby and (2) estuary mouth dynamics: fully tidal or seasonally closing (Fig. 2).

Collections and otolith analysis

Recently settled gobies were collected between August and October of 2011 (Table 1). Due to permitting logistics and to minimize temporal sampling bias, arrow goby collections began in late August and were completed within a 24 d period. Because of low initial sample sizes collected at Colorado Lagoon, an additional collection date was later added in late October. Tidewater goby collections began in late September and were completed within a 36 d period. Small (9.8 to 24.8 mm) juvenile gobies were collected from each site, though previous estimates of the length at larval to juvenile transformation for the tidewater goby (13.4–13.8 mm) and arrow goby (14.2–17.2 mm) were used as target sizes (Moser 1996). Sampling occurred for both species in water depths < 1.2 m between 05:00 and 11:00 h, using either a 3.7 × 1.2 m beach seine with a 1.6 mm mesh, or a 1-man push net with 1.6 mm mesh (Strawn

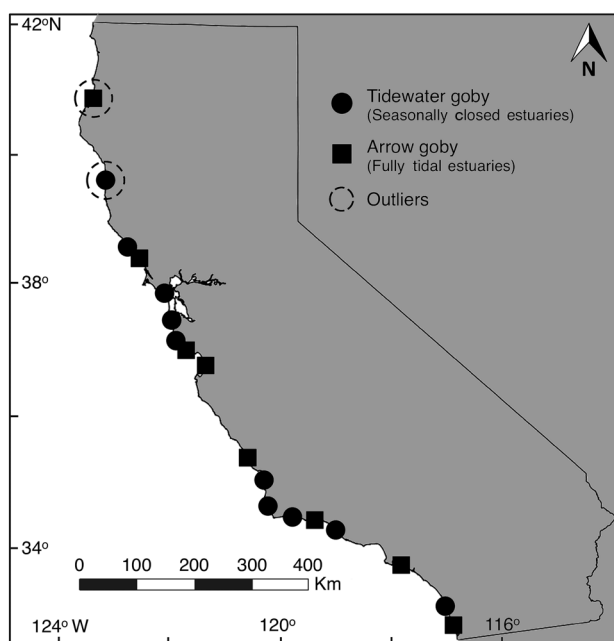


Fig. 1. California coastline showing the 18 study sites. The arrow goby (square) was collected at 8 fully tidal estuaries; the tidewater goby (circle) was collected at 10 seasonally closed estuaries. The 2 sites furthest north, Ten Mile River and Arcata Bay, were treated as outliers (broken circle) for portions of the analysis (see 'Materials and methods')

Table 1. Collection and habitat information for the tidewater goby and arrow goby. The 18 study sites are listed from north to south for each species

Collection date	Study site	Latitude (°N)	Habitat size*	Closed/Open @ collection	Temp. (°C) Mean ± SD	Std. length (mm) Mean ± SD	n
Tidewater goby							
10/15/11	Ten Mile River	39° 32' 43.86"	Large	Open	15.33 ± 0.14	14.69 ± 1.16	39
09/25/11	Salmon Creek	38° 21' 10.87"	Medium	Closed	18.70 ± 0.34	14.93 ± 1.36	40
09/30/11	Rodeo Lagoon	37° 49' 54.41"	Medium	Closed	17.40 ± 0.31	15.48 ± 1.15	40
10/16/11	San Gregorio Creek	37° 19' 14.29"	Medium	Closed	18.86 ± 0.32	14.48 ± 1.60	40
10/07/11	Moore Creek	36° 57' 04.50"	Small	Closed	21.59 ± 0.20	16.17 ± 1.31	40
10/07/11	San Luis Obispo Creek	35° 11' 13.35"	Large	Closed	18.92 ± 0.30	18.47 ± 2.78	39
09/22/11	Santa Ynez River	34° 41' 30.57"	Large	Open	18.51 ± 0.32	16.53 ± 1.96	40
10/07/11	Arroyo Burro Lagoon	34° 24' 11.77"	Medium	Closed	24.95 ± 0.44	20.53 ± 1.53	40
09/25/11	Santa Clara River	34° 14' 07.19"	Large	Closed	21.47 ± 0.21	13.68 ± 0.95	38
10/28/11	Las Flores Marsh	33° 17' 25.79"	Large	Closed	25.11 ± 0.18	18.58 ± 1.48	39
Arrow goby							
09/17/11	Arcata Bay	40° 51' 30.57"	Large	Open	20.41 ± 0.10	13.85 ± 1.33	41
09/02/11	Bodega Bay	38° 18' 59.42"	Large	Open	16.12 ± 0.33	14.33 ± 1.49	42
09/10/11	San Lorenzo River	36° 57' 56.41"	Large	Open	17.62 ± 0.12	16.13 ± 1.84	40
09/09/11	Elkhorn Slough	36° 48' 40.14"	Large	Open	18.37 ± 0.18	13.84 ± 2.22	41
08/25/11	Morro Bay	35° 20' 34.52"	Large	Open	17.96 ± 0.07	14.50 ± 2.02	40
08/24/11	Carpinteria Salt Marsh	34° 23' 52.97"	Medium	Open	19.70 ± 0.12	13.10 ± 1.96	43
08/28/11; 10/31/11	Colorado Lagoon	33° 45' 10.52"	Large	Open	20.50 ± 1.00	18.01 ± 3.01	29
08/27/11	Los Peñasquitos	32° 55' 57.84"	Large	Open	21.37 ± 0.34	13.21 ± 2.09	41

* (USFWS 2005) Habitat size classification: large = systems of >2 hectares (5 acres) in surface area; medium = systems with a surface area <2 ha but >0.4 ha (1 acre); small = systems with surface area < 0.4 ha

1954). This size mesh likely selected for individuals on the larger end of our target size range, and smaller gobies were seen passing through the net at times. We used this mesh because it was large enough to allow algae and other material to pass through the net while still capturing fish spanning the targeted size range. Although our nets were size selective, which could bias estimates of density and assemblage composition (Weinstein & Davis 1980), our tests for correlations between larval traits and temperature or latitude are unlikely to be similarly biased because the same mesh size was used at all study sites. Once the fishes were collected, they were euthanized and then preserved in 95% ethanol. This preservation technique is considered the best choice when using otoliths to assess life history traits, despite the possibility that specimens can shrink over time (Fowler & Smith 1983).

The standard length of each goby was measured to the nearest 0.01 mm using a digital caliper before extraction of lapillar otoliths. Lapilli were used for all otolith analyses because they were clearer and required less processing than sagittal otoliths. Previous work has validated daily increment deposition in sagittal otoliths for the tidewater goby (Hellmair 2010), and we found no differences in increment counts between sagittal and lapillar otoliths based on

a small subsample in this study. Due to the similarity and close phylogenetic relationship with the tidewater goby (Ellingson et al. 2014), it was assumed that increments in otoliths of the arrow goby were also deposited daily. Furthermore, otolith-based estimates of larval traits appear to be suitable for both species due to a strong relationship between body length and age (Spies et al. 2014).

A total of 712 otoliths (arrow goby: n = 317; tidewater goby: n = 395) were prepared using techniques similar to Samhuri et al. (2009), which included placing the otoliths in immersion oil for >30 d to clear them to facilitate interpretation. Otoliths were then viewed under a compound microscope with a polarizing filter and camera at 200× magnification. Any otoliths that were irregularly shaped or unreadable were discarded from the study (tidewater goby n = 5; arrow goby n = 8). The remaining otoliths were viewed and measured twice by the same individual (B. T. Spies). Following methods from Wilson et al. (2009), if the 2 readings were >10% different, they were not included in any analysis. If the 2 readings were <10% different, but not the same, then the second reading (which we considered more accurate) was used for the analysis.

Increment measurements were made along the longest axis, from the core to the outermost complete



Fig. 2. (A) Moore Creek, located in Santa Cruz County, represents a seasonally closed estuary inhabited by the tidewater goby. (B) Carpinteria Salt Marsh, located in Santa Barbara County, represents a fully tidal system inhabited by the arrow goby

ring, using Image-Pro Plus image analysis software. The settlement band was interpreted as the point where the daily increment transitioned distinctly in band width (Wilson & McCormick 1999, Spies et al. 2014). Date of settlement was found by subtracting the number of post-settlement increments counted for each otolith from the date the fish was collected. Larval duration was estimated as the number of rings from the hatch mark to the settlement mark. The first dark ring outside the core was considered to be the hatch mark (Hellmair 2010), although this interpretation has not been validated. Average pre-settlement growth rates (larval growth rate) were estimated from otolith growth as a proxy for larval somatic growth, by dividing the otolith radius to the settlement mark by the age at settlement (McCormick & Molony 1995, Spies et al. 2014).

Temperature measurement

Hourly temperature measurements were recorded at all 18 sites, from 10 July to 3 October 2011, allowing for at least 6 wk of temperature data to be recorded before fish collections began on 24 August 2011. At 17 of the 18 sites, iButton thermocron temperature recorders (model DS1921G with 0.5°C resolution) were used. Before placement in the field, 10 iButtons were randomly chosen to test for accuracy and precision in controlled laboratory conditions. All hourly temperature measurements were within the stated 0.5°C resolution range. iButtons were then waterproofed using ¾ inch (1.9 cm) schedule 40 PVC male and female caps with silicone thread tape and a small dehydration packet inserted to absorb moisture.

Two iButton recorders were placed within each site and spaced approximately 100 m apart to provide representative measurements of temperature ranges around the area where goby collections would occur. The site where iButtons were not used (Elkhorn Slough) had multiple, fixed, long-term temperature loggers already put in place and maintained by the Elkhorn Slough National Estuarine Research Reserve. At the time of fish collections, water temperature was also measured, using a YSI 556 water quality meter, and these data were compared to the temperature measurements of the fixed loggers to assess their accuracy. In addition, 2 other sites (Arcata Bay and Morro Bay) also had fixed, long-term temperature loggers in place during this study. Temperature measurements from the fixed loggers and iButton recorders were very similar (within 1.0°C). The fixed loggers were located much closer to where fish collections took place and had a much finer resolution; therefore, we used temperature measurements from them instead of iButton data recorders at those 2 sites. Those data were provided by the US Fish and Wildlife Service (Arcata Bay) and the San Luis Obispo Science and Ecosystem Alliance (Morro Bay).

Hourly temperature recordings were used to calculate the mean temperature experienced by each fish through its larval phase. The date of hatching and the date of settlement for each fish were estimated from otolith-derived age estimates and the date of capture. This allowed us to calculate the mean temperature history for each individual fish during its larval phase (McCormick & Molony 1995). The mean temperature experienced by each fish was then used to calculate the mean temperature for each study site. Additionally, temperature variance was calculated for each site as a possible additional temperature metric for analysis. However, preliminary analysis showed that temperature variance was not a good predictor of differences in larval traits, so it was not used in the analyses reported.

Statistical analysis

To test the effects of temperature and latitude on 3 larval traits (larval duration, otolith based size at settlement, and growth rate) and whether these effects differed between the 2 species (i.e. test whether slopes differed between the 2 species), a general linear model (GLM) was used in SYSTAT 13. The model included Species as a categorical factor and Temperature and Latitude as continuous variables. Whether

the effects of temperature or latitude were consistent between species was tested by the interaction terms between these 2 covariates and the factor Species. To allow unbiased tests of the main effects of Species, Temperature, and Latitude, all non-significant interaction terms involving the continuous variables (covariates) were sequentially eliminated if $p > 0.05$, following Winer et al. (1991).

In addition to the GLM, linear regression was used to test the fit of larval trait data for the 2 species to temperature or latitude. This was done, in addition to GLM models mentioned above, to evaluate whether the extent to which temperature or latitude predicted variation in larval traits differed between the 2 species. The GLM analyses tested whether 2 slopes were equal (via the interaction term), and if the slopes were equal, whether the pooled slope differed from zero, but those analyses did not test whether the fits to those 2 lines were equal. These linear regressions were used only to generate r^2 -values, not p-values (because doing so would increase the probability of Type I error by testing the same hypothesis — that the slope was equal to zero — with the same data set twice). Linear regression was also used to examine the relationship between temperature and latitude of the 18 study sites. For all analyses, assumptions of normality and homogeneity of variance were checked by inspection of normal probability plots and residual plots.

The 2 most northern sites (Arcata Bay and Ten Mile River) were unusual, and all analyses were conducted with and without them (i.e. treating them as outliers) to evaluate their influence on the results. Thus, all GLM and linear regressions were analyzed under 2 alternative conditions: (1) a full model with all study sites ($n = 18$) and (2) a reduced model with both unusual sites removed ($n = 16$). Ten Mile River is typically a seasonally closed estuary that is inhabited by the tidewater goby, but it remained open to tidal influence throughout the duration of this project, while maintaining an abundant population of tidewater gobies. This unexpected occurrence could bias results for the tidewater goby since this estuary was more similar to the fully tidal sites occupied by arrow gobies. Arcata Bay, a site occupied by the arrow goby, was a major outlier in the relationship between temperature and latitude because it was much warmer than predicted. We are confident that our estimate of temperature is accurate because both fixed data loggers and iButtons measured essentially the same temperature. This site is shallow and has extensive sun-exposed mudflats, which probably accounts for its warm temperature.

RESULTS

Mean site temperature and latitude

Larval tidewater gobies experienced a greater range of average temperatures at the study sites than did larvae of the arrow goby. Average temperatures at the sites occupied by the tidewater goby differed by $\sim 10^{\circ}\text{C}$ from the coolest to warmest site (range: 15.33 ± 0.14 to 25.11 ± 0.18 , $n = 10$), whereas those occupied by the arrow goby differed by only $\sim 5^{\circ}\text{C}$ (range: 16.12 ± 0.33 to 21.27 ± 0.34 , $n = 8$). Removal of the 2 unusual sites slightly reduced the temperature range to 7.7°C for tidewater goby sites but did not change the temperature range of arrow goby sites.

The study sites showed the expected trend of lower temperature at higher latitude, but when all sites were included in the regression models, this trend was only statistically significant for estuaries inhabited by the tidewater goby (Table 1, Fig. 3: tidewater goby $n = 10$, $r^2 = 0.603$, $p = 0.008$; arrow goby $n = 8$, $r^2 = 0.167$, $p = 0.315$). The removal of the 2 unusual sites, however, strengthened the temperature-latitude relationship such that it was significant for estuaries inhabited by both species (Table 1, Fig. 3: tidewater goby: $n = 9$, $r^2 = 0.465$, $p = 0.043$; arrow goby: $n = 7$, $r^2 = 0.936$, $p < 0.001$).

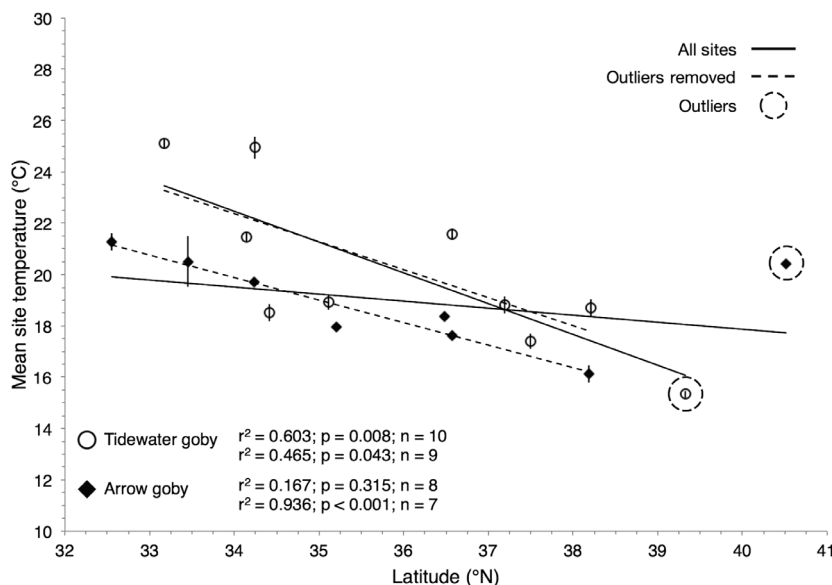


Fig. 3. Temperatures for each site ($^{\circ}\text{C}$, mean hourly ± 1 SD) as a function of latitude. Solid trend lines represent relationships for all 18 study sites inhabited by the tidewater goby and arrow goby ($n = 10$ and 8 sites, respectively). Dashed trend lines represent relationships for 16 study sites with the 2 outlier sites removed ($n = 9$ and 7 sites, respectively)

Effects of temperature and latitude on larval traits

Larval traits were significantly related to temperature, but not latitude, and these relationships between larval traits and temperature were stronger in the tidewater goby than the arrow goby. This variation between species was likely due to the larger range of temperatures in estuaries inhabited by the tidewater goby (10 or 7.7°C with an outlier removed), compared to those inhabited by the arrow goby (5°C). There were no significant Species \times Temperature or Species \times Latitude interactions found in the full model (Species \times Temperature: larval duration $F_{1,10} = 0.25$, $p = 0.63$; size at settlement $F_{1,10} = 2.75$, $p = 0.13$; growth rate $F_{1,10} = 0.73$, $p = 0.41$; Species \times Latitude: larval duration $F_{1,10} = 0.36$, $p = 0.56$; size at settlement $F_{1,10} = 2.99$, $p = 0.11$; growth rate $F_{1,10} = 0.61$, $p = 0.45$; Figs. 4 & 5) or in the model with outliers removed (Species \times Temperature: larval duration $F_{1,8} = 2.46$, $p = 0.16$; size at settlement $F_{1,8} = 0.92$, $p = 0.37$; growth rate $F_{1,8} = 0.48$, $p = 0.51$; Species \times Latitude: larval duration $F_{1,8} = 3.43$, $p = 0.10$; size at settlement $F_{1,8} = 1.28$, $p = 0.29$; growth rate $F_{1,8} = 0.68$, $p = 0.44$; Figs. 4 & 5). Thus, there was no statistical evidence for a difference in the pattern of response of the 2 species to the effects of temperature and latitude. All 3 larval traits were significantly related to temperature (Table 2). Larval duration decreased as temperature increased (full model: $p = 0.001$; outliers removed: $p = 0.01$; Fig. 4A), as did size at settlement ($p = 0.03$ and $p = 0.056$; Fig. 4B), whereas larval growth rates increased with temperature (full model: $p = 0.04$; outliers removed: $p = 0.05$; Fig. 4C). However, latitude was not related to the 3 larval traits: larval duration ($p = 0.76$ and $p = 0.95$ for full and reduced models, respectively; Fig. 5A), settlement at size ($p = 0.15$ and $p = 0.92$; Fig. 5B), or growth rate ($p = 0.07$ and $p = 0.71$; Fig. 5C).

Linear regressions evaluating the fit of the larval traits to temperature and latitude including all study sites revealed that traits of the tidewater goby were more predictable based on temperature or latitude than were those of the arrow goby. Strong negative relationships with temperature predicted 73% and 71% of the variation in larval duration and settlement size, respectively, for the tidewater goby but only 40% and 8% in the arrow goby (Fig. 4). Positive linear rela-

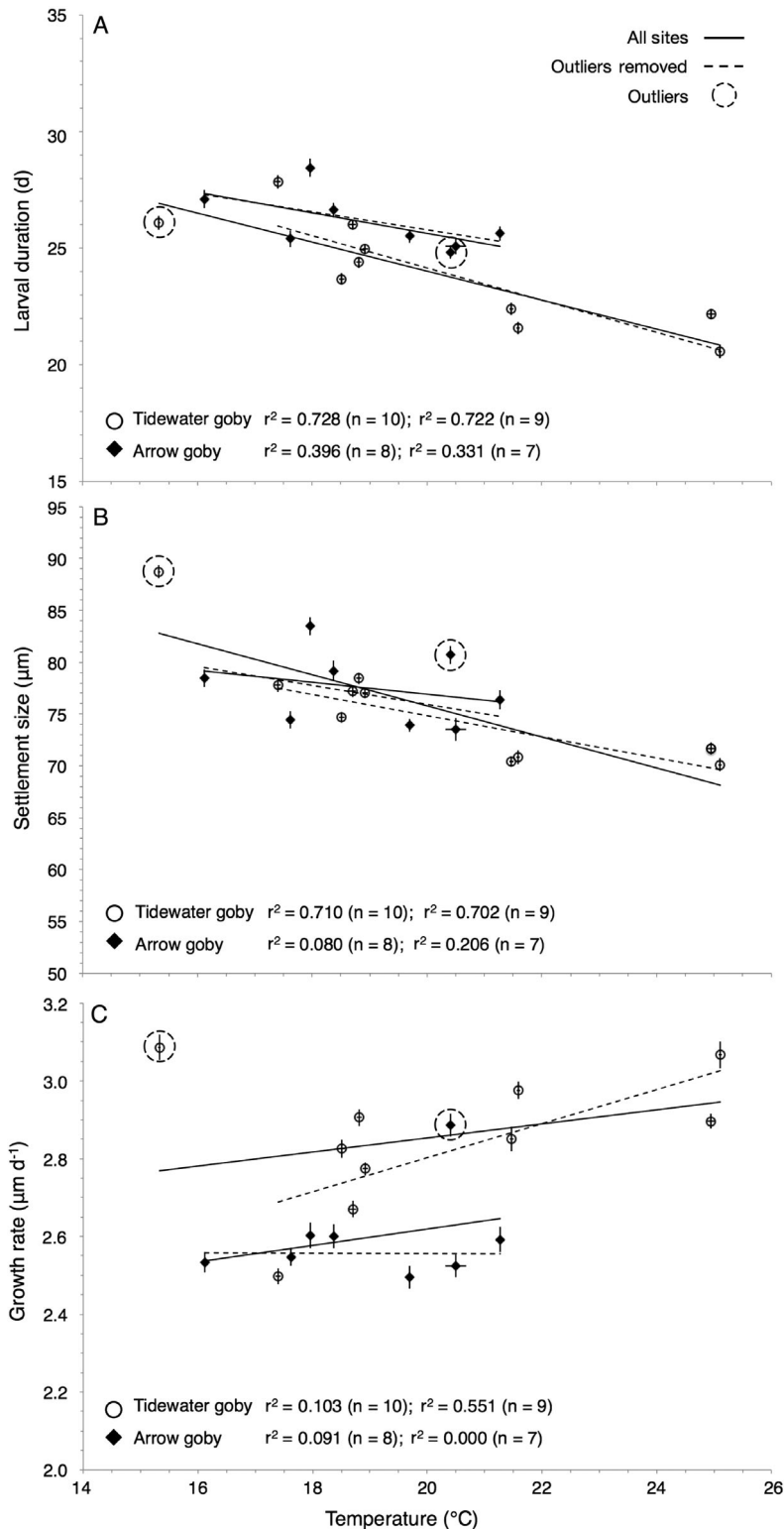


Fig. 4. Relationships between site temperature ($^{\circ}\text{C}$, mean hourly ± 1 SD) and (A) larval duration, (B) size at settlement, and (C) and pre-settlement growth rate for the tidewater goby and arrow goby in all 18 study sites (solid lines: n = 10 and 8 sites, respectively) and with outlier sites removed (dashed lines: n = 9 and 7 sites, respectively). Error bars for all 3 response variables represent ± 1 SE

tionships with latitude predicted a somewhat smaller fraction of the variation in larval duration and settlement size, 56% and 64%, respectively, for the tidewater goby, and <1% and 23% for the arrow goby (Fig. 5). The better fits of the relationships with temperature rather than latitude suggest that temperature was a stronger driver of larval duration and settlement size. Also, these findings suggest that the effects of temperature on larval duration and settlement size found in the GLM analyses were driven mostly by the tidewater goby, which experienced a greater range of temperatures than the arrow goby.

Larval growth rate was not strongly predicted by temperature in either species: tidewater goby ($r^2 = 0.10$; Fig. 4C) or arrow goby ($r^2 = 0.09$). However, the removal of an outlier site significantly improved this relationship for the tidewater goby ($r^2 = 0.55$; Fig. 4C). Furthermore, larval growth rate had a moderately strong positive relationship with latitude for the arrow goby ($r^2 = 0.46$; Fig. 5C) but a weak negative relationship for the tidewater goby ($r^2 = 0.03$). When the 2 outlier sites were removed from the analyses, there was no relationship between growth rate and latitude for the arrow goby ($r^2 = 0.00$), but the strength of the negative relationship increased for the tidewater goby ($r^2 = 0.36$).

Overall, considering all 18 study sites, the tidewater goby had a larval duration that was approximately 35% shorter in the warmest sites, as well being 27% smaller in body size and growing 24% faster in those sites. In comparison, the arrow goby had an approximately 15% shorter larval duration in the warmest sites, was 14% smaller in body size, and grew 16% faster.

DISCUSSION

This study demonstrates the variable nature of temperature in estuarine habitats and the influence temperature can have on the traits of larval fishes. Larval duration, settlement size, and growth rate were significantly affected by temperature,

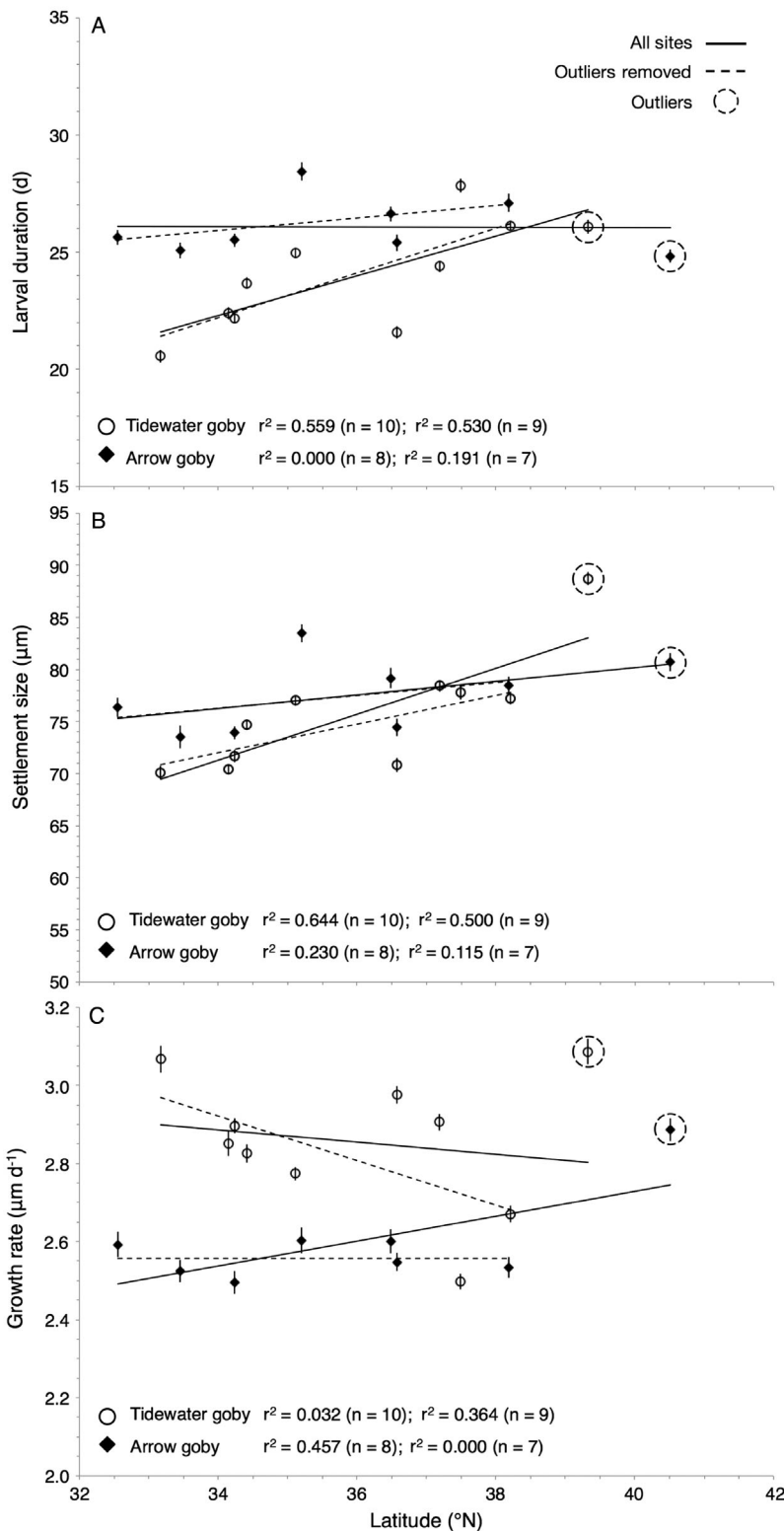


Fig. 5. Relationships between latitude and (A) larval duration, (B) size at settlement, and (C) and pre-settlement growth rate for the tidewater goby and arrow goby in all 18 study sites (solid lines: n = 10 and 8 sites, respectively) and with outlier sites removed (dashed lines: n = 9 and 7 sites, respectively). Error bars represent ± 1 SE

with no statistically significant effects of latitude. Temperature, however, had a stronger effect on larvae of the tidewater goby than the arrow goby, presumably due to the larger difference among sites in temperatures this species experienced. Nonetheless, in warmer estuarine environments, both the arrow goby and tidewater goby spent fewer days in the larval phase, were smaller in size at settlement, and exhibited faster growth rates.

The smaller, seasonally closed estuaries inhabited by the tidewater goby exhibited a greater range of mean temperature (10°C) than the larger, tidal estuaries inhabited by the arrow goby (5°C). Excluding the unusually warm Arcata Bay, temperature was also better predicted by latitude in the tidal estuaries inhabited by arrow gobies than in the seasonally closed estuaries inhabited by tidewater gobies. However, in testing the relationships between larval traits and temperature, including Arcata Bay was useful because doing so helped to remove the covariance between temperature and latitude. Models with and without the 2 unusual sites (Arcata Bay and Ten Mile River) showed generally similar results, strengthening the conclusion that variation in all 3 larval traits was more closely linked to temperature than to other attributes that change with latitude.

A number of studies have found similar effects of temperature on larvae of a variety of temperate and tropical marine fishes, using both field and laboratory approaches. Several species of flatfish in temperate systems have exhibited shorter larval durations with increased temperature in controlled laboratory experiments (Laurence 1975, Fonds 1979, Policansky 1982, Seikai et al. 1986, Gadomski & Caddell 1991, Keefe 1993). Green & Fisher (2004) found that a decrease of 3°C resulted in a slower swimming development in laboratory reared tropical anemone fish *Amphiprion melanopus*, including longer larval duration and reduced growth rates. Similar results were found in the tropical goatfish *Upeneus tragula*, which showed a significant negative relationship between

Table 2. Results of general linear models (GLM) testing for effects of temperature, latitude, and species on larval duration, settlement size, and pre-settlement growth rate for all study sites (n = 18) and with 2 unusual sites removed (n = 16)

n	Source	df	SS	F	p	r ²
Larval duration						
18	Species	1	8.523	5.931	0.029	0.745
	Latitude	1	0.140	0.097	0.760	
	Mean temp.	1	27.195	18.923	0.001	
	Error	14	20.12			
16	Species	1	5.067	3.132	0.102	0.748
	Latitude	1	0.007	0.005	0.948	
	Mean temp.	1	13.825	8.545	0.013	
	Error	12	19.415			
Settlement size						
18	Species	1	2.979	0.271	0.611	0.603
	Latitude	1	25.267	2.300	0.152	
	Mean temp.	1	66.498	6.054	0.027	
	Error	14	153.785			
16	Species	1	3.100	0.412	0.533	0.560
	Latitude	1	0.088	0.012	0.916	
	Mean temp.	1	33.713	4.479	0.056	
	Error	12	90.332			
Growth rate						
18	Species	1	0.191	9.473	0.008	0.597
	Latitude	1	0.078	3.847	0.070	
	Mean temp.	1	0.106	5.261	0.038	
	Error	14	0.283			
16	Species	1	0.106	8.925	0.011	0.736
	Latitude	1	0.002	0.141	0.714	
	Mean temp.	1	0.055	4.650	0.052	
	Error	12	0.143			

water temperature and size and age at metamorphosis both in the field and in laboratory experiments (McCormick & Molony 1995). Furthermore, water temperature in the upper Florida Keys explained 78% of the variation in the pelagic larval duration (PLD) in the Caribbean reef fish *Thalassoma bifasciatum* (Sponaugle et al. 2006), with larval growth explaining 85% of the variation in PLD. This resulted in warmer water cohorts exhibiting a shorter PLD as a product of faster growth rates. Fish from warmer periods were larger at age, but due to the shorter time spent in the larval phase, they were smaller in body size at settlement. Larvae of the Hawaiian amphidromous goby *Lentipes concolor* settled after a shorter PLD and at smaller sizes during months of warmer ocean temperature (Radtke et al. 2001).

Although temperature was a good predictor of larval traits in this study, these traits are likely influenced by other biotic or abiotic factors (e.g. water quality, predator abundance, and food availability). For example, variations in precipitation and stream flow are 2 of the main hydrological drivers of mouth

dynamics (open or closed) in seasonally closed estuaries throughout California (Rich & Keller 2013). During the summer months when precipitation is typically low in California, which is when this study took place, a net increase in evaporation can lead to hypersaline environments (Largier et al. 1997). Salinity can influence nutrient availability, physiological rates, and behavior responses (Cowen & Sponaugle 2009). For example, unusually low or high salinity can interfere with egg development and the hatching process and reduce survival of larvae (Fonds 1979). In seasonally closed estuaries, dissolved oxygen levels can drop precipitously, especially during warm months (Collins & Melack 2014). Lower than normal levels of dissolved oxygen can lead to reduced growth rates (Boeuf et al. 1999, Pichavant et al. 2001, Ekau et al. 2010) and altered swimming behavior (Ciuhandu et al. 2007). Increased flow rates when estuary mouths are open, and the resulting turbulence, could enhance prey encounters and, thus, ingestion rates (Kjørboe & MacKenzie 1995). In addition, genetic differences among populations can contribute to latitudinal differences in larval traits, as Billerbeck et al. (2001) found for growth of the Atlantic silverside *Menidia menidia*.

Due to the permitting and logistical constraints, we were unable to examine the temporal variation in larval traits. This would be an interesting topic for future research. Temporal variability in the abiotic environment can differ strongly between fully tidal estuaries and seasonally closed estuaries, depending on their frequency and degree of closure/breaching. Temperature changes of only a few degrees Celsius, as might occur during a breaching event, have been found to strongly influence key physiological processes in fish, such as developmental rate, growth rate, swimming ability, and reproductive performance (Munday et al. 2008, 2009). Physiological responses to temperature commonly follow a dome-shaped relationship where a maximum is reached as rates increase with temperature, but responses thereafter decrease rapidly if temperatures exceed the thermal optimum (Munday et al. 2009). The thermal optima for the arrow goby and tidewater goby larvae are unknown but are likely different given what is known about the natural history of these 2 species. Such a difference could be a result of local adaptation or acclimation to site-specific temperature regimes (Hawkins 1996). However, if water temperature rises above the thermal optimum for reproduction, this could potentially cause a decline in adult reproductive output, which could result in significant consequences for population connectivity (Munday et al. 2008, 2009).

It is not clear how climate change will affect temperature regimes in closed vs. open estuarine habitats along the coast of California because variations in climate patterns over large latitudinal scales can affect the overall size, function, and distribution of estuarine habitats (Scavia et al. 2002, Day et al. 2008). A rise in global temperature is expected to alter the evaporation/precipitation regime, causing increased evaporation at lower latitudes and increased precipitation in the higher latitudes (Roesig et al. 2004). This could cause longer durations of estuary closure in southern California and more frequent opening conditions in northern California. Increased global temperatures will likely accelerate larval development for many species. This could be beneficial or harmful for an individual species depending on their local conditions and community structure. For example, if a species such as the arrow goby experienced a decrease in larval duration, this change could decrease the spatial scale over which it disperses, reducing connectivity (O'Connor et al. 2007, Munday et al. 2009). However, reductions in larval duration could potentially increase survival to settlement by reducing exposure to mortality agents in the plankton. This change could prove beneficial in habitats where there are numerous predators of larvae, but could be detrimental in habitats where increased body size at settlement gives a competitive advantage in the juvenile and adult stage (Fontes et al. 2010).

In summary, larval traits of the 2 estuarine gobies in this study differed, both between species and within species, among estuaries spanning the coast of California. Differences in larval traits between species followed the pattern proposed by Dawson et al. (2002), with the tidewater goby predicted to have a shorter larval duration than the arrow goby, based on an apparent lack of marine larval dispersal in the tidewater goby. Variation in larval traits for both species was strongly related to differences in the mean temperature experienced during larval development and not strongly related to latitude per se. Our findings also reveal the variable nature of estuarine habitats and their abiotic conditions. Closed estuaries exhibited a greater range of thermal environments than did estuaries open to the ocean. These systems, however, have not been studied in great detail. We know relatively little about latitudinal trends in the abiotic environment and even less about biotic interactions. For example, how do the types and abundance of prey for larval fishes change with latitude, and are these changes similar between open and seasonally closed estuaries? What about predators

of larvae, and densities of conspecifics? Further research on these systems is needed to better understand how local biotic and abiotic conditions can affect population dynamics of endangered taxa, as well as provide vital information on the ability of endemic and threatened species to acclimate or adapt to the projected increases in temperature due to climate change.

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LITERATURE CITED

- Barlow M (2002) Phylogeographic structure of the tidewater goby, *Eucyclogobius newberryi* (Teleostei: Gobiidae), in the San Francisco Bay area and Ventura County: implications for conservation management. MA thesis, University of California at Los Angeles, CA
- Billerbeck JM, Lankford TM, Conover DO (2001) Evolution of intrinsic growth and energy acquisition rates. I. Trade-offs with swimming performance in *Menidia menidia*. *Evolution* 55:1863–1872
- Boeuf G, Boujard D, Person-Le-Ruyet J (1999) Control of the somatic growth in turbot. *J Fish Biol* 55:128–147
- Bond MH, Hayes SA, Hanson CV, MacFarlane RB (2008) Marine survival of steelhead (*Oncorhynchus mykiss*) enhanced by a seasonally closed estuary. *Can J Fish Aquat Sci* 65:2242–2252
- Brothers EB (1975) The comparative ecology and behavior of three sympatric California gobies. PhD dissertation, University of California, San Diego, CA
- Brown JH, Gillooly JF, Allen AP, Savage VM, West GB (2004) Toward a metabolic theory of ecology. *Ecology* 85: 1771–1789
- Ciuhandu CS, Wright PJ, Goldberg JI, Stevens ED (2007) Parameters influencing the dissolved oxygen in the boundary layer of rainbow trout (*Oncorhynchus mykiss*) embryos and larvae. *J Exp Biol* 210:1435–1445
- Collins DG, Melack JM (2014) Biological and chemical responses in a temporarily open/closed estuary to variable freshwater inputs. *Hydrobiologia* 734:97–113
- Cooper SD, Lake PS, Sabater S, Melack JM, Sabo JL (2013) The effects of land use changes on streams and rivers in Mediterranean climates. *Hydrobiologia* 719:383–425
- Cousins M, Stacey MT, Drake JL (2010) Effects of seasonal

- stratification on turbulent mixing in a hypereutrophic coastal lagoon. *Limnol Oceanogr* 55:172–186
- Cowen RK (1991) Variation in the planktonic larval duration of the temperate wrasse *Semicossyphus pulcher*. *Mar Ecol Prog Ser* 69:9–15
- Cowen RK, Sponaugle S (2009) Larval dispersal and marine population connectivity. *Annu Rev Mar Sci* 1:443–466
- Dawson MN, Louie KD, Barlow M, Jacobs DK, Swift CC (2002) Comparative phylogeography of sympatric sister species, *Clevelandia ios* and *Eucyclogobius newberryi* (Teleostei, Gobiidae), across the California Transition Zone. *Mol Ecol* 11:1065–1075
- Day JW, Christian RR, Boesch DM, Yáñez-Arancibia A and others (2008) Consequences of climate change on the ecomorphology of coastal wetlands. *Estuar Coasts* 31:477–491
- Earl DA, Louie KD, Bardeleben C, Swift CC, Jacobs DK (2010) Rangelwide microsatellite phylogeography of the endangered tidewater goby, *Eucyclogobius newberryi* (Teleostei: Gobiidae), a genetically subdivided coastal fish with limited marine dispersal. *Conserv Genet* 11:103–114
- Ekau W, Amel H, Pörtner HO, Gilbert D (2010) Impacts of hypoxia on the structure and processes in pelagic communities (zooplankton, macro-invertebrates and fish). *Biogeosciences* 7:1669–1699
- Ellingson RA, Swift CC, Findley LT, Jacobs DK (2014) Convergent evolution of ecomorphological adaptations in geographically isolated Bay gobies (Teleostei: Gobionellidae) of the temperate North Pacific. *Mol Phylogenet Evol* 70:464–477
- Fonds M (1979) Laboratory observations on the influence of temperature and salinity on development of the eggs and growth of the larvae of *Solea solea* (Pisces). *Mar Ecol Prog Ser* 1:91–99
- Fontes J, Afonso P, Santos RS, Caselle JE (2010) Temporal variability of larval growth, size, stage duration and recruitment of a wrasse, *Coris julis* (Pisces: Labridae), from the Azores. *Sci Mar* 74:721–729
- Fowler GM, Smith SJ (1983) Length changes in silver hake (*Merluccius bikinearis*) larvae: effects of formalin, ethanol, and freezing. *Can J Fish Aquat Sci* 40:866–870
- Gadomski DM, Caddell SM (1991) Effects of temperature on early-life-history stages of California halibut *Paralichthys californicus*. *Fish Bull* 89:567–576
- Gillooly JF (2001) Effects of size and temperature on metabolic rate. *Science* 293:2248–2251
- Gillooly JF, Charnov EL, West GB, Savage VM, Brown JH (2002) Effects of size and temperature on developmental time. *Nature* 417:70–73
- Green BS, Fisher R (2004) Temperature influences swimming speed, growth and larval duration in coral reef fish larvae. *J Exp Mar Biol Ecol* 299:115–132
- Hawkins AJS (1996) Temperature adaptation and genetic polymorphism in aquatic animals. In: Johnston IA, Bennett AF (eds) *Animals and temperature: phenotypic and evolutionary adaptation*. Cambridge University Press, Cambridge, p 103–126
- Hayhoe K, Cayan D, Field CB, Frumhoff PC and others (2004) Emissions pathways, climate change, and impacts on California. *Proc Natl Acad Sci USA* 101:12422–12427
- Hellmair M (2010) Manual for otolith based age determination of tidewater goby, *Eucyclogobius newberryi*. USFWS Scientific Report
- Houde ED (1989) Comparative growth, mortality, and energetics of marine fish larvae: temperature and implied latitudinal effects. *Fish Bull* 87:471–495
- Jacobs DK, Stein ED, Longcore T (2011) Classification of California estuaries based on natural closure patterns: templates for restoration and management. Southern California Coastal Water Research Project. Technical Report 619.a
- Keefe M (1993) Patterns of metamorphosis in summer flounder, *Paralichthys dentatus*. *J Fish Biol* 42:713–728
- Kjørboe T, MacKenzie B (1995) Turbulence-enhanced prey encounter rates in larval fish: effects of spatial scale, larval behavior and size. *J Plankton Res* 17:2319–2331
- Klausmeyer KR, Shaw MR (2009) Climate change, habitat loss, protected areas and the climate adaptation potential of species in Mediterranean ecosystems worldwide. *PLoS ONE* 4:e6392
- Lafferty KD, Swift CC, Ambrose RF (1999a) Postflood persistence and recolonization of endangered tidewater goby populations. *N Am J Fish Manag* 19:618–622
- Lafferty KD, Swift CC, Ambrose RF (1999b) Extirpation and recolonization in a metapopulation of an endangered fish, the tidewater goby. *Conserv Biol* 13:1447–1453
- Largier JL, Hollibaugh JT, Smith SV (1997) Seasonally hypersaline estuaries in Mediterranean-climate regions. *Estuar Coast Shelf Sci* 45:789–797
- Laurence GC (1975) Laboratory growth and metabolism of the winter flounder *Pseudopleuronectes americanus* from hatching through metamorphosis at three temperatures. *Mar Biol* 32:223–229
- McCormick MI, Hoey AS (2004) Larval growth history determines juvenile growth and survival in a tropical marine fish. *Oikos* 106:225–242
- McCormick MI, Molony BW (1995) Influence of water temperature during the larval stage on size, age and body condition of a tropical reef fish at settlement. *Mar Ecol Prog Ser* 118:59–68
- Meekan MG, Carleton JH, McKinnon AD, Flynn K, Furnas M (2003) What determines the growth of tropical reef fish larvae in the plankton: food or temperature? *Mar Ecol Prog Ser* 256:193–204
- Miller DJ, Lea RN (1972) Guide to the coastal marine fishes of California. California Department of Fish and Game. *Fish Bull* 157:1–235
- Mills LS, Allendorf FW (1996) The one-migrant-per-generation rule in conservation and management. *Conserv Biol* 10:1509–1518
- Moser GH (1996) The early stages of fishes in the California Current region. California Cooperative Oceanic Fisheries Investigation Atlas no.33, NOAA/NMFS-SFSC, Allen Press, Lawrence, KS
- Moyle PB (2002) *Inland fishes of California*. Revised and expanded. University of California Press, Berkeley, CA
- Munday PL, Jones GP, Pratchett MS, Williams AJ (2008) Climate change and the future for coral reef fishes. *Fish Fish* 9:261–285
- Munday PL, Leis JM, Lough JM, Paris CB, Kingsford MJ, Berumen ML, Lambrechts J (2009) Climate change and coral reef connectivity. *Coral Reefs* 28:379–395
- O'Connor MI, Bruno JF, Gaines SD, Halpern BS, Lester SE, Kinlan BP, Weiss JM (2007) Temperature control of larval dispersal and the implications for marine ecology, evolution, and conservation. *Proc Natl Acad Sci USA* 104:1266–1271
- Pankhurst NW, Munday PL (2011) Effects of climate change on fish reproduction and early life history stages. *Mar*

- Freshw Res 62:1015–1026
- Pichavant K, Person-Le-Ruyet J, Le Bayon N, Severe A, Le Roux A, Boeuf G (2001) Comparative effects of long-term hypoxia on growth, feeding and oxygen consumption in juvenile turbot and European sea bass. *J Fish Biol* 59: 875–883
 - Policansky D (1982) Influence of age, size, and temperature on metamorphosis in the starry flounder, *Platichthys stellatus*. *Can J Fish Aquat Sci* 39:514–517
 - Prasad RR (1959) Reproduction in *Clevelandia ios* (Jordan and Gilbert), with an account of the embryonic and larval development. *Proc Nat Inst Sci India B Biol Sci* 25B: 12–30
 - Radtke RL, Kinzie RA, Shafer DJ (2001) Temporal and spatial variation in length of larval life and size at settlement of the Hawaiian amphidromous goby *Lentipes concolor*. *J Fish Biol* 59:928–938
 - Rich A, Keller EA (2013) A hydrologic and geomorphic model of estuary breaching and closure. *Geomorphology* 191:64–74
 - Roessig JM, Woodley CM, Cech JJ Jr, Hansen LJ (2004) Effects of global climate change on marine and estuarine fishes and fisheries. *Rev Fish Biol Fish* 14:251–275
 - Rutherford ES, Houde ED (1995) The influence of temperature on cohort-specific growth, survival and recruitment of striped bass, *Morone saxatilis*, larvae in Chesapeake Bay. *Fish Bull* 93:315–332
 - Samhuri JF, Steele MA, Forrester GE (2009) Inter-cohort competition drives density dependence and selective mortality in a marine fish. *Ecology* 90:1009–1020
 - Scavia D, Field JC, Boesch DF, Buddemeier RW and others (2002) Climate change impacts on US coastal and marine ecosystems. *Estuaries* 25:149–164
 - Seikai T, Tanangonan JB, Tanaka M (1986) Temperature influence on larval growth and metamorphosis of the Japanese flounder *Palulichthys olivaceus* in the laboratory. *Bull Jpn Soc Sci Fish* 52:977–982
 - Spies BT, Tarango BC, Steele MA (2014) Larval duration, settlement, and larval growth rates of the endangered tidewater goby (*Eucyclogobius newberryi*) and the arrow goby (*Clevelandia ios*) (Pisces, Teleostei). *Bull South Calif Acad Sci* 113:165–175
 - Sponaugle S, Grorud-Colvert K, Pinkard D (2006) Temperature-mediated variation in early life history traits and recruitment success of the coral reef fish *Thalassoma bifasciatum* in the Florida Keys. *Mar Ecol Prog Ser* 308: 1–15
 - Steele MA, Schroeter SC, Page HM (2006) Sampling characteristics and biases of enclosure traps for sampling fishes in estuaries. *Estuaries Coasts* 29:630–638
 - Strawn K (1954) The pushnet, a one-man net for collecting in attached vegetation. *Copeia* 1954:195–197
 - Swenson RO (1999) The ecology, behavior, and conservation of the tidewater goby, *Eucyclogobius newberryi*. *Environ Biol Fishes* 55:99–119
 - Swift CC, Nelson JL, Maslow C, Stein T (1989) Biology and distribution of the tidewater goby, *Eucyclogobius newberryi* (Pisces: Gobiidae) of California. *Natural History Museum of Los Angeles County. Contrib Sci* 404: 1–19
 - Swift CC, Haglund TR, Ruiz M, Fisher RN (1993) The status and distribution of the freshwater fishes of Southern California. *Bull South Calif Acad Sci* 92:101–167
 - Takahashi M, McCormick MI, Munday PL, Jones GP (2012) Influence of seasonal and latitudinal temperature variation on early life-history traits of a coral reef fish. *Mar Freshw Res* 63:856–864
 - USFWS (US Fish and Wildlife Service) (2005) Recovery plan for the tidewater goby (*Eucyclogobius newberryi*). US Fish and Wildlife Service, Portland, OR
 - Valiela I, Kinney E, Bulbertson J, Peacock E, Smith S (2009) Global losses of mangroves and salt marshes. In: CM Duarte (ed) *Global loss of coastal habitats: rates, causes and consequences*. Fundación BBVA, Madrid, p 107–142
 - Wang J (2004) Application of the one-migrant-per-generation rule to conservation and management. *Conserv Biol* 18:332–343
 - Weinstein MP, Davis RW (1980) Collection efficiency of seine and rotenone samples from tidal creeks, Cape Fear River, North Carolina. *Estuaries* 3:98–105
 - Wilson DT, McCormick MI (1999) Microstructure of settlement-marks in the otoliths of tropical reef fishes. *Mar Biol* 134:29–41
 - Wilson DT, Meekan MG (2002) Growth-related advantages for survival to the point of replenishment in the coral reef fish *Stegastes partitus* (Pomacentridae). *Mar Ecol Prog Ser* 231:247–260
 - Wilson JA, Vigliola L, Meekan MG (2009) The back-calculation of size and growth from otoliths: Validation and comparison of models at an individual level. *J Exp Mar Biol Ecol* 368:9–21
 - Winer BJ, Brown DR, Michels KM (1991) *Statistical principles in experimental design*. McGraw-Hill, New York, NY
 - Wright S (1931) Evolution in Mendelian populations. *Genetics* 16:97–159

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