

# Expression analysis and identification of antimicrobial peptide transcripts from six North American frog species

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*Diseases of Aquatic Organisms* 104:225–236 (2013)

**Supplement 1.** Supplementary figures show the location of frog capture sites (Fig. S1), primers used to sequence putative antimicrobial peptide sequences (Fig. S2), alignment of predicted antimicrobial peptides identified in this study (Fig. S3), and the mRNA abundance of antimicrobial peptide precursor temporin-1BYa in *Rana boylei*, by sex (Fig. S4)

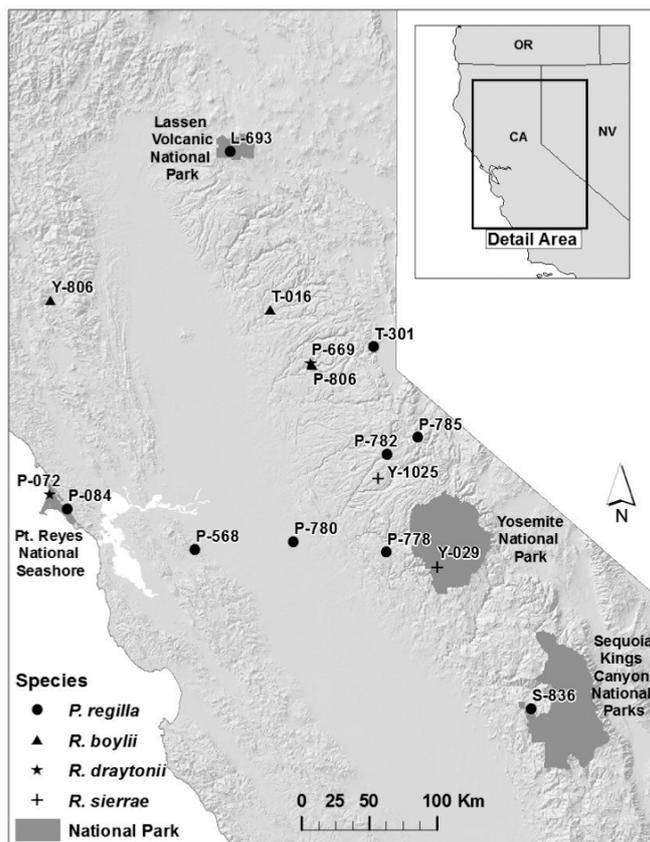


Fig. S1. Sites where *Pseudacris regilla*, *Rana boylei*, *R. draytonii*, and *R. sierrae* were captured to collect skin secretions

A:

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LRO-335a
LRO-209
LRO-208
1 GCT TTG TAG GAT AGA CCT GCA CTG AAG TCT TCC AGC CGT CTA CAT TCT GAG CAC CAA CTG AAC TAC CCG AGC CCA AAG ATG TTC ACC TTG 90
M F T L 4

LRO-335a AAG AAA TCC CTT TTA C
LRO-213
LRO-212
LRO-210
91 AAG AAA TCC CTG TTA CTC CTC TTT TTC CTT GGG ACC ATC AAC TTA TCT CTC TGT GAG GAA GAG AGA AAT GCA GAA GAA GAA AGA AGA GAT 180
5 K K S L L L L F F L G T I N L S L C E E E E R N A E E E R R D 34

181 GAA CCA GAT GAA AGG GAT GTT CAA GTG GAA AAA CGA CTT TCA CCA AAC CTG CTC AAG AGC TTG TTG GGA AAA TAA CCA AAA ATG TTA AGA 270
35 E P D E R D V Q V E K R L S P N L L K S L L G K * 58

LRO-214b GAA TTG GAA RTC ATC TGA TGT G
LRO-216b GA ATA TCA TTT AGC TAA ATG CTA AAT G
271 ATG GAA TTG GAA ATC ATC TGA TGT GGA ATA TCA TTT AGC TAA ATG CGC AAC AGA TGT CTT ATT TAA AAA ATA AAT ATG TTG CAT
354

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B:

<u>Forward primers:</u>		<u>Reverse primers:</u>	
LRO-208	ACCAACTGAAC <u>Y</u> ACCCGAGC	LRO-214	<u>C</u> CATCAGATGA <u>Y</u> TTCCAATTC
LRO-333	ACCAACTGAAC <u>T</u> ACCTGAGC	LRO-215	<u>C</u> CATCADATG <u>R</u> YTTCCAAT <u>T</u> B
LRO-525	ACCAACTGAAC <u>T</u> ACCCGAGC	LRO-330	<u>C</u> CATCAGATG <u>A</u> CTTCCAATTC
LRO-209	GAGCCCAAAGATGTT <u>C</u> ACC <u>W</u> TG	LRO-331	<u>C</u> CATCAGATG <u>A</u> TTTCCAATTC
LRO-526	GAGCCCAAAGATGTT <u>C</u> ACC <u>T</u> TG	LRO-332	<u>T</u> CATCAGATG <u>A</u> TTTCCAATTC
LRO-210	<u>C</u> CTTGGGACCAT <u>C</u> A <u>A</u> CTTATC	LRO-216	<u>C</u> AT <u>T</u> TAGC <u>A</u> TTTAGCTAAATGATATTC
LRO-211	<u>T</u> CTTGGGACCAT <u>T</u> CCTTATC	LRO-217	<u>T</u> GR <u>T</u> GTGCATTTAGCTAAATGATATTC
LRO-212	CATTTCCTTATCTCTCTGTGAAC	Antisense primer	GACAT <u>C</u> TGG <u>T</u> GTGC <u>A</u> ATTAGCT
LRO-213	GGAAGAGARATGCHGATG		
LRO-334	GTATGTTACCTTGAA <u>T</u> AAAT <u>T</u> CC		
LRO-335	ATGTTACCTTGAA <u>G</u> AAAT <u>C</u> CCCTTTAC		
Sense primer	ATGT_CACCT_GAAG <u>A</u> AAAT <u>C</u> CCCTC		

Fig. S2. Primers used to sequence antimicrobial peptide genes from amphibian skin secretions. (A) The locations of several primers are indicated relative to the putative protein coding sequence for *Rana temporaria* mRNA for temporin H precursor (GenBank accession number Y09394). (B) The sequences (5' to 3') for all primers used to identify antimicrobial peptide genes are listed, grouped according to location relative to the putative protein coding sequence. Nucleotides that differ within each group of primers are underlined in **bold**. The alignment of 1 primer from each group is shown in (A)

<sup>a</sup>Primer sequence is divided between 2 lines

<sup>b</sup>Reverse complement of primer sequence is shown

	10	20	30	40	50	60	70	80
<b>brevinin-1Pb</b>	MFTLNKFLLL	LFPLGTINLS	LCEEERNAE	EERRDEPDET	DVEVEKRFLP	IIAGIAAKVF	PKIFCAISKK	C
<b>brevinin-1BY</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDADE	EERRDDPDET	NVEVEKRFLP	ILASLAAKFG	PKLFLCLVTKK	C
<b>brevinin-1DR</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDADE	EERRDDPDES	NVEVEKRFLP	ILAGLATKIV	PKVFCLITKK	C
<b>esculentin-1Pa</b>	MFTLKKSLLL	IVLLGIIISLS	LCEQERNADE	DEESEIKRGI	FPKIIGKGIK	TGIVNGIKSL	VKGVGMKVPK	AGLNNIGNTG CNED-EC
<b>esculentin-1Pb</b>	MFTLKKSLLL	IVLLGIIISLS	LCEQERNADE	DEESEIKRGI	FPKIIGKGIK	TGIVNGIKNL	VKGVGMKVPK	AGLSNIGNTG CNED-EC
<b>esculentin-1Pc</b>	MFTLKKSLLL	IVLLGIIISLS	LCEQERNADE	DEESETKRGL	FTKINKKKAK	TGVFNIIKTI	GKEAGMDVIR	AGIDTIS--- CKIKGEC
<b>esculentin-2BY</b>	MFTLKKSLLL	LFPLGTISLS	LCEQERDADE	EDGEKEVKRS	IFSLLTAGAK	LLGKTLFKMA	GKAGAHLAC	KATNQC
<b>esculentin-2DR</b>	MFTLKKSLLL	LFPLGTISLS	LCEQERDADE	EDGEKEVKRG	ILSLITGAK	LLGKTLFKMA	GKAGAHLAC	KATNQC
<b>ranacyclin-P</b>	MFTLKKSLLL	LFPLGTISLS	LCEQERDSD	DDQGEVTEQV	VKRLVRCWT	KSYPKPCFV	RG	
<b>ranacyclin-1BY</b>	MFTLKKSLLL	LFPLGTISLS	LCEEERDADE	DDQGEVTEEV	VKRVLKGWT	KSYPKPCFG	KR	
<b>ranalexin-1C</b>	-----	---LGTINLS	LCEEERNAE	EERRNDPDER	VEVEKRFLGG	LKAPFALIC	AVTKK	
<b>ranatuerin-2PRa</b>	MFTMKKSLLL	FFFLGTISLS	LCEEERGADE	DDGVELTEEE	VKRGLLSSFK	GVAKGVAKDL	AGKLEKLC	KITG-C
<b>ranatuerin-2PRb</b>	MFTLEKSLLL	FFFLGTISLS	LCEEERGADE	DDVEMTEEE	VKGGIMDSVK	---GVAKNL	AAKLEKLC	KITG-C
<b>ranatuerin-2PRc</b>	MFTMKKSLLL	FFFLGTISLS	LCEEERDADD	DQG-EVVKKE	VKRAFFTTVK	---NLVTNV	AGTVIDMKK	KLTGQC
<b>ranatuerin-2BYa</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDAGD	DQG-EVVKQE	VKRAFFTFK	---NLVTNV	AGTVIDMKK	KLTGEC
<b>ranatuerin-2BYb</b>	MFTLKKSLLL	FFFLGTISLS	LCEEERGADE	DDGVELTEEE	VKRGILSTFK	GLAKGVAKDL	AGKLLDKFK	KITG-C
<b>ranatuerin-2DR</b>	MFTLKKSLLL	FFFLGTISLS	LCEEERGADE	DDGVELTEEE	VKRGIMDTFK	GIAKGVAKDL	AGKLLDELK	KMTG-C
<b>ranatuerin-2SRa</b>	MFTMKKSLLL	FFFLGTISLS	LCEEERGADE	DDGVELTEEE	VKRGLLSSFK	GVAKGVAKDL	AGKLEKLC	KITG-C
<b>ranatuerin-2SRb</b>	MFTLKKSLLL	FFFLGTISLS	LCEEERGADE	DDVEMTEEE	VKRGIMDSVK	---GVAKNL	AAKLEKLC	KITG-C
<b>temporin-1C</b>	-----	---LGTINLS	LCEEERDSDQ	EERRDDPGER	NVEVEKR-FL	PLFASLIGKL	LG	
<b>temporin-1Pb</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERNADE	EERRDDPEM	NVEVEKR-FL	PLVGKILSGL	IGK	
<b>temporin-1PR</b>	MFTMKKSLLL	LFPLGTINLS	LCEEERDADE	EERRDDPEER	NVEVEKR-FL	PIIAKVLGNL	LGK	
<b>temporin-1BYa</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERNADE	DERRDDPEER	NVEVEKR-FL	PIIAKVLGNL	LGK	
<b>temporin-1DRa</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDADE	EERRDDPEER	NVEVEKR-FL	PIIAKVLGNL	LGK	
<b>temporin-1DRb</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERNADE	EERRDDPEER	AVEVEKRNFL	GTLVNLAKKI	LGK	
<b>temporin-1SR</b>	MFTMKKSLLL	LFPLGTINLS	LCEEERDADE	EERRDDPEER	NVEVEKR-FL	PIIAKVLGNL	LGK	
<b>temporin-2SR</b>	---LKKSLLL	LFPLGTINLS	LCEEERNA-E	EERRDDPEEI	NVEVEKR-FP	ELSEDALASL	LGK	
<b>temporin-3C</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDADQ	EERRDDPGER	NVEVEKR-FF	PFFGKILSPA	FGK	
<b>temporin-3PR</b>	MFTLKKSLLL	LFPLGTINLS	LCEEERDADQ	EERRDDPGER	NVEVEKR-FF	PFFGKILSPA	FGK	
<b>odorrainin-M-Pa</b>	MFTLKKLLLL	LFPLGVAFSS	PCYRKREADE	EGNDGEAKTE	GIKRATAWGP	RHGLLPIRPI	RIRPLCGNDK	S
<b>odorrainin-M-Pb</b>	MFTLKKLLLL	LFPLGVAFSS	PCYRKREADE	EGNDGEAKTE	GIKRGIPWRP	PHGLKPRPPT	R-KLFCGKDK	S
<b>odorrainin-M-Pc</b>	MFTLKKLLLL	LFPLGVAFSP	PCYRKREADE	EGNDGEAKTE	GIKRGLPWRP	HHVLKPLPSP	RMKPFCKDK	S
<b>odorrainin-M-PR</b>	MFTLKKLLLL	LFPLGVAFSS	PCYRKREADE	EGNDGEAKTE	GIKRGIPWRP	PHGLKPRPPT	R-KLFCGKDK	S
<b>odorrainin-M-DR</b>	MFTLKKLLLL	LFPLGVAFSP	PCYRKREADE	EGNDGEAKTE	GIKRGLPWRP	HHVLKPLPSP	RMKPFCKDK	S

Fig. S3. Predicted antimicrobial peptide precursor sequences, listed by name of the predicted mature peptide and aligned within each mature peptide group. Species for each antimicrobial peptide precursor is indicated by capital letters at the end of each mature peptide name: C = *Lithobates clamitans*, P = *L. pipiens*, PR = *Pseudacris regilla*, BY = *Rana boylii*, DR = *R. draytonii*, SR = *R. sierrae*. Putative cleavage site for release of mature peptide (KR) is shaded. Partial coding sequences that lack the amino terminus were identified for 3 predicted antimicrobial peptide precursors: ranalexin-1C, temporin-1C, and temporin-2SR

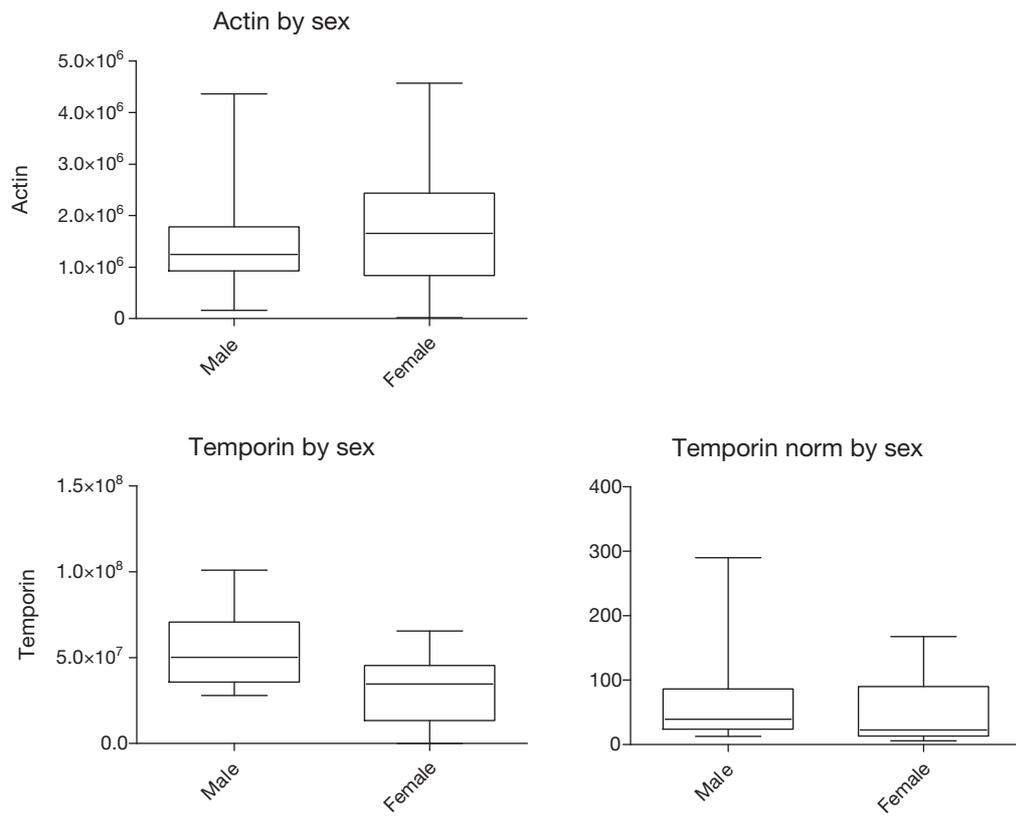


Fig. S4. *Rana boylei*. mRNA abundance of actin, mRNA abundance of antimicrobial peptide precursor temporin-1BYa, and mRNA abundance of antimicrobial peptide precursor temporin-1BYa normalized to actin in frog skin secretions by sex. Box plots show lower quartile, median, and upper quartile; whiskers indicate minimum and maximum values